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Título: Theoretical Screening of Therapeutic Peptides with Potential Anticancer Activity

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Autora

Romero Herdoiza Maylin Fernanda

Tutora

Ph.D Rodríguez Hortensia

Co-tutor

Ph.D Marrero-Ponce Yovani

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0704627900

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*This work is completely dedicated to my parents:
Juan Romero and Yolanda Herdoiza*

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Maylin Romero

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Resumen

La inespecificidad de los fármacos quimioterapéuticos y la resistencia a múltiples fármacos (MDR) adquirida por las células cancerígenas generan la necesidad de encontrar alternativas para tratar el cáncer. Los fármacos basados en péptidos son enfoques prometedores en el tratamiento del cáncer, ya que presentan valiosas ventajas como bajo peso molecular, alta especificidad y baja toxicidad. En particular, los péptidos localizadores de tumores (THP) destacan por la capacidad de unirse específicamente a los receptores de las células cancerígenas y a la vasculatura tumoral. Por otro lado, el descubrimiento de fármacos *in silico* ha demostrado ser una forma eficaz y rápida para predecir agentes quimioterapéuticos. Actualmente, hay dos predictores de THP disponibles, TumorHPD y THPep, basados en aprendizaje automático (ML) supervisado. Aquí, se desarrolla una metodología alternativa para descubrir THPs utilizando ciencia de redes y búsqueda por similitud en starPep toolbox (<http://mobiosd-hub.com/starpep/>). Este enfoque se beneficia de la Red de Espacio Químico (CSN). Se diseñaron algunos modelos basados en THPs representativos y no redundantes de la CSN para descubrir nuevos THPs a través de la búsqueda por similitud y fusión de grupos. Su rendimiento se validó con tres conjuntos de datos de referencia de THPs/no-THPs. Se alcanzaron precisiones entre 92.64-99.18% y coeficientes de correlación de Matthews entre 0.894-0.98, superando a los clasificadores de ML. Estos resultados demuestran el potencial de la búsqueda por similitud y la ciencia de redes para la predicción de actividad. Además, el mejor modelo se utilizó para reutilizar péptidos de starPepDB. Se sometieron a una optimización multiobjetivo para mejorar su farmacocinética. Por último, se propone una pequeña biblioteca de péptidos, que consta de 27 THP y 14 péptidos localizadores de tumores anticancerígenos (ACP) putativos. Estos 41 péptidos no han sido relacionados con estas actividades hasta ahora. Por lo tanto, son agentes terapéuticos prometedores para una futura validación experimental.

Palabras clave: cáncer, péptido localizador de tumores, péptido anticancerígeno, descubrimiento de fármacos *in silico*, ciencia de redes, búsqueda por similitud, red de espacio químico, fusión de grupos.

Abstract

Unspecificity of chemotherapeutic drugs and multi-drug resistance (MDR) acquired by cancer cells generate the necessity to find alternatives to treat cancer. Peptide-based drugs are promising approaches in cancer treatments since they present valuable benefits as low molecular weight, high specificity, and low toxicity. Particularly, tumor homing peptides (THPs) are highlighted by their ability to specifically bind towards receptors from cancer cells and tumor vasculature. On the other hand, *in silico* drug discovery has demonstrated being an effective and rapid way to predict chemotherapeutic agents. Currently, there are two available THP predictors, TumorHPD and THPep, based on supervised Machine Learning (ML). Herein, an alternative methodology to discover THPs is developed using network science and similarity searching in starPep toolbox (<http://mobiosd-hub.com/starpep/>). The approach benefits from Chemical Space Network (CSN). Some models were designed based on representative and non-redundant THPs from the CSN to discover novel THPs through similarity searching and group fusion. Their performance was validated with three benchmarking datasets of THPs/non-THPs. Accuracies between 92.64-99.18% and Matthews correlation coefficients between 0.894-0.98 were achieved, outperforming ML classifiers. These results demonstrate the potential of similarity searching and network science for activity prediction. Moreover, the best model was used to repurpose peptides from starPepDB. They were subjected to multi-objective optimization to enhance their pharmacokinetic. Finally, a small peptide library is proposed, consisting of 27 putative THPs and 14 putative tumor homing anticancer peptides (ACPs). These 41 peptides are not related with these activities up to now. Thus, they are promising therapeutic agents for future experimental validation.

Keywords: cancer, tumor homing peptide, anticancer peptide, *in silico* drug discovery, network science, similarity searching, Chemical Space Network, group fusion.

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Chapter 1

Introduction

Cancer is the second leading cause of death worldwide [1]. It is treated with radiotherapy, surgery, or systematic therapy [2]. Nevertheless, they carry short- and long-term health effects [2]. Notably, side effects of chemotherapy are caused by their unspecificity towards cancer cells [3]. For that reason, the attempts of the scientific community to find alternatives to currently used drugs do not cease. On this basis, peptides emerge as potential therapeutic agents for cancer treatment.

Peptides are characterized by having a low molecular weight in comparison to proteins and antibodies, short half-life time in the organism, binding to cancer cells selectively, and being non-toxic [4–7]. Hence, peptide-based drugs are opening a new door to an improved cancer diagnosis and treatment. Especially, tumor homing peptides (THPs) are highlighted by their capability to specifically bind to tumor cells and tumor vessels' receptors [8]. Therefore, they can be conjugated to therapeutic agents that present restrictions for their alone application in cancer therapy, acting as drug carriers.

THPs are discovered using *in vitro* and *ex vivo/in vivo* phage display technology [9]. However, wet-lab drug discovery procedures take much time, high investment, and need collaborative work from mastery in different fields [10]. Thus, prior *in silico* studies are employed for drug discovery, reducing resources and time [11]. In this way, short sets of molecules become the candidates for posterior experimental verification. Databases, web servers, and software, mainly based on Machine Learning (ML) approaches, are the bioinformatic tools applied to discover novel drugs [12]. Particularly about THPs, two databases recollect information about the experimentally proved peptides with tumor homing activity: TumorHoPe [13], and starPepDB [14], and two THPs web servers for prediction: TumorHPD [9], and THPep [15], both based on supervised ML approaches. Moreover, TumorHPD, and THPep design new THPs by the generation of random libraries where the peptide undergoes stochastic substitutions in different positions.

1.1 Scope of Research

This work seeks to broaden the chemical space of therapeutics peptides used for chemotherapeutic drug delivery, which contributes to solving one of the major problems found in cancer treatments: side-effects by unspecific targeting.

This study focuses on a set of potential THPs with anticancer activity found by an alternative methodology that combines network science and similarity searching. The main tools used are starPep toolbox software to perform the network analysis, scaffold extractions, and similarity searching; Chemical Space Network (CSN) to represent the chemical space of THPs as a coordinate-free system in starPep toolbox; freely available web servers for activity prediction and design of peptides; and, the evolutionary algorithm ROSE to generate peptide libraries.

1.2 Objectives

1.2.1 Principal Objective

The main objective is to discover potential THPs with a potential anticancer activity using an alternative methodology based on network science and similarity searching.

1.2.2 Specific Objectives

- To design a representative THPs model from starPepDB, which contains experimentally tested peptides.
- To carry out a similarity searching using the THPs model in the starPep toolbox to identify potential THPs from starPepDB.
- To discover new motifs in the set of potential THPs.
- To perform a multi-objective optimization of tumor homing, cell penetrability, anticancer capabilities, and half-time of potential THPs by punctual mutations and shortening sequence using webservers.
- To design THPs with anticancer activity using ROSE, an evolutionary algorithm.

Chapter 2

Background Information

2.1 Peptides as Therapeutic Agents

Peptides are short chains of amino acids joined together by peptide bonds, a covalent amide linkage (Figure 2.1). The peptide sequences are read from amino- to carboxyl-terminus [16]. Peptides have different biological roles in the organism, acting as biological regulators, inhibitors, neurotransmitters, antibiotics, hormones, ion channel ligands, or enzyme substrates [17, 18]. They can be synthesized, obtained from natural sources, or through genetic, recombinant, or chemical libraries [19, 20]. The worldwide methodology applied for peptide synthesis is the solid-phase synthesis, discovered by Merrifield in 1963 [21].

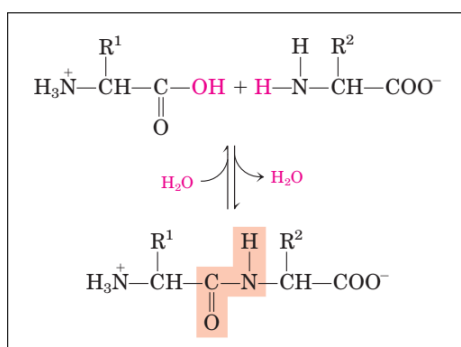


Figure 2.1: Peptide formation by an amide bond. Taken from [16].

Peptides have different biochemical and therapeutic characteristics than small molecules and proteins, making them attractive to the pharmaceutical and biotechnological industry to act as antimicrobials, antivirals, anticancer, cardiovascular agents, and treat diabetes, even vaccines [22, 23]. From 2015 to 2019, 15 peptides or peptide-containing molecules were approved by the U.S. Food and Drug Administration (FDA) as drugs demonstrating the growing interest of the scientific community [24].

Being smaller than proteins allows them to penetrate tissues more easily, have low cost, easier synthesis, and do not require folding to be biologically active [25]. In contrast to small molecules, they have higher specificity and efficacy due to representing the smallest functional part of a protein [19]. Moreover, they are not supposed to interact with the

immune system, are biocompatible, have tunable bioactivity, and have low cytotoxicity due to the degradation products being amino acids [4, 17, 25].

The low oral bioavailability and rapid metabolism are significant challenges that peptides must face up to be potential drug candidates [22]. The main reasons are that, in general, their hydrophobicity prevents crossing physiological barriers, and proteases can quickly degrade them in the blood and digestive system [5, 19]. Then, peptides have low stability and short half-life time being removed from the circulation by the kidneys and liver in minutes. Consequently, commercially available therapeutic peptides are administered via subcutaneous, intravenous, or intramuscular injections [26]. Figure 2.2 summarizes the advantages and pitfalls of using therapeutic peptides.

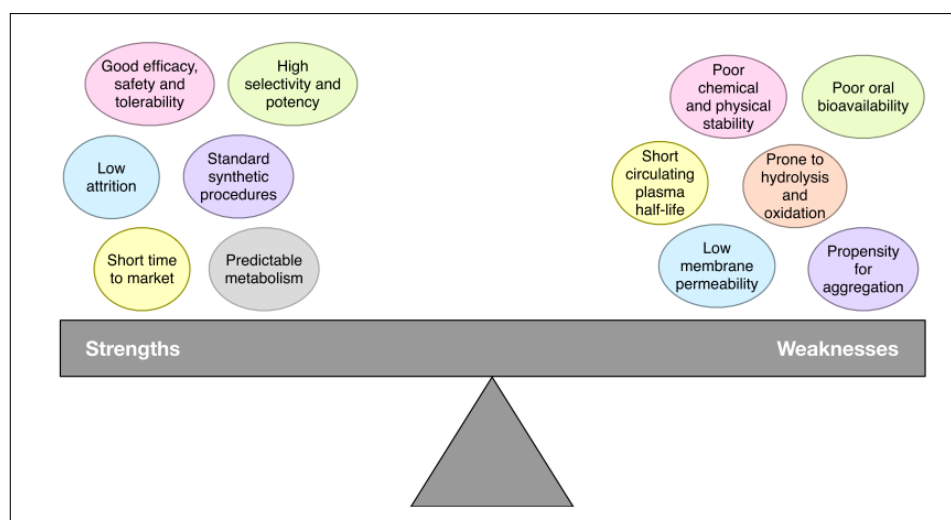


Figure 2.2: Summary of strengths and weaknesses of the application of peptides as therapeutic agents. Taken from [27].

However, studies reveal that some chemical modification and residue mutation improve their stability in plasma [28–30]. Indeed, it is reported that cysteine residues increase half-life by disulfide bond formation of the peptide with plasma albumin [31, 32]. The most common attempts to increase half-life are increasing molecular weight, cyclization, terminal modifications such as PEGylation, or replacing both terminals L-amino acids with D-amino acids [33–35].

2.1.1 Peptides in Cancer Therapy

Cancer is a disease that can be developed in different cell and tissue types. According to the World Health Organization, it is the second leading cause of death worldwide,

with approximately 9.6 million deaths (one of six deaths) in 2018 [1]. It is based on the abnormal growth of cells due to an inherited genetic mutation or induced by the environment [3]. Cells are considered cancerous if acquired the following capabilities [36]:

- Generation of their signals and to respond to weak ones that are not identified by normal cells.
- Does not respond towards antiproliferative signals.
- Resistance towards apoptotic signals.
- Replication without limit.
- Angiogenesis, i.e., stimulation of new blood vessel formation to feed them and growth.
- Metastasis and tissue invasion, i.e., spreading the invasion through the body after the localized invasion of tissue.

Tumor blood and lymphatic vasculature differ in biomarkers expression and morphology from normal lymphatic and blood vessels [37, 38]. These differences are known as the “vascular zip codes” [39]. Besides, cancer cells commonly present a higher negatively charged and fluid outer membrane and greater surface area than normal mammalian cells (Figure 2.3) [40]. The high negatively charged membrane is granted by the presence of negative glycoproteins, phosphatidylserines, O-glycosylated mucins, and chaperone proteins. The high fluidity is a consequence of low cholesterol levels. Indeed, as cancer progresses, its membrane fluidity increases. Moreover, cancer cells increase the microvilli, which concede a higher surface.

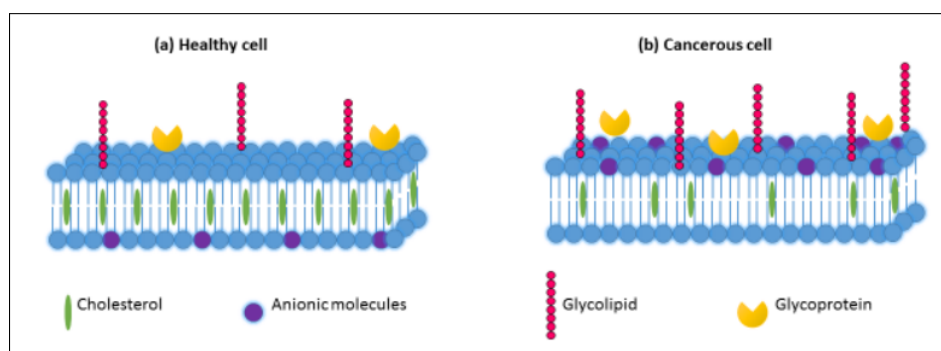


Figure 2.3: Comparison between (a) healthy and (b) cancerous cells. Taken from [40].

Localized cancers are treated with radiotherapy, by surgery, or both. However, in the case of metastatic or advanced cancer, they are treated with chemotherapy [41].

Chemotherapy is also used before local approaches to reduce the tumor size, known as neoadjuvant chemotherapy [42]. The main drawback of chemotherapy is that drugs in clinical use cannot differentiate between healthy and cancer cells, causing adverse side effects in patients [43]. Additionally, cancer cells are generating multi-drug resistance (MDR) [44]. For that reason, in the pharmaceutical industry, there is a necessity to develop new anticancer agents with a different mode of action to fight the current drug resistance of cancer cells without being cytotoxic to healthy cells [3]. Advantageously, peptides present some characteristics such as specificity towards cancer cells and low toxicity in healthy cells, allowing their application in diagnosis, treatment, and prognostic of cancer [6]. Chemotherapy based on peptides can be classified according to their mode of action.

- Mimetic peptides: to influence interactions between molecules that are relevant for cancer viability. In this way, they can induce apoptosis, immune response, tumor regression, inhibition of cancer growth, and angiogenesis [45–47].
- Biomarker peptides: to act as cancer-targeting of molecular imaging techniques in cancer diagnosis, such as magnetic resonance imaging (MRI), single-photon emission computed tomography (SPECT), or positron emission tomography (PET) [45, 48].
- Drug delivery systems: to penetrate biological barriers and/or home tumor cells or vessels to provide selective anticancer drug delivery [45, 46].

The FDA has already approved some anticancer peptides (ACPs) which are in clinical use [45]. In the years from 2015 to 2019, 5 FDA-approved drugs were destined for oncology [24].

2.1.2 Tumor Homing Peptides (THPs)

THPs are short peptides composed of 3-to-15 amino acids. They easily cross membranes by their small length and home tumor cells and vessels, taking advantage of cancer cells and tumor vessels' peculiarities [38]. THPs are widely investigated as drug carriers and for imaging purposes on oncology treatments and diagnosis since they decrease side effects [48, 49]. Moreover, nowadays, the application of nanomaterials to cancer treatment is of great interest, but they present size limitations causing low drug delivery efficiency [50]. Then, the development of peptides-conjugated nanomaterials is a promising drug delivery

system.

First-generation of THPs have RGD and NGR motifs. RGD peptides have the characteristic of selectively binding to α integrin receptors of angiogenic blood vessels, metastatic tumor cells, and tumor endothelial cells, while NGR to aminopeptidase N receptors (Figure 2.4) [51, 52].

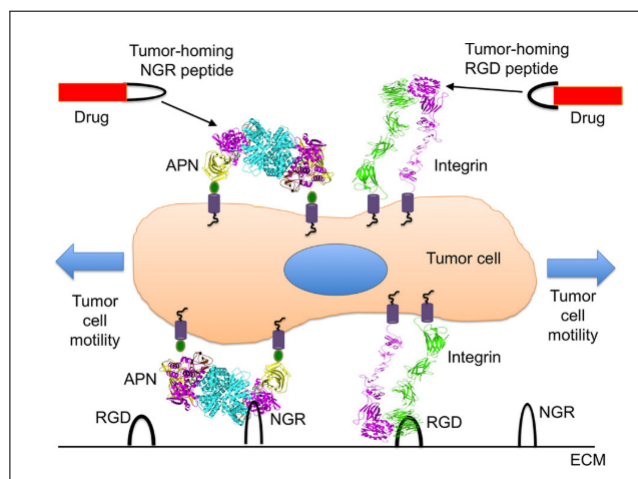


Figure 2.4: Interactions between NGR and RGD from extracellular matrix (ECM) proteins with aminopeptidase N (APN) and integrin, respectively. Taken from [53]

There are non-RGD neither NGR peptides that home tumor vasculature and cancer cells by interactions with other receptors, such as EGFR. The table 2.1 shows some of the reported motifs in THPs.

THPs and their target receptors are commonly identified through *in vitro* and *ex vivo/in vivo* phage display (Figure 2.5).

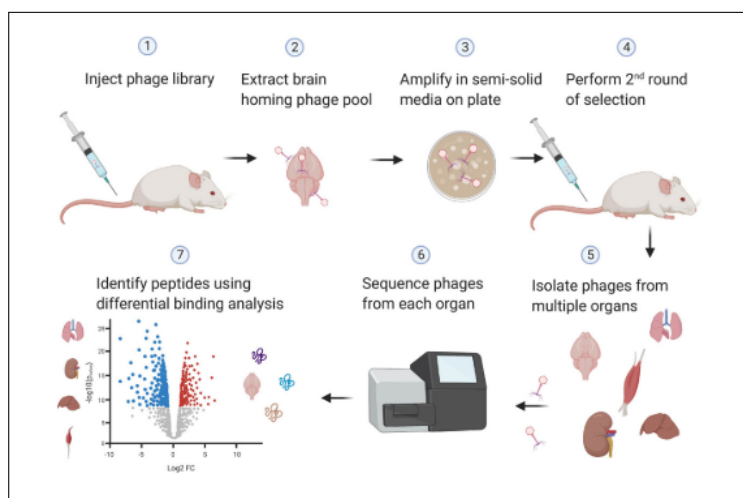


Figure 2.5: Procedure of *in vivo* phage biopanning. Taken from [54].

Phage display technology generates peptides by random peptide libraries. Random

peptide libraries are constructed based on the insertion of random oligonucleotides in the genome of phages which will encode random peptide sequences on their surfaces [55]. The general process is that phages encoding different peptides are injected into the tail of mice and let them circulate through the body for 5 to 15 minutes [51]. Then, the specific sequence binds to the target receptor and is amplified to collect the tumor-specific phages. This process is called biopanning. The main advantages of these libraries are that they can simultaneously contain up to 10^9 variants [56]. Without previous knowledge of existing interactions, it is possible to identify the sequence interacting with a target molecule [57]. However, this task is uphill, and may not translate to humans due to differences between animal model and humans, such as peptide binding and vasculature [58].

Table 2.1: Classical (well-known) tumor homing motifs. **Taken from TumorHoPe (outside parenthesis), and starPepDB (inside parenthesis).

No.	Motif	Frequency**	THP examples	Receptor	Target	Ref.
1	NGR	45/(41)	RGDPAYNGRFL CNGRCVSGCAGRC NGRSL CRGDK RGDGWK CEKRGDSVC	APN and integrin APN APN NRP-1 and integrin Integrin $\alpha_5\beta_1$ Integrins $\alpha_6\beta_3$ and $\alpha_6\beta_5$ NRP-1	Breast cancer cells (MDA-MB-435 and MCF-7) Contains RGD motif. Breast cancer (MDA-MB-435), Kaposi's sarcoma, melanoma cells. Angiogenic endothelial cells. Containing VSG motif. Neuroblastoma (GL-ME-N, GL-LI-N, HTLA-230, IMR-32, and SH-SY5Y, KS1767). Angiogenic endothelial cells. Breast cancer (MDA-MB-321, MDA-MB-231). Penetrating peptide Melanoma (B16F10). Tumor vessels. Prostate cancer. Endothelial and tumor cells.	[59] [60-66] [67] [68, 69] [70] [71]
3	RVS	7/(6)	RSGRVSN CFVSRQNK ARVFWRYSSFAPTY SKSSGVN	EGFR EGFR Gal $\beta_1 \rightarrow 3GalNAc\alpha$ disaccharide of T antigen	Ovarian cancer (SKOV3). Lung (H460), stomach adenocarcinoma cells (SNU484). Breast cancer (MDA-MB-435) cells. Breast cancer (MDA-MB-435).	[72] [73] [74] [75]
4	GVS	7/(7)	ATLDGVS	EGFR	Ovarian cancer (SKOV3).	[72]
5	AEGEF	7/(7)	AEGEFIHNRNRRFFYYWYGDPAK AEGEFMYWGDHSHWLQYWYEGDPAK AEGEFWGDHSHWLQYWYEGDPAK	HER2 (Human epidermal growth factor receptor 2)	Breast cancer (SKBr3, MCF7), ovarian cancer, melanoma. MeWo cells.	[79, 80]
6	VSG	5/(3)	RHSVSG CVSGPRC	EGFR	Ovarian cancer (SKOV3). Breast cancer.	[72] [75]
7	CSD (CSDxxHxWC)	5/(4)	CSDSWHYWC CSDYNHHWC CSDWQHPWC	VEGFR-3	Tumor metastasis.	[81]
8	WRP	5/(5)	ASSYPLHWRPWAR IHWRPWAR DRWRPALP	VEGF-C	Melanoma (B16BL6). Angiogenic endothelial cell. Tumor growth suppressor.	[82, 83]

Table 2.1 (cont.): Classical (well-known) tumor homing motifs. **Taken from TumorHoPe (outside parenthesis), and starPepDB (inside parenthesis).

No.	Motif	Frequency**	THP examples	Receptor	Target	Ref.
9	RPM	5/(14)	CPIEDRPMC GRRPMKLNKTP ALDRRPM	Integrin $\alpha_5\beta_1$	Colorectal cancer cells (HT29). Liver metastatic gastric cancer cells (XGC9811-L). Colorectal cancer cells (HT29).	[84, 85]
10	SVR	4/(4)	NSVRGSR IASVRWA CADPNSVRAMC	EGFR	Ovarian cancer cells (SKOV3). Melanoma tumor (B16B15b). Cervical cancer (SiHa).	[72] [75] [86, 87]
11	PRP	0/(6)	SVSVGMKPSRP APRPG WTHHSYPRPL	VEGF-stimulated HUVECs	Angiogenic endothelial cells. Tumor vasculature. Proved in Meth A sarcoma, and Colon 26 NL-17 carcinoma cells.	[86, 87] [82, 83]
12	GSL	0/(3)	GSLACQNIVGVKKQCNALC CLSGSLSC CGSLVRC		Breast carcinoma, Kaposi's sarcoma, and malignant melanoma.	[63]
13	KGD	0/(0)		β_3 integrin	Melanoma cells, inhibit lung metastasis	[88, 89]
14	PSP	6/(6)	SVSVGMKPSRP	VEGF- stimulated HUVECs	To the tumor neovasculature of both humans and mice. Human lung (H460), colon (HCT116), breast (BT483), prostate (PC3), liver (Mahlavu), and pancreatic (PaCa) cancer.	[90]
15	RGR	1/(1)	CRGRRST	platelet-derived growth factor receptor β (PDGFR β) expressed	Angiogenic cells from tumor vasculature. NIH-3T3.	[73, 91]

2.1.3 Cell-Penetrating Peptides (CPPs)

Cell-penetrating peptides (CPPs) are short chains of 5-to-30 residues that can internalize into cells' cytosol without cell damage [92]. Trans-activator of transcription (TAT) protein from the human immunodeficiency virus 1 (HIV-1) is the first reported CPP [93]. CPPs are rich in basic amino acids (K, R, H, and Orn) [94]. However, arginine is the one that contributes the most to enhance penetrability [95]. The majority of CPPs are cationic, but they also can be amphipathic or hydrophobic [96, 97].

CPPs are used to transport cargo to targeted cells by conjugation or co-administration. The uptake depends on physicochemical properties of peptide, cell type, cargo, interactions with membranes, concentration, temperature, and peptide to cell ratio [48, 98]. The entry mechanism is energy-dependent endocytosis, direct penetration (energy-independent), or through multiple mechanisms [99]. Endocytosis can occur by phagocytosis, macropinocytosis, caveolae/lipid raft-mediated endocytosis, or clathrin-mediated endocytosis (Figure 2.6) [100, 101]. Direct penetration occurs at higher concentrations of CPPs by electrostatic interactions with membranes and then forming inverted micelles, carpets, or pores (barrel-stave and toroidal models) [92, 96].

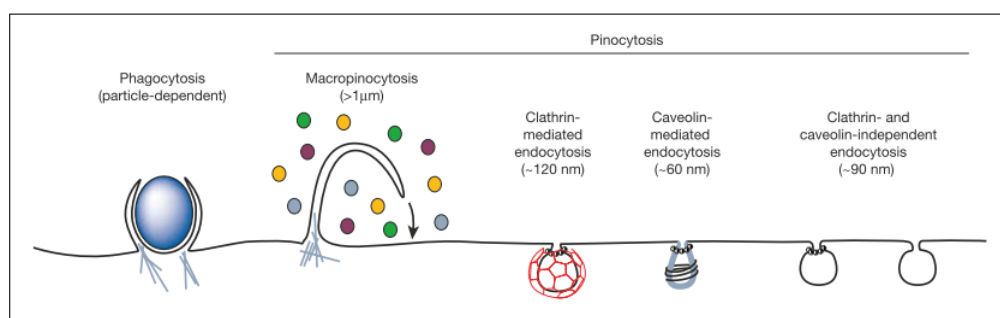


Figure 2.6: Different pathways of endocytosis. Taken from [100]

CendR (R/KXXR/K) represents an important CPP motif where X can be any amino acid different from R or K. It binds to neurophil-1 or -2 (NRP-1 and NRP-2, respectively) and initiates an endocytic transport (CendR pathway), but it is only active when it is located at the C-terminus [71]. When CendR is located into the sequence, protease can cut the sequence letting the motif in the C-terminus when the sequence is bonded to an integrin (Figure 2.7), for example, through the RGD motif, as happens in iRGD tumor-penetrating peptide [39].

The major disadvantage of CPPs is that they are not selective, then they cannot

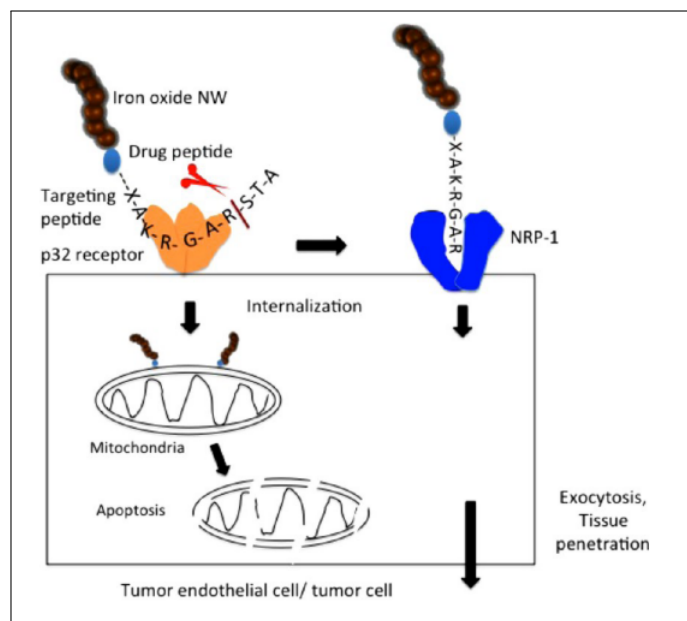


Figure 2.7: Mechanism of internalization of a peptide that contains the R/KXXR/K motif. Taken from [102].

differentiate between cancer and healthy cells [103]. Therefore, systems that penetrate tumor tissue and deliver at tumor-specific sites are desired in chemotherapy [104–107]. In this context, tumor-penetrating peptides have enhanced drug delivery of coupled and non-coupled drugs [108].

2.2 Computational-Aided Drug Discovery

Computer-aided drug discovery is a useful strategy to save time and resources, contributing to the fast introduction of peptides-based drugs in the global market [109]. In this work, some ML-based bioinformatic tools and chemical space networks are employed.

2.2.1 Machine Learning (ML)

ML, a subgroup of artificial intelligence, is the main approach applied for *in silico* drug discovery. ML uses algorithms to build mathematical models from training data sets to perform automated predictions of test sets [110].

Training data sets correspond to unlabeled and labeled data used as the sample sets. Meanwhile, test sets are unknown data sets that are going to be analyzed. The nature of the training data determines the ML model type [110], which can be: supervised learning, unsupervised learning, semi-supervised learning, reinforcement learning, or transfer

learning.

In supervised ML models, training data are labeled, while unsupervised models are unlabeled. Then, unsupervised ML models must find patterns to predict the outcomes. Semi-supervised ML combines parts of supervised and unsupervised models; its training data set contains labeled and unlabeled data, but the amount of unlabeled data is bigger. In reinforcement learning, the training data are used as feedback. Finally, transfer learning techniques consider that data is constantly changing, and data are transferred from one domain to another. By the way, supervised learning is the most widely applied for therapeutic peptides predictions [109].

ML models can classify or regress the training data to classify or regress the test sets, and the model performance will depend on the quality and quantity of training data [109, 110]. Therapeutic peptides models are commonly predicted through classifiers of supervised learning, particularly Random Forest (RF) and Support Vector Machine (SVM) [111].

RF applies classification or regression algorithms and is based on decision trees [112]. SVM classifies unlabeled data. It performs a binary classification using a linear hyperplane to maximize the separation between classes [7, 112]. When the space is not linear, SVM uses a kernel function to construct a linearly separating feature space, such as radial, polynomial, or Gaussian function [7].

2.2.2 Chemical Similarity Networks

The representation of all synthetically and natural molecules is known as chemical space. Nevertheless, as the amount of molecules in the chemical space is vast, small segments of the chemical space are used according to the activity of interest, namely, the biologically relevant chemical space where compounds that participate in biological systems are represented [113].

Chemical space is commonly visualized as a multi-dimensional coordinate system, where numerical features or computational vectors characterize molecules to represent their physicochemical properties, known as molecular descriptors [113]. Each molecular descriptor represents one dimension; thus coordinate-based maps require dimensionality reduction for visualizing in 2- or 3-Dimension maps [114]. This pitfall is known as a curse of dimensionality [115]. In this scenario, coordinate-free chemical spaces, such

as Chemical Space Networks (CSNs), emerge to visualize the chemical space with lower complexity [116]. Figure 2.8 illustrates the differences between chemical space represented as a coordinate-free system based on similarity and a coordinate-based map.

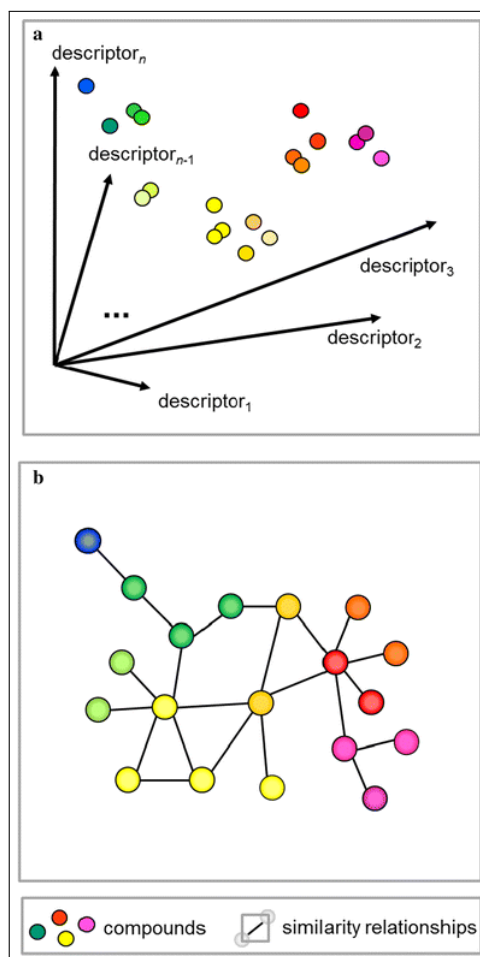


Figure 2.8: Chemical space representation as (a) a multi-dimensional coordinate-based map, and as (b) a coordinate-free similarity network. Taken from [117].

Chemical Space Network (CSN)

CSN is an undirected coordinate-free system (G) where molecules are represented as nodes joined by an edge if they have any similar relationship between their descriptors. Thus, it is defined as $G = (V, E)$, where V is the set of nodes present in network G , and E is the set of overall edges that connects V [118]. The connection between nodes depends on the selected similarity threshold. The similarity metric used in this work is the min-max normalized Euclidean, then the distance between two nodes is based on the Euclidean distance ($d(u, v)$). Two nodes are connected if $s(u, v)$ is equal or greater than the similarity threshold.

Layout algorithms determine the appearance of CSNs, where nodes are considered springs that repulse each other or are attracted by their similarity relationship. Distances between nodes are not based on how related they are but on the applied layout since its algorithm transforms pairwise similarity into the distance [119]. In this work, two algorithms for layout were used, Fruchterman Reingold and Force Atlas 2.

Additionally, some properties, and statistical measurements from networks science are applied to better understand these networks, such as clustering, modularity and centrality [120].

Similarity Threshold

Similarity threshold is an important concept in network science since it defines the network's topology and appearance [120]. It establishes the lower limit value of similarity between node pairs connected by an edge [121]. In other words, if two nodes have an equal or greater similarity value than the established, they are connected.

Node Degree

Node degree or vertex degree is the number of edges bonded to a node [122]. In other words, it represents the number of nodes with which it is attached.

Density

Network density is the ratio between the number of edges present in the network and all possible edges [120]. It depends on the similarity threshold value and determines the properties of the network. Then, network density is given by

$$\rho = \frac{2m_t}{n(n-1)} \quad (2.2.1)$$

where m_t is the number of edges at a threshold value t , and n is the number of nodes of the network. Generally, density decreases as the similarity threshold increases [114].

Clustering

Clustering is a concept with outstanding importance in unsupervised learning [114]. It is based on the division of the graph data into different communities or groups according

to the similarity between nodes [120]. Consequently, similar nodes reside in the same community, and nodes from distinct communities are different. In a network, modularity measures how good is the classification of nodes into communities [123]. Modularity can be positive or negative with a maximum value of 1 [124], and is given by

$$Q = \frac{1}{2m_t} \sum_{uv} \left(a_{uv} - \frac{k_u k_v}{2m_t} \right) \delta(c_u, c_v) \quad (2.2.2)$$

where a_{uv} is the weight of the edge, i.e., similarity value between node u and node v , k_u is the sum of the weight of edges joined to node u , c_u is the community of u , and $\delta(c_u, c_v)$ is defined as

$$\delta(c_u, c_v) = \begin{cases} 1 & \text{if } c_u = c_v \\ 0 & \text{if } c_u \neq c_v \end{cases} \quad (2.2.3)$$

Modularity represents the number of edges that connect nodes intra-community minus the expected number of edges that are randomly settled in an identical network [124]. Modularity increases as density decreases, then community structures are better resolved. Hence, modularity must be optimized as much as possible to obtain the best partition of the network.

Louvain Clustering algorithm has demonstrated the best accuracy and computing time belong reported algorithms applied for modularity optimization [114, 125]. This algorithm begins assigning all nodes in different communities and consists of two phases [125]. In phase I, one node is moved to the community of its neighbor, and its new modularity value is evaluated. When the movement increases modularity, the node is changed to this community; otherwise, it keeps in the original community. This process is performed with the full nodes until no modularity improvement occurs. In phase II, a new network is constructed based on the resultant communities from the first phase. Finally, the process (phase I and II) is repeated until no modularity changes occur.

Moreover, the network ability to cluster together can be measured through the average clustering coefficient (ACC). The clustering coefficient is the ability to connect two nodes that share a neighbor [120]. Therefore, ACC is a global measurement of the neighborhood connectivity.

Centrality

In network science, centrality is one of the essential measurements since nodes rank according to how representative they are in the network [122, 126]. There are different methods to calculate centrality, but this research is focused on harmonic, community hub-bridge, betweenness, and weighted degree.

- Betweenness centrality: is based on short path lengths. The betweenness centrality of node u is the number of shortest paths between node pairs (without considering node u) that pass through it [127]. It is given by

$$C_B(u) = \frac{1}{(N-1)(N-2)} \sum_{x \neq u, x \neq v, v \neq u} \frac{SP_{xv}(u)}{SP_{xv}} \quad (2.2.4)$$

where N is the number of total nodes, $(N-1)(N-2)$ is the number of node pairs excluding node u , $SP_{xv}(u)$ is the number of shortest paths between nodes x and v that cross node u , and SP_{xv} is the total number of shortest paths between nodes x and v .

- Harmonic centrality: is a global centrality measurement based on the distance between two nodes [128]. The harmonic centrality of node u is given by

$$C_H(u) = \sum_{v \neq u} \frac{1}{d(u, v)} \quad (2.2.5)$$

where $d(u, v)$ is the distance from node u to node v .

- Weighted degree: is based on the similarity between a node pair, known as the weight of the edge [114]. It is given by the internal and external strength as follows.

$$k_u^{in} = \sum_{v \in c_u} a_{uv} k_u^{ex} = \sum_{v \notin c_u} a_{uv} \quad (2.2.6)$$

where k_u^{in} is the internal strength, k_u^{ex} is the external strength, and a_{uv} is the similarity value between u and v .

- Community hub-bridge centrality: is a local centrality measurement based on where the node is located into the community, and nodes can be considered hubs or bridges [114]. Local hub nodes are those that connect various internal nodes, while bridge nodes are those located at the boundary of a community being the attachment

between two neighboring communities [129]. Hub-bridge centrality of node u is given by

$$C_{HB}(u) = k_u^{in} * CS(u) + k_u^{ex} * NC(u) \quad (2.2.7)$$

where k_u^{in} , and k_u^{ex} are internal and external strength, respectively, defined as in Weighted degree. $CS(u)$ is the community size of u , and $NC(u)$ is the number of neighboring communities directly attached to u by other nodes from its community.

2.3 Bioinformatic Tools

For the development of this study, some databases, web servers, and software are of particular interest.

2.3.1 Databases

TumorHoPe is a database intended to recollect reported THPs and contains 744 experimentally proved THPs [13]. Furthermore, starPepDB is a graph-based database that contains 45210 reported peptides where 659 corresponds to THPs [14].

2.3.2 Web Servers

To date, TumorHPD [9], and THPep [15] are the only web servers based on ML approaches for predicting of tumor homing activity of peptides.

TumorHPD developed by Sharma et al., was the pioneer web server [9]. Its prediction model is a SVM that uses three input features: amino acid composition, dipeptide composition, and binary profile pattern. The reported accuracy of their predictions is 86.56% [9]. However, the data set used for training and testing contains peptides with high similarity sequences and does not present statistical representations [15]. Additionally, the performance of the SVM model is not well described [15].

Alternatively, Shoombuatong et al. construct THPep where RF is used as prediction model, and amino acid composition, dipeptide composition, and pseudo amino acid composition as features achieving 90.13% of overall accuracy [15]. They removed the sequences with higher than 90% similarity to avoid overestimated predictions.

Moreover, other webservers were used to predict other activities, including cell-penetrating, anticancer, hemolysis, toxicity, antibacterial, among others (see Table 2.2).

2.3.3 Software for Network Visualization and Analysis

StarPep toolbox

StarPep toolbox is a software that uses FASTA files as inputs, and includes the starPepDB. Peptides are represented as nodes joined by an edge if they have any relationship. It can perform querying, filtering, visualization of networks, scaffold extractions, single or multiple queries similarity searching, and analysis of peptides by graph networks [14].

Networks can be built based on the metadata of peptides or based on the similarity between them. In metadata networks, nodes are connected by a specific parameter in common, such as origin, the target which assessed against, functionality, the database where they come from, the cross-reference, N-terminus, C-terminus, or amino acid composition. In similarity networks, peptides are defined by descriptors, such as length, net charge, isoelectric point, molecular weight, Boman index, indexes based on aggregation operators, hydrophobic moment, average hydrophilicity, hydrophobic periodicity, aliphatic index, and instability index. Moreover, networks are visualized using different layouts, such as Fruchterman Reingold or Force Atlas 2.

Networks can be clustered, and communities are optimized using the Louvain method. Moreover, centrality of each node can be measured, particularly, harmonic, community hub-bridge, betweenness, and weighted degree. Centrality is highly important to perform scaffold extractions due to peptides are ranked according to their centrality score, and then redundant sequences are removed, prioritizing the most central. Thus, scaffold extractions depend on the type of centrality applied.

On the other hand, similarity searching, which is the basis of this study, is performed using a set of queries against a target dataset, where different percentage of identity can be applied. The identity score is a number between 0-1, and it is calculated using Smith-Waterman local alignment and Blosum 62 substitution matrix. Multiple queries similarity searching works using the group fusion model.

Gephi

Gephi is open-source software for the visualization and analysis of network graphs. It calculates relevant data from the networks, such as average degree, diameter, radius, density, modularity, clustering coefficient, average clustering coefficient (ACC), average

path length, number of edges, and nodes.

Table 2.2: Webservers used for activity predictions of peptides. Algorithms for classification are Support Vector Machine (SVM), Random Forest (RF), eXtreme Gradient Boosting (SGBoost), Artificial Neural Network (ANN), WEKA (package of classifier algorithms), Determinant Analysis (DA), and (Meta)genomic AMP Classification and Retrieval system (MACREL).

No.	Webserver	Predicted activity	Classifier	Ref.
1	TumorHPD	Tumor homing	SVM	[9]
2	THPep	Tumor homing	SVM	[15]
3	AntiCP	Anticancer	SVM	[130]
4	ACPred	Anticancer	SVM and RF	[131]
5	iACP	Anticancer	SVM	[132]
6	ENNAACT	Anticancer	ANN	[133]
7	CellPPD	Cell penetrating	SVM	[134]
8	C2Pred	Cell penetrating	SVM	[135]
9	MLACP	Cell penetrating	RF	[136]
10	CpACpP	Anticancer and cell penetrating	XGBoost, SVM and RF	[137]
11	ToxinPred	Toxic	SVM	[138]
12	HemoPI	Hemolytic	SVM	[139]
13	HemoPred	Hemolytic	RF	[140]
14	PlifePred	Half-life in blood	SVM and WEKA	[28]
15	HLP	Half-life in acid media	SVM	[141]
16	ANuPP	Amyloidogenic	Regression	[142]
17	SGnn	Prion-like domains	ANN	[143]
18	AlgPred2	Allergens	RF	[144]
19	PepSolubility 1.0	Solubility	-	-
20	SolupHred	pH-dependent aggregation	-	[145]
21	IL2Pred	IL-2 induction	RF	[146]
22	IL4pred	IL-4 induction	SVM	[147]
23	IL-10Pred	IL-10 induction	SVM and RF	[148]
24	AIPpred	Inflammatory	RF	[136]
25	ProInflam	Inflammatory	SVM and RF	[149]
26	AntiInflam	Inflammatory	SVM	[150]
27	PRRpred	Pattern recognition receptor	SVM	[151]
28	QSPpred	Quorum sensitivity	SVM	[152]
29	AMPfun	Various (anticancer, antimicrobial, etc)	SVM and RF	[153]
30	CAMPPr3	Antimicrobial	SVM, RF, ANN and DA	[154]
31	AxPEP	Antimicrobial	RF	[155]
32	Macrel	Antimicrobial and hemolytic	MACREL	[156]
33	AMPDiscover	Various (antimicrobial, antiviral, etc)	RF and ANN	[157]
34	ClassAMP	Antibacterial, antiviral and antifungal	SVM	[112]
35	iAMPpred	Antibacterial, antiviral and antifungal	SVM	[158]
36	Antifp	Antifungal	SVM	[159]
37	Meta-iAVP	Antiviral	RF	[160]
38	AntiTbPred	Antitubercular	SVM	[161]
39	dPABBs	Bio-film active	SVM and WEKA	[162]

Chapter 3

Experimental Procedure

The overall workflow of this study, shown in Figure 3.1, is based on 3 steps: i) selection of the model of representative THPs from starPepDB, ii) prediction of potential THPs, and iii) multi-objective optimization of potential THPs. In the first step, some models of representative THPs from starPepDB were built using different centrality measures to rank the nodes and extract the representative and less redundant sequences by local alignment; then, the best model was selected in accordance with the performance and its capacity to correctly retrieve THPs from well-known THPs databases using similarity searching and group fusion. In the second step, the model was used to perform similarity searching with the aim to repurpose peptides as THPs from starPepDB, and their tumor homing activity was optimized using TumorHPD server. Additionally, sequence motifs were found from the set of potential THPs using multiple sequence alignments, alignment-free methods, and PROSITE server. In the last step, cell-penetrability, anticancer, and stability of potential THPs were optimized by three methodologies: punctual mutations and shortening the sequences in freely available webservers, creating a family of related peptides from a root peptide by applying a probabilistic model of evolution called ROSE, and by the addition of TAT sequence at the *C*-end.

3.1 Model Selection

3.1.1 Data Extraction

The dataset of reported THPs was extracted from starPepDB in starPep toolbox. All 45120 peptides contained in starPepDB were filtered by the query “Tumor Homing” in the metadata function, where 659 entries were obtained.

3.1.2 Similarity Threshold Analysis

Network analysis of peptides was performed building CSN of 659 THPs in starPep toolbox. In order to choose the appropriate similarity threshold to build the network of THPs, CSNs were built varying 0.05 the cut-off value from 0.10 to 0.90 (17 CSNs in total). Some

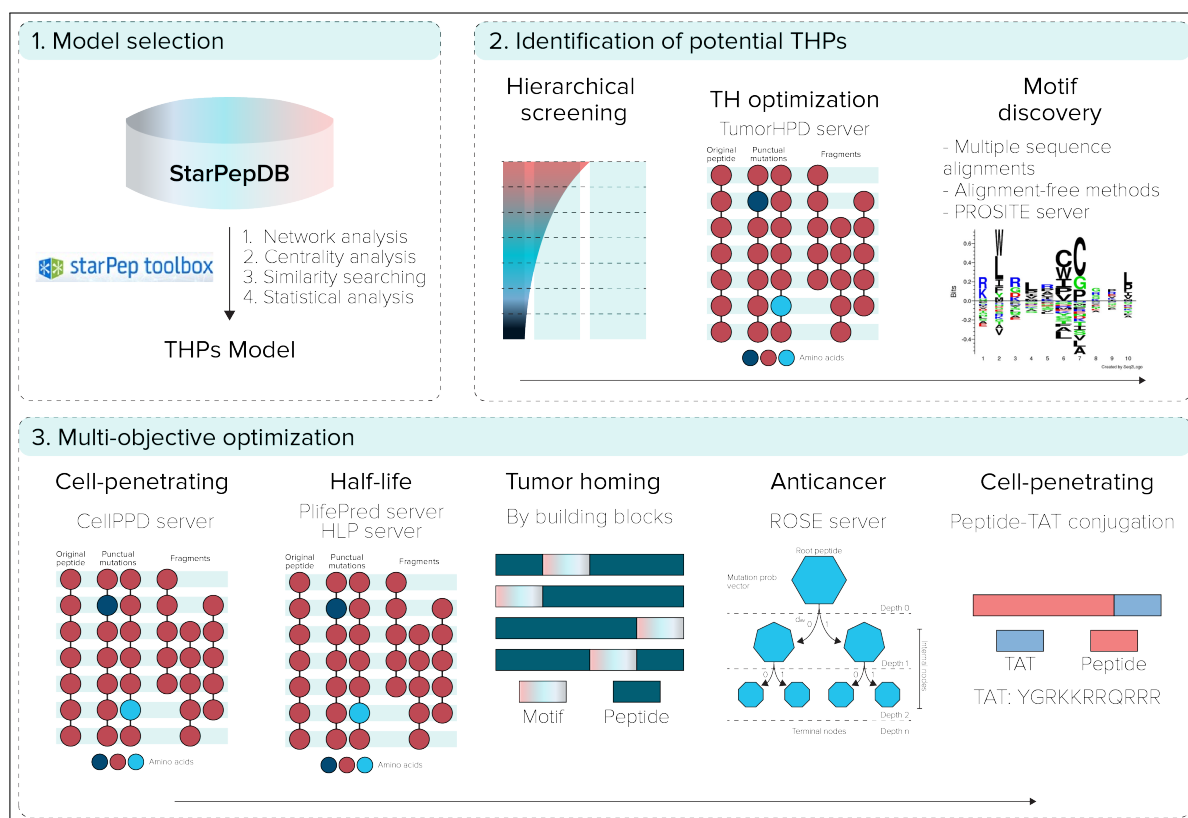


Figure 3.1: General overview of the experimental procedure.

metrics were retrieved from each CSN using starPep toolbox, such as density, number of communities, modularity, and number of singletons.

By default, when CSN is built, nodes with higher than 98% of similarity were eliminated using the local alignment Smith-Waterman algorithm and Blosum 62 substitution matrix. The similarity metric used to establish the pairwise similarity relationships between nodes was the min-max normalized Euclidean. Then, the community hub-bridge centrality was calculated with which outliers, nodes with 0 as vertex degree, were identified and removed, remaining the giant (or connected) components of the CSN, i.e., subgraph where all nodes are connected. After that, the network was clustered and the modularity optimized using the Modularity optimization clustering algorithm which is based on the Louvain method [125].

The network was saved as a Graph ML file to open in Gephi [163] for subsequent calculation of ACC. Finally, density, modularity, and ACC as a function of similarity threshold were graphed in Origin to decide which similarity threshold is better.

3.1.3 Network Characterization

CSN of the giant components using the best similarity threshold is characterized by the number of nodes, edges, outliers, density, number of communities, and modularity, which were parameters obtained from starPep toolbox; ACC, diameter (larger shortest path), average path length, and a total of triangles, which were obtained from Gephi; and the distribution degree. These parameters allow knowing the topology, and structural patterns of the CSN.

For network visualization, Force Atlas 2 was used as a layout algorithm, colors represent different clusters, and node size depends on how central is according to the community hub-bridge centrality. Network visualization aims to obtain an aesthetically pleasing and understandable graph where nodes are not overlapped.

Outliers

CSN of outliers was built with a cut-off of 0.30 to procure an appropriate density and, then, it was clustered. Moreover, a subsequent scaffold extraction was applied based on hub-bridge centrality, and 30% identity by local alignment was applied.

The network was characterized according to the number of nodes, edges, and communities, density, modularity, average degree, ACC, and diameter obtained before scaffold extraction, and the number of nodes and edges obtained after scaffold extraction. For network visualization, Fruchterman Reingold was used as a layout algorithm, colors represent different clusters, and node size depends on how central is according to hub-bridge.

3.1.4 Similarity Searching Model for THPs Prediction

In this study, the proposed method for discovering potential THPs was based on similarity searching. For that reason, multiple query similarity searching models (SSMs) composed by several queries of the most important and less redundant nodes of CSN and a similarity threshold were tested against datasets that contain well-known THPs/non-THPs through similarity searching. The recoveries from the similarity searching were statistically evaluated to choose the best model which was used to identify potential THPs.

Centrality analysis

The most influential nodes were used to find the new potential THPs, and centrality is the key parameter that provides this information. Thus, the four available centrality types in starPep toolbox, weighted degree, community hub-bridge, betweenness, and harmonic, were calculated and normalized using the min-max method. Then, redundant peptides were removed applying scaffold extraction, where peptides were ranked based on the scores obtained after centrality calculation, and using as similarity identity cutoff 30% based on local alignment algorithm Smith-Waterman and Blosum 62 substitution matrix. Subsequently, nodes with 10% lower centrality than the most central node were removed in each metric.

On the other hand, harmonic and weighted degree were calculated, normalized, and redundant peptides were removed applying 4 different percentages of similarity identity, 30, 40, 50, and 60%.

Query datasets (reference sequences)

The retrieved sets after applying scaffold extractions at each centrality measure and the two sets of outliers were used as queries. Additionally, combinations of outliers with sets obtained from centrality-based scaffold extractions, and combinations between sets obtained from scaffold extractions performed using different centrality metrics, were used as queries. In total, 22 sets of most influential nodes were used as queries, where 12 sets came from each applied percentage of scaffold extraction, 2 sets of outliers, and 8 sets came from the combination between sets.

Target Databases

Three training datasets that consider well-known THPs and randomly generated non-THPs [27] were used as the target or calibration for the recovery. THPep and TumorHPD employ these datasets for training their supervised ML classifiers [9, 27].

- Main dataset: 651 experimentally validated THPs and 651 random non-THPs. They were collected from TumorHoPe[13], and the literature [9].
- Small dataset: 469 experimentally validated THPs and 469 random non-THPs. They are peptides derived from the Main dataset with a length of 4-to-10 aa residues.

- Main90 dataset: 176 THPs and 443 non-THPs. They are peptides from the Main dataset with equal or lower than 90% of sequence similarity.

Main and Small datasets were retrieved from Ref. [9], while Main90 from Ref. [27].

Group fusion

Group fusion is based on the variation of a query (reference molecule), but keeping constant the identity measure [164]. The identity score is calculated to each peptide from target dataset varying the queries. The fusion group’s algorithm associates a fused score to each target peptide, i.e., the maximum similarity (MAX-SIM) score from all obtained identity scores varying the query. Therefore, considering peptide S from target dataset, reference peptide Q from queries, and the identity score $I(S,Q)$ the MAX-SIM score obtained, the algorithm assigns $I(S,Q)$ as the fused score to peptide S. The identity scores were calculated with the Smith-Waterman local alignment algorithm with Blosum62 substitution matrix, and is a number between 0-1, being 1 the maximum similarity score.

Retrospective Similarity Searching

Main Dataset was imported to starPep toolbox, and the similarity searching based on local alignment Smith-Waterman and Blosum 62 substitution matrix were performed using the “Multiple query sequences” option of the software and the sets obtained from and 30% of scaffold extraction followed by removing nodes with 10% lower centrality than the most central node as queries. During the similarity searching the group fusion is applied by default, and results were ranked according to the fused score corresponding to the MAX-SIM value. Subsequently, seven different percentages of identity (similarity thresholds), 30, 40, 50, 60, 70, 80, and 90%, were tested, where peptides with similarity scores equal or higher than the applied threshold were retrieved as predicted THPs. The rescued nodes, i.e., predicted THPs, were statistically evaluated to validate the prediction. The procedure is illustrated in Figure 3.2. Here, it was possible to identify the two centrality measures and percentages of sequence identity with the best performance.

Then, similarity searching was performed using only sets of the best two centrality measures as queries: harmonic and weighted degree, and 30, 40, 50, 60, and 70% of identity. In Small and Main90 datasets, only sets of harmonic and weighted degrees were used, applying 40, 50, and 60% of identity for recovery.

In total 98 different SSM were evaluated. Figure 3.2 illustrates how similarity searching works.

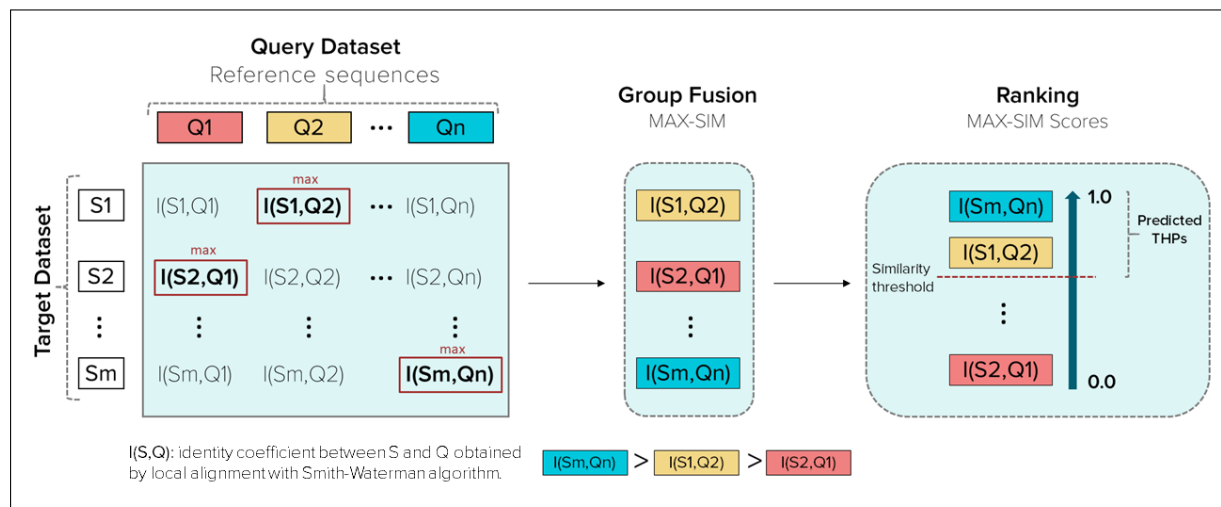


Figure 3.2: Schematic representation of the similarity searching process. Q is a peptide from a query dataset, n the number of peptides contained in a query dataset, S is a peptide from the target dataset (Main, Small or Main90 dataset), m is the number of peptides contained in the target dataset (1302, 938 or 619, respectively).

Statistical Analysis

The ability of the SSMs to predict THPs was validated by the measurement of accuracy (Ac), kappa (κ), recall (R), precision (P), Matthews correlation coefficient (MCC), and false accept rate ($FAR\%$) using the following formulas.

$$Ac = \frac{TP + TN}{TP + TN + FP + FN} \quad (3.1.1)$$

$$\kappa = \frac{Po - Pc}{1 - Pc} \quad (3.1.2)$$

$$R_{TP} = \frac{TP}{TP + FN} \quad (3.1.3)$$

$$R_{TN} = \frac{TN}{TN + FP} \quad (3.1.4)$$

$$P_{pos} = \frac{TP}{TP + FP} \quad (3.1.5)$$

$$P_{neg} = \frac{TN}{TN + FN} \quad (3.1.6)$$

$$MCC = \frac{TP * TN - FP * FN}{\sqrt{(TP + FP) * (TP + FN) * (TN + FP) * (TN + FN)}} \quad (3.1.7)$$

$$FAR\% = \frac{FP}{FP + TN} * 100 \quad (3.1.8)$$

where TP is the number of true positives, TN is the number of true negatives, FP is the number of false positives, FN is the number of false negatives, R_{TP} is the recall of true positive or sensitivity, R_{TN} is the recall of true negative or specificity, P_{pos} is the precision of positive predictions, P_{neg} is the precision of negative predictions, Po is the relative observed agreement between the observers equal to the Ac formula, and Pc is the expected chance agreement calculated by the formula $Pc = \frac{(TP+FP)*(TP+FN)+(FN+TN)*(FP+TN)}{(TP+TN+FP+FN)^2}$.

Finally, the best 9 SSMs were compared and ranked using the Friedman test-based analysis performed in KEEL [165], open-source software from Java. The Friedman test identified the best model based on the statistical metrics previously shown [166]. Moreover, it allows us to compare the models and determine if the difference between them is statistically significant and not due to chance. The confusion or classification matrix of the best model was constructed. Additionally, the best models were compared with reported ML models used for THPs prediction, TumorHPD and THPep, by using the same 3 calibration datasets.

3.2 Potential THPs Prediction

3.2.1 Hierarchical Virtual Screening

Pipeline Prospective Screening

The first step to identify potential THPs was to carry out a drug repurposing in the starPep toolbox, which is to find new target activities of known molecules [167]. Currently, this alternative methodology to discover drugs is widely applied due to reduced approval time for their clinical use [168]. In this sense, peptides from starPepDB were repurposing as THPs.

First, peptides without reported TH activity and toxicity with a sequence length between 3 to 25 residues were filtered from the chemical space of starPepDB. Secondly, peptides with higher than 95% of sequence similarity by local alignment Smith-Waterman and Blosum 62 substitution matrix were removed using the Scaffold extraction option. Thirdly, multiple query similarity searching was performed using the best SSM, obtained in the previous section, as the query against the chemical space of non-THPs, non-toxic,

and non-redundant peptides with a length of 3-25 aa (amino acids), using 60% as similarity threshold. In the recovered set, peptides with a similarity score of 1 were removed.

Activity Prediction

Peptides with reported tumor homing activity in the literature were removed since the main objective of this study is to identify novel THPs. Then, theoretical activities of virtual hits were predicted using webserver 1-3, 7, 11, and 12 from Table 2.2, to corroborate their potential as THPs and prioritize those that do not harm healthy cells. The activities of interest were tumor homing, anticancer, cell-penetrating, toxicity, and hemolysis. The SVM thresholds used were 0.30 in servers 1, 3, and 7, and 0 in server 11.

Redundancy Reduction by Network Analysis

CSN of hits was built, clustered, and the modularity was optimized using the Louvain method in starPep toolbox. Then, harmonic and weighted degree centralities were calculated to perform a scaffold extraction using a 60% identity as threshold.

Visual Mining

The neighborhood of well-known THPs of each potential THPs was visualized using starPep toolbox. CSN of 659 THPs in starPepDB was built using 0.60 as cut-off, clustered, and optimized modularity. Hits obtained in the previous step after scaffold extraction were embedded into the CSN of 659 THPs to study the neighborhood of each peptide. Hence, the 3-nearest neighbors from 659 THPs which are directly attached to each hit, were visualized. When two peptides had the same two or three neighbors, one of them was prioritized, choosing the one with better-predicted activities.

3.2.2 Tumor Homing Optimization

Lead hits obtained from hierarchical virtual screening are peptides from starPepDB with a natural or designed activity different from tumor homing. This is why their tumor homing action should be enhanced. Lead hits were optimized by punctual amino acid mutations using the “Designing of Tumor Homing Peptides” module of TumorHPD (<https://webs.iiitd.edu.in/raghava/tumorhpd/peptide.php>) (Figure 3.3). Moreover, lead

and mutated sequences were shortened into fragments 5, 10, and 15 residues in length using the same server.

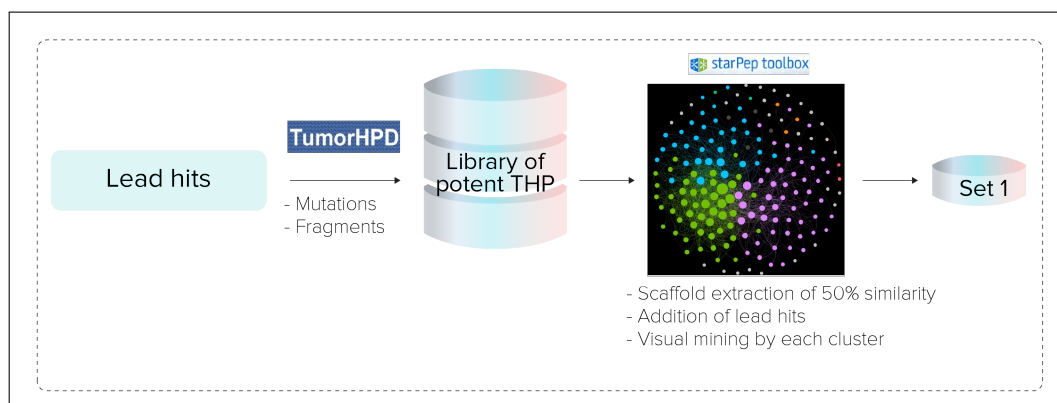


Figure 3.3: Procedure to optimize tumor homing activity of lead hits.

The selected optimized sequences, which must show a higher tumor homing activity score than parent hits, were analyzed through CSN in starPep toolbox using 0.60 as similarity threshold to build the network. Besides, tumor homing, toxicity, hemolytic, anticancer, and cell-penetrability were predicted using servers 2, 3, 7, 11, and 12 from Table 2.2. Redundant sequences with higher than 50% of similarity were removed by scaffold extraction.

Finally, the optimized sequences and parent hits were merged, and its CSN was built using 0.50 of cut-off and clustered. Moreover, harmonic centrality was calculated. Each cluster was analyzed separately in order to prioritize the most central, potent, non-toxic, and non-hemolytic potential THPs.

The heat map and histogram of pairwise sequence identity of lead compounds were constructed to study their structural diversity.

3.2.3 Discovery of THP Motifs

Multiple Sequence Alignments

The resulting potential THPs were hard-to-align sequences because of their short length and variability, they were grouped into seven clusters according to the neighborhood in the CSN. Given that clusters 1 and 5 were underrepresented by 2 peptides each, they were fused in a cluster labeled 1-5. Thus, peptide clusters (2-4, 1-5, and singletons) were aligned independently by using multiple sequence alignments (MSA), publicly available

at <https://www.ebi.ac.uk/Tools/msa/>. Four different MSA algorithms were applied with their default parameters to determine consensus motifs within each cluster.

1. Clustal-Omega v 1.2.4 [169].
2. MAFFT (Multiple Alignment using Fast Fourier Transform) v7.487 with the iterative refinement FFT-NS-i option [170].
3. MUSCLE (Multiple Sequence Comparison by Log- Expectation) v3.8 [171].
4. T-Coffee (Tree-based Consistency Objective Function for Alignment Evaluation) v1.83 [172].

The resulting MSAs were employed to extract the conserved motifs by considering the consensus sequences estimation from the programs Jalview v2.11.1.4 [173], EMBOSS Cons v6.6.0 (https://www.ebi.ac.uk/Tools/msa/emboss_cons/) and Seq2Logo v2.1 (<http://www.cbs.dtu.dk/biotools/Seq2Logo/>) [174].

Alignment-Free Method

Peptides were analyzed in STREME [175] (Sensitive, Thorough, Rapid, Enriched Motif Elicitation) to discover fixed-length patterns (ungapped motifs) that were enriched with respect to a control set generated by shuffling input peptides [173]. The analyses were performed via its webserver <https://meme-suite.org/meme/tools/streme>, by considering both total peptides and by each cluster. The motif width was set between 3-5 amino acids length. STREME applies a statistical test at p-value threshold = 0.05 to determine the enrichment of motifs in the input peptides compared to the control set.

Motif Search in PROSITE

Peptides were queried by the Motif Search tool (<https://www.genome.jp/tools/motif/>), integrated into the GenomeNet Suite (<https://www.genome.jp/>). PROSITE Pattern and PROSITE Profile libraries were only considered for the motif search.

3.3 Multi-Objective Optimization of THPs

3.3.1 Cell-Penetrating Activity

The penetration ability of 54 THPs was optimized by amino acid mutations using “Design Cell-Penetrating Peptide & Generate Its Mutants” module from CellPPD (<https://webs.iiitd.edu.in/raghava/cellppd/submission.php>). Toxicity, tumor homing, anticancer, and hemolytic activities of optimized sequences were also predicted using the servers 1-3, 7, 11, and 12 from Table 2.2. Then, 54 THPs were combined with optimized sequences, and its CSN was built in starPep using 0.65 as threshold and clustered. Harmonic centrality was calculated, followed by the scaffold extraction of sequences with lower than 90% similarity. Then, a set of sequences was selected, analyzing the neighborhood of each cluster and prioritizing optimized sequences with higher TH activity, non-toxic, and non-hemolytic. Finally, multiple reference similarity searching was performed using the THPs model. Figure 3.4 shows the overall procedure for the optimization.

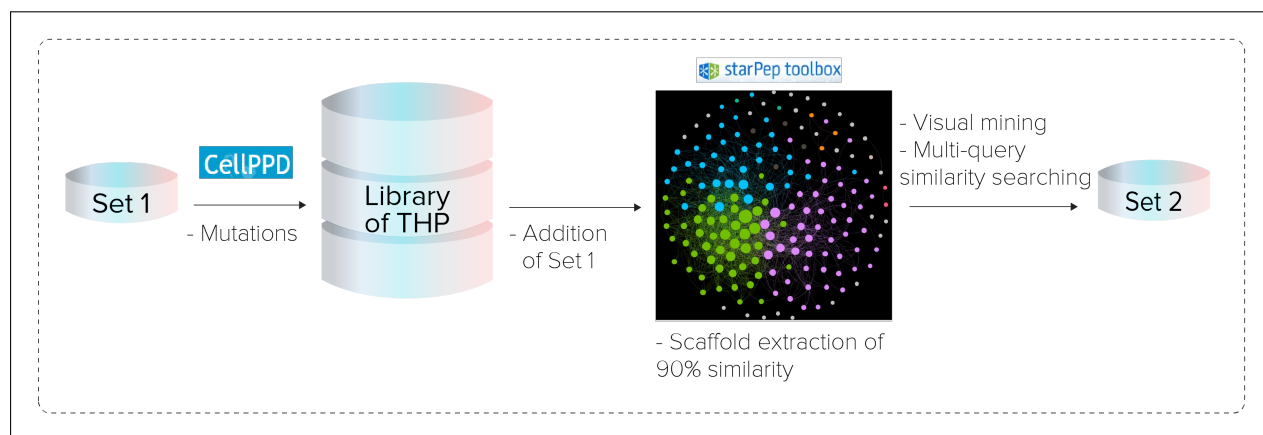


Figure 3.4: Procedure to optimize cell-penetrating activity.

3.3.2 Half-Life Time

Half-life in the blood of sequences obtained after cell-penetrating optimization was optimized by punctual amino acid mutations and shortening in fragments of 5 and 10 residues using the “Analog Generation” module from PlifePred (<https://webs.iiitd.edu.in/raghava/plifepred/analog.php>). Toxicity, tumor homing, cell-penetrating, anticancer, and hemolytic activities of optimized sequences were predicted using the servers 1-3, 7, 11, and 12 from Table 2.2. Then, multiple sequences searching was performed using the

model of THPs in starPep toolbox. On the other hand, CSN of unrecovered sequences from multiple searching was built using 0.65 as threshold and clustered. Harmonic centrality was calculated, and sequences with higher than 60% similarity by local alignment were removed. Figure 3.5 shows the overall procedure for the optimization.

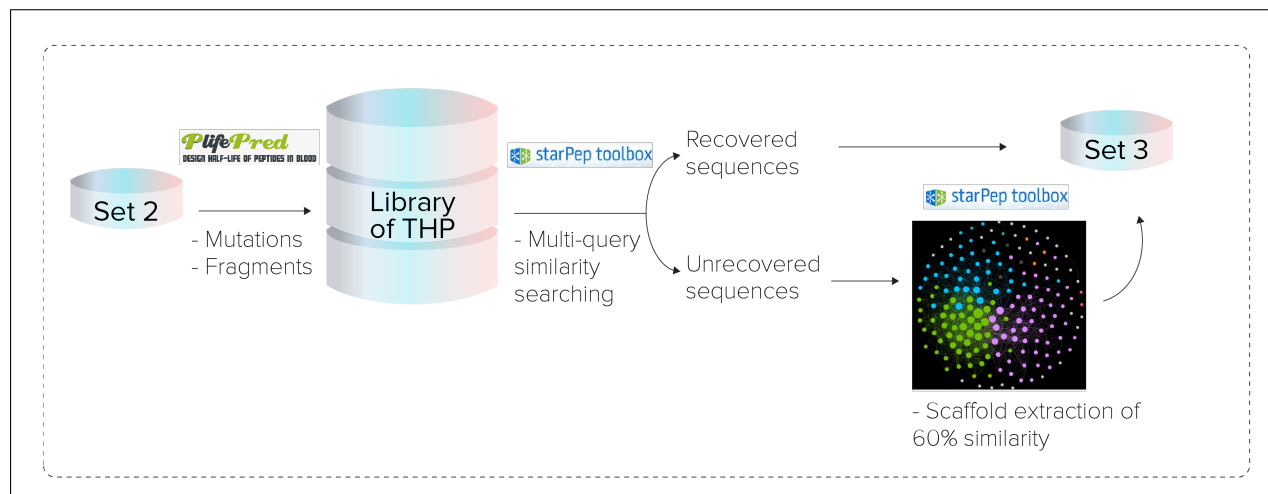


Figure 3.5: Procedure to optimize stability in blood.

Subsequently, half-life in an intestinal-like environment was optimized by punctual amino acids mutations and shortening in fragments of 5, 10 and 15 residues using the “Submission form for Designing Stable Peptide” module from HLP (http://crdd.osdd.net/raghava/hlp/pep_both.htm). Toxicity, tumor homing, cell-penetrating, anticancer, hemolytic activities, and half-time in the blood of optimized sequences were predicted using the servers 1-3, 7, 11, 12, and 14 from Table 2.2. Then, multiple sequences searching was performed with the model of THPs in starPep toolbox. In this case, the CSN of both recovered and unrecovered sequences was built using 0.65 as threshold. Finally, harmonic centrality was calculated, and sequences with higher than 90 and 65% similarity by local alignment were removed, respectively. Figure 3.6 shows the overall procedure for the optimization.

To deeply characterize the optimized THPs, more activities were predicted, such as allergen reaction using AlgPred2, aggregation-prone regions (APR) using AnuPP, and hemolysis using another server, HemoPred. In those sequences with unfavorable predicted activities, they were replaced with a better variant obtained by punctual mutations. They were filtered to keep a stronghold of the best performing multi-target sequences following the prediction specified in Table 3.1. Finally, multiple sequence searching was performed using 60% of identity with the model of THPs. Therefore, potential peptides are tumor

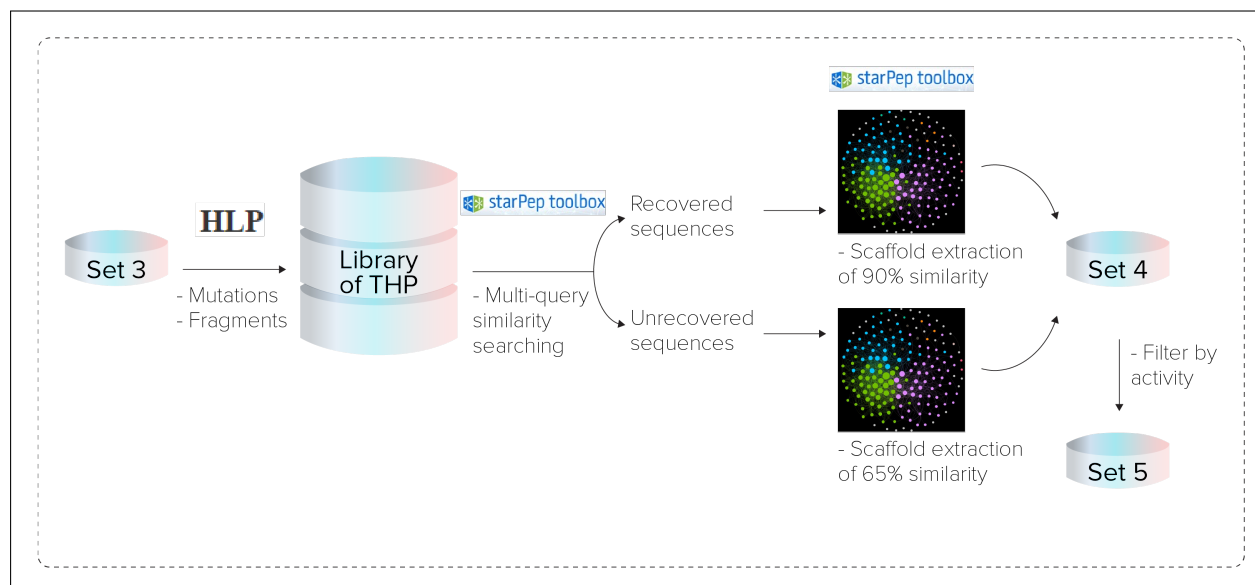


Figure 3.6: Procedure to optimize stability in the gastrointestinal tract.

homing active by all available models to predict THPs, the developed here, THPep, and TumorHPD.

Table 3.1: Parameters to be met by peptides in activity predictions.

Server	Prediction
TumorHPD	Score >2
THPep	TH
AntiCP	Score >0.60
ToxinPred	Non-toxic
HemoPred	Non-hemolytic
AlgPred	Score <0.40
ANuPP	no APR
HL in blood	>800 s

3.3.3 Building Blocks

Using three sets of motifs, new sequences were designed by building blocks.

1. Discovered motifs: Tables 4.10, 4.11, and 4.12.
2. Reported motifs: Table 2.1.
3. Short sequences from optimized hits: Table 4.13.

As two motifs found in PROSITE (Table 4.12) have 10 and 7 aa, respectively, motif 1 was divided into two sequences of 5 aa each, and only the last 4 aa of motif 2 were

considered as a motif since the first 4 aa was also found as a motif by STREME (Table 4.11). Motifs were added in *N*-terminus, *C*-terminus, and inserted into three random positions in the sequences from SET 4 and also in motif sequences. Then, their activities (tumor homing, anticancer, cell-penetrating, toxicity, hemolysis, allergen, and half-life in blood) were predicted using the servers 1-3, 7, 11, 12, 14, 16, and 18 from Table 2.2, and sequences were filtered considering the parameters shown in Table 3.1.

CSN of the obtained sequences was built and clustered in starPep toolbox. Harmonic centrality was calculated, followed by a scaffold extraction of sequences with lower than 70% similarity by local alignment. Toxic sequences were identified and removed by the multi-sequence searching using as query a set of representative 105 venom peptides (Attachments **A**) obtained from starPepDB and 50% of identity as a cut-off. Then, TH active by the developed model and ACPs were identified by two independently multiple query similarity searching. One was performed using 60% of identity and the model of THPs as query, and the second was performed using 50% of identity with a set of representative 162 ACPs (Attachments **B**) obtained from starPepDB. Both recoveries were joined. In unrecovered peptides, CSN was constructed and clustered. Then, hub-bridge centrality was calculated, followed by a scaffold extraction of sequences with lower than 50% similarity.

On the other hand, active CPPs by the two models of CellPPD from the library of building blocks were filtered. Their CSN was built and clustered in starPep toolbox, followed by the scaffold extraction of sequences with lower than 50% similarity.

3.3.4 Final Selection of Potential THPs

Sequences obtained from the activity optimization (SET 5) and building blocks were joined. CSN was built and clustered, followed by a scaffold extraction of 70% similarity based on harmonic centrality in starPep toolbox. Then, a multiple sequence search was performed using 60% of identity with the THPs model and an ACPs model found in Attachments **B**. In unrecovered peptides, CSN was constructed and clustered. Then, hub-bridge centrality was calculated, followed by a scaffold extraction of sequences with higher than 50% similarity.

Finally, the set of potential THPs was reduced to 27 sequences in order to provide a pool of the most potent sequences considering other predicted activities, such as solubility

(server 20 from Table 2.2). The biological profile of 27 sequences was characterized by predictions using the remaining servers from Table 2.2.

De novo Design of ACPs

An alternative optimization methodology was performed to improve tumor homing activity, penetrability, solubility but most importantly, anticancer activity while maintaining low toxicity and hemolysis. For this purpose, 14 sequences with a higher compromise between their activities were chosen and some of them were optimized using ROSE (<https://bibiserv.cebitec.uni-bielefeld.de/rose>) [176].

The ROSE program is an algorithm that creates a family of related peptides from a root peptide by applying a probabilistic model of evolution. The algorithm inserts, deletes, and substitutes amino acid residues from the sequences guided by the topology and branch lengths of a predefined evolutionary tree. ROSE was calibrated, hence the generated peptides retained at least 60% of identity with the corresponding root sequence. ROSE's internal parameters were tuned as follows: the binary mutation guide trees with 1023 nodes and depth $k = 9$, and average distance (d_{av}) of 5–20 PAMs. The diversity of the resulting peptides also depends on the root sequence, which is represented by a mutation probability vector. Each position/residue in the vector is weighted by variability or conservation degree shown in the sequence consensus, where the “zero” value indicates no mutations (high conservation degree), while the “one” value represents high mutation probability. Figure 3.7 illustrates the binary mutation guide tree used by ROSE.

Then, the well-known cell-penetrating sequence TAT (YGRKKRRQRRR) was added to the *C*-end of 14 peptides to evaluate if their anticancer activity is kept while penetrability increases. TAT was added directly and via a non-steric hindrance amino acid, A. Moreover, their 3D structures were generated using PEP-FOLD 3.0 [177], to study whether TAT affects the structural conformation of the sequence, resulting in loss of activity. Finally, the sequences were fully characterized using all of the servers listed in Table 2.2, and their activities were compared.

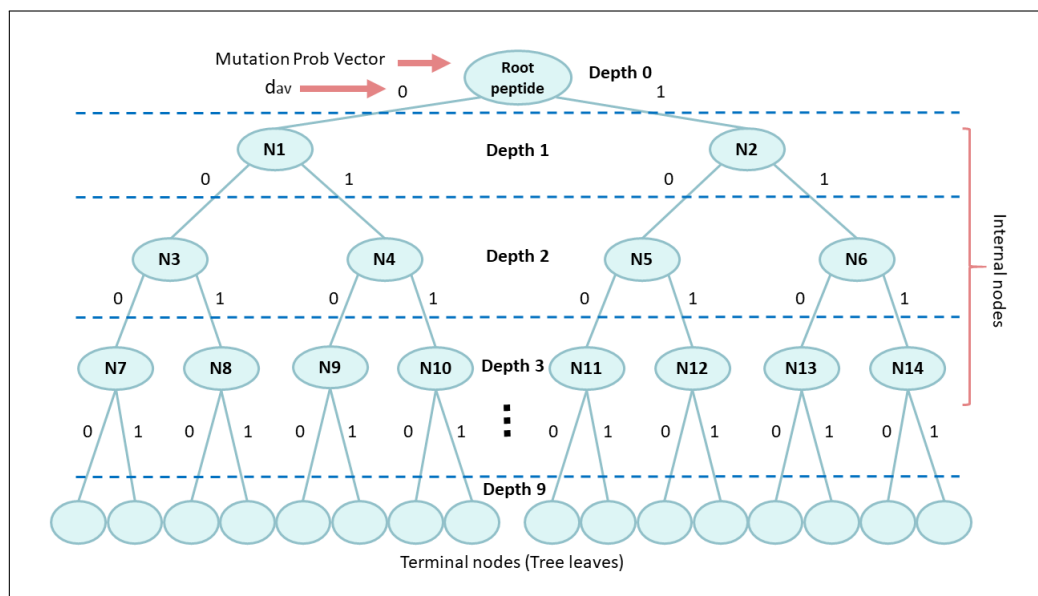


Figure 3.7: Binary mutation guide tree used by ROSE to mutate the root peptide. Peptide libraries may be selected either from the internal nodes (peptides closely related to the root) or from terminal nodes/tree leaves (distantly-related to the root).

Chapter 4

Results and Discussion

4.1 Model Selection

4.1.1 Similarity threshold analysis

From the set of 659 THPs retrieved from starPepDB, 627 peptides were filtered with lower than 98% similarity by local alignment. Before building CSN of 627 peptides, the adequate similarity threshold was chosen. This step is non-trivial since it is the parameter that defines the topology and networks parameters, such as density, modularity, ACC, and singletons [121]. Hence, the appropriate cutoff to build the CSN was determined based on how density, modularity, ACC, and singletons change varying the similarity cutoff. Attachments **C** shows the obtained parameters at each cutoff.

The graph of density, modularity, and ACC as a function of the similarity threshold (Figure 4.1) shows that density is lower at a higher similarity threshold, and ACC follows the same pattern until 0.65 similarity threshold. On the contrary, as the similarity threshold increases, modularity increases, and clustering is optimized.

A well-defined network needs a compromise between the density, modularity, and ACC parameters, but also the number of outliers because they are atypical peptides with particular properties. Networks with very low density result in too many outliers (Attachments **C**), while networks with very high density show a massive connection. In both cases, information is lost and interpretation becomes difficult. Literature reports that, generally, the best density percentages are around 1% or 2.5% due to displaying high modularity and allowing an adequate interpretation of the network [121]. As modularity indicates the existence of community structures, the ideal value must show an equilibrium between non-clustered network, and a network with artificial clusters due to too high modularity value. Based on that, the selected similarity threshold was 0.60 where CSN shows the best parameters and connectivity: 2.3% of density, 0.47 of modularity, 0.428 of ACC, and 99 outliers (15.8% of overall nodes). Therefore, the giant components of the network were 528 nodes.

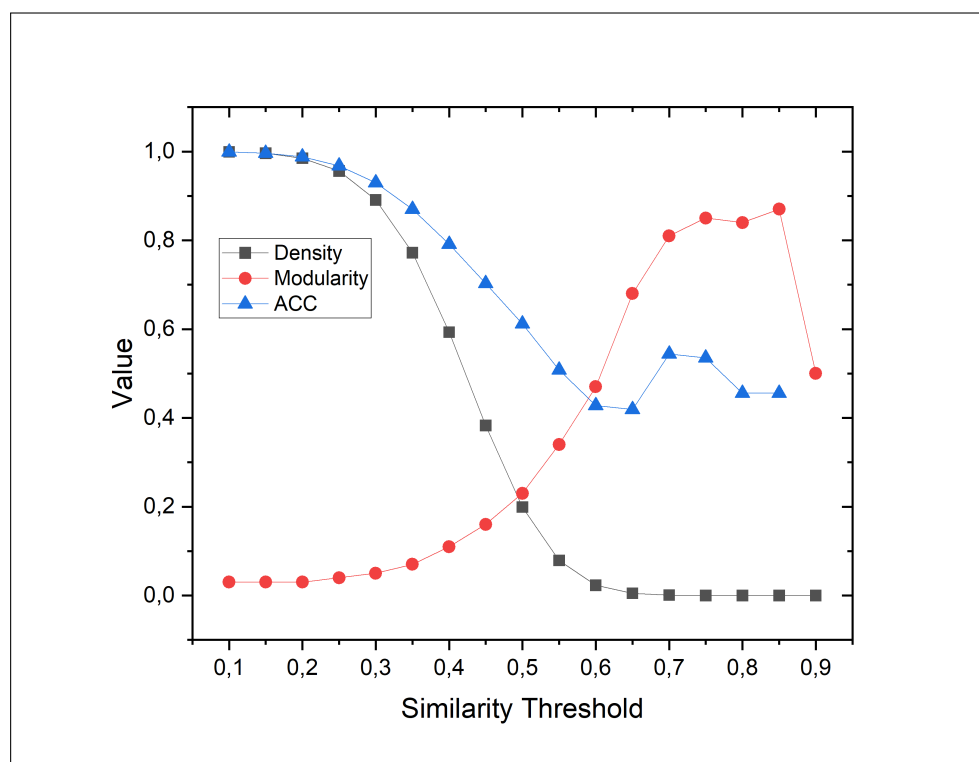


Figure 4.1: Density, modularity, and average clustering coefficient (ACC) as a function of similarity threshold of 627 THPs CSN.

4.1.2 Network characterization

To get a general comprehension of CSNs of the giant component (Figure 4.2) and outliers (Figure 4.4), some parameters were calculated (Table 4.1): density, number of clusters, modularity, average degree, ACC, and diameter.

Additionally, the degree of distribution of the giant components is shown in Figure 4.3. It gives some information about the structure of the CSN. In this case, it can be observed that the degree of distribution is concentrated in the nodes with low vertex degrees, but it has a tail associated to the nodes with higher vertex degrees that are in lower proportion. The nodes with higher degrees correspond to the most central nodes, which, as can be seen in Figure 4.2, are few.

Table 4.1: Global networks properties of CSN of 528 nodes and outliers. Density, number of clusters, and modularity were calculated in starPep toolbox, while average degree, ACC, and diameter in Gephi.

	Nodes	Edges	Density	Clusters	Modularity	Average degree	ACC	Diameter	Nodes after scaffold extraction	Edges after scaffold extraction
THPs	528	4452	0.023	10	0.47	16.864	0.428	8	-	-
Outliers	99	2691	0.891	3	0.13	54.364	0.733	3	34	384

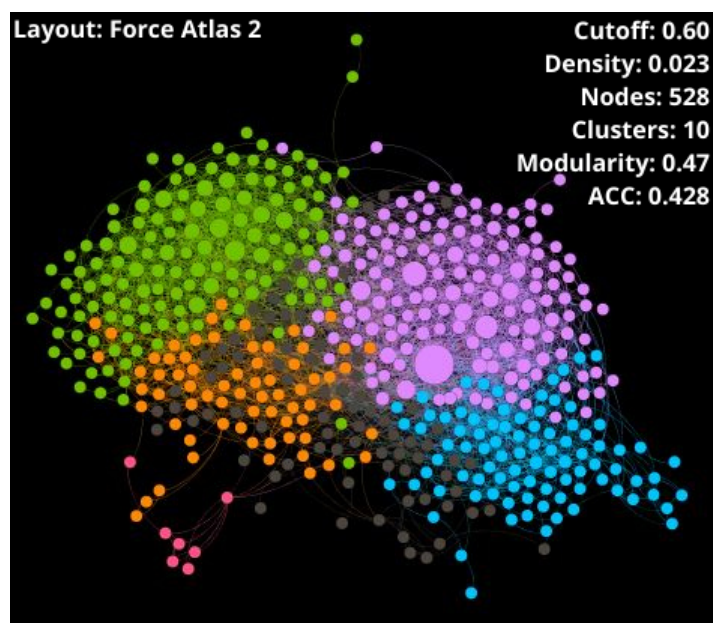


Figure 4.2: CSN of giant component conformed by 528 THPs retrieved from starPepDB. Nodes color represent the community, and size how central the node is.

Outliers are relevant THPs because they present particular characteristics that 528 nodes do not have. CSN of singletons was built using 0.30 of similarity threshold (Figure 4.4a). Then, sequences with higher than 30% similarity by local alignment were removed based on hub-bridge centrality ranking, where 34 outliers with orthogonal sequences were obtained (Figure 4.4b).

4.1.3 Similarity Searching Model for THPs Prediction

Centrality is the key parameter to build the model by which the novel THPs are going to be proposed since it allows the identification of the most influential sequences of the giant components. Moreover, outliers are nodes with unique properties that enriches the model of influential sequences. Therefore, sets from centrality measurements and sets of outliers represented the chemical space of THPs and were used as queries to perform the similarity searching against the target datasets. In total 98 different SSMs were generated, that were based on 22 query sets and similarity thresholds between 0.3 and 0.9.

Table 4.2 shows the results of the similarity searching using the sets obtained after applying 30% of scaffold extraction in the CSN of giant components followed by removing nodes with 10% lower centrality than the most central node as queries against the Main dataset. There, 7 different percentages of identity (30, 40, 50, 60, 70, 80, and 90%) were applied.

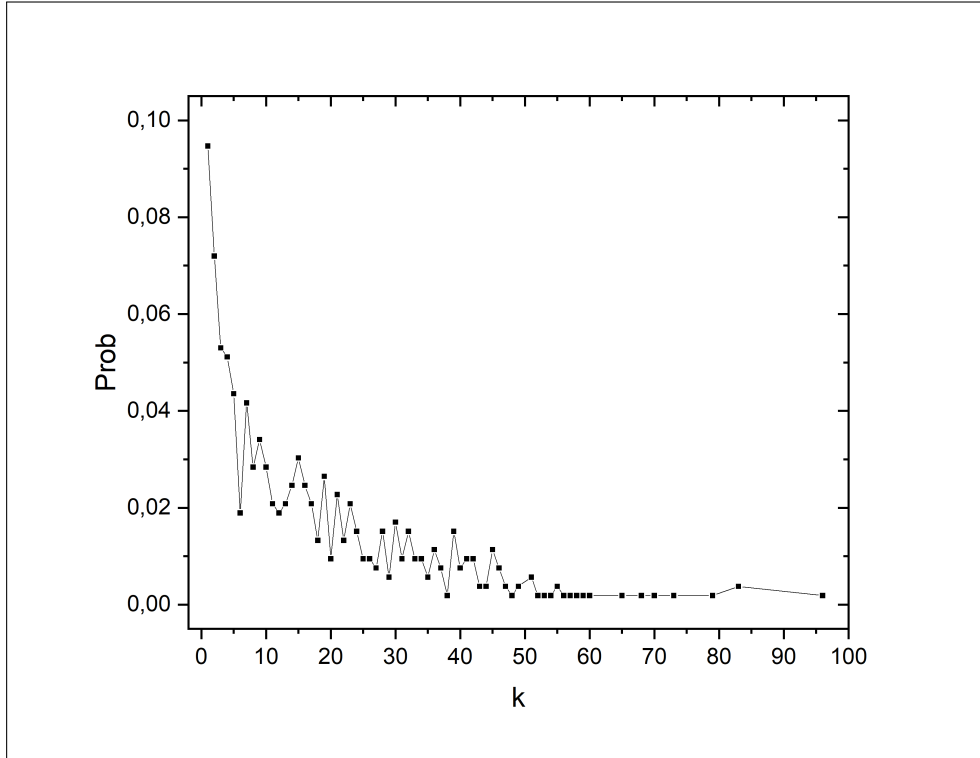


Figure 4.3: Degree distribution of the 528 giant components, where k is the vertex degree.

Results, shown in Table 4.2, indicated that harmonic centrality and weighted degree using 30, 40, 50, 60, and 70% of identity have a better performance with accuracy between 56-58%, recovering between 86-116 nodes. However, these results were not satisfactory considering that the Main dataset consists of 651 active nodes, and the recoveries contained low positive, and many FP nodes, which were reflected in the low R_{TP} and P_{neg} obtained, respectively.

On the other hand, using the sets of outliers as queries, results were also unsatisfactory due to R_{TP} and P_{neg} are low (Table 4.3). However, comparing the two sets of outliers, the set of 99 outliers showed better performance than 34 outliers, and similar statistics as previously obtained using harmonic and weighted degree. This behavior was expected because the outliers are unique sequences with high structural diversity whose similarity to the 528 THPs is less than 60%, so using a set with a higher number of outliers allows recovering more sequences by local alignment.

In order to increase the number of nodes used as queries, sets obtained from the harmonic and weighted degree, and 99 outliers were joined and used as queries (Table 4.4). However, results showed that R_{TP} and P_{neg} remained low.

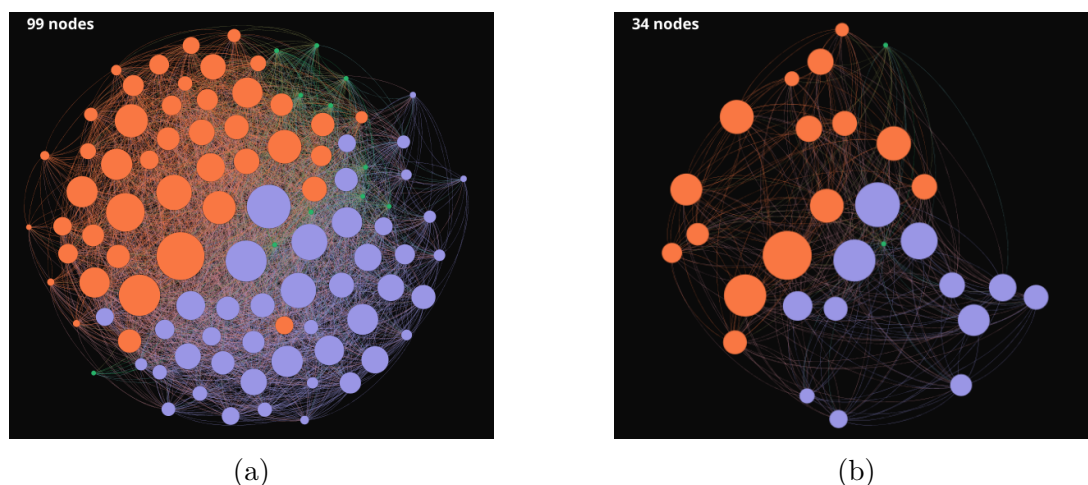


Figure 4.4: CSN of (a) 99 outliers with a density of 0.30 and (b) 34 remaining outliers obtained after 30% similarity extraction scaffold. Layout: Fruchterman Reingold.

Then, sets using 30, 40, 50, and 60% of scaffold extraction based on harmonic and weighted degree were obtained and used as queries. Statistics showed better recovery using 60% of scaffold extraction, where R_{TP} , P_{neg} , kappa, and a number of recovery increase significantly (Table 4.5).

In the last attempt to maximize the model’s performance, sets were obtained from harmonic and weighted degree using 60% of scaffold extraction, and outliers were joined and used as queries in the similarity searching (Table 4.6). Table 4.6 shows that the best performance was achieved using the union of harmonic, weighted degree and 99 outliers sets as queries, in total 479 queries. Moreover, the best percentage of identity at which a compromise of all statistical parameters was achieved with 60%. All statistical parameters showed values greater than 0.88 (4.6).

In general, it is observed that the best performance of query datasets followed the tendency of *weighteddegree* > *harmonic* > *hub – bridge* > *betweenness* > *singletons*. Although, the combination of query datasets from different centrality types exceeds the performance of the sets obtained with only one centrality measure. Moreover, the addition of the sets of outliers improved the performance of the combination sets since it generates the complete representation of the chemical space of THPs.

On the other hand, the performance of the 9 best SSMs was validated in Small and Main90 Datasets. The models used as queries were the union of the set of harmonic with outliers, the set of weighted degree with outliers, and the sets of harmonic, weighted and 99 outliers, all using 60% of scaffold extraction by local alignment, and 40, 50, and 60%

Table 4.2: Results from statistical analysis of recovery performance of using models obtained from 30% of scaffold extraction in the CSN of giant components followed by removing nodes with 10% lower centrality than the most central node as queries. Ac is the accuracy, R_{TP} is the recall of true positives, R_{TN} is the recall of true negatives, PP is the precision of positives, and NP is the precision of negatives.

Centrality	Nodes	% Id	Ac	Correct class.	Incorrect class.	κ	R_{TP}	R_{TN}	P_{pos}	P_{neg}
Weighted degree	54	30	0.588	765	537	0.175	0.177	0.998	0.991	0.548
		40	0.587	764	538	0.174	0.175	0.998	0.991	0.548
		50	0.585	762	540	0.171	0.172	0.998	0.991	0.547
		60	0.582	758	544	0.164	0.164	1	1	0.545
		70	0.566	737	565	0.132	0.132	1	1	0.535
		80	0.558	726	576	0.115	0.115	1	1	0.531
Hub-bridge	31	90	0.549	715	587	0.098	0.098	1	1	0.526
		30	0.564	734	568	0.127	0.129	0.998	0.988	0.534
		40	0.563	733	569	0.126	0.127	0.998	0.988	0.534
		50	0.562	732	570	0.124	0.126	0.998	0.988	0.533
		60	0.562	732	570	0.124	0.124	1	1	0.533
		70	0.547	712	590	0.094	0.094	1	1	0.525
Betweenness	25	80	0.538	701	601	0.077	0.077	1	1	0.52
		90	0.531	692	610	0.063	0.063	1	1	0.516
		30	0.56	729	573	0.12	0.121	0.998	0.988	0.532
		40	0.558	727	575	0.117	0.118	0.998	0.987	0.531
		50	0.558	727	575	0.117	0.118	0.998	0.987	0.531
		60	0.558	727	575	0.117	0.117	1	1	0.531
Harmonic	63	70	0.54	703	599	0.08	0.08	1	1	0.521
		80	0.531	692	610	0.063	0.063	1	1	0.516
		90	0.525	684	618	0.051	0.051	1	1	0.513
		30	0.574	747	555	0.147	0.151	0.997	0.98	0.54
		40	0.573	746	556	0.146	0.149	0.997	0.98	0.539
		50	0.572	745	557	0.144	0.146	0.998	0.99	0.539
		60	0.568	739	563	0.135	0.135	1	1	0.536
		70	0.566	737	565	0.132	0.132	1	1	0.535
		80	0.565	735	567	0.129	0.129	1	1	0.534
		90	0.559	728	574	0.118	0.118	1	1	0.531

identity in the similarity searching. Tables 4.7 and 4.8 show the results obtained and validate the performance of the models.

The best 9 SSMs were compared and ranked using the Friedman test by comparing the multiple statistical metrics of each SSM on the three target datasets (details in Attachments D). According to the test, the best SSM is the set **CSN-TH-0.60Sc-479-H+W+s-0.6-583**. It is composed by the union of nodes with identity lower than 60% from the global centrality harmonic with those obtained applying weighted degree and the set of 99 outliers, in total 479 nodes. The best percentage of identity used to carry out the similarity searching was 60%. The confusion matrices of the better SSM (THP1) are

Table 4.3: Results from statistical analysis of recovery performance when sets of 99 and 34 outliers were used as queries. Ac is the accuracy, R_{TP} is the recall of true positives, R_{TN} is the recall of true negatives, P_{pos} is the precision of positives, and P_{neg} is the precision of negatives.

	Nodes	% Id	Ac	Correct class.	Incorrect class.	κ	R_{TP}	R_{TN}	P_{pos}	P_{neg}
Outliers	99	30	0.585	762	540	0.171	0.174	0.997	0.983	0.547
		40	0.578	752	550	0.155	0.158	0.997	0.981	0.542
		50	0.578	752	550	0.155	0.158	0.997	0.981	0.542
		60	0.577	751	551	0.154	0.157	0.997	0.981	0.542
		70	0.577	751	551	0.154	0.157	0.997	0.981	0.542
		80	0.576	750	552	0.152	0.154	0.998	0.99	0.541
		90	0.576	750	552	0.152	0.152	1	1	0.541
Outliers (30%Sc)	34	30	0.528	688	614	0.057	0.058	0.998	0.974	0.515
		40	0.526	685	617	0.052	0.054	0.998	0.972	0.513
		50	0.526	685	617	0.052	0.054	0.998	0.972	0.513
		60	0.526	685	617	0.052	0.054	0.998	0.972	0.513
		70	0.526	685	617	0.052	0.054	0.998	0.972	0.513
		80	0.526	685	617	0.052	0.052	1	1	0.513
		90	0.526	685	617	0.052	0.052	1	1	0.513

shown in Attachments **E**. It can be seen that the prediction of the model was not random due to MCC was much greater than 0 [178]. Moreover, the performance of statistical metrics showed good results with accuracy, recall, precision, and kappa statistic values higher than 0.85.

Finally, Friedman test of the THP1 versus the reported models used in TumorHPD[9] and THPep[15] servers revealed that the similarity searching methodology to discover potential THPs is superior (details in Attachments **F**). These ML results present a weak predictive ability, where accuracy is 86.56% and 90.13%, and maximal MCC is 0.70 and 0.76, respectively [9, 15]. The test found significant differences between THP1 and the ML models from TumorHPD and THPep servers. Table 4.9 shows the comparison between them on all benchmarking datasets.

4.2 Potential THPs Prediction

4.2.1 Hierarchical Virtual Screening

Molecules to be repurposed using the THP1 were a stronghold of peptides from the chemical space of starPepDB. Starting from 45120 peptides, and after applying the previously

Table 4.4: Results from statistical analysis of recovery performance when the mixture of sets were used as queries. **H** is the set obtained when harmonic centrality was calculated, **W** is the set obtained when the weighted degree was calculated, **Ac** is the accuracy, R_{TP} is the recall of true positives, R_{TN} is the recall of true negatives, P_{pos} is the precision of positives, and P_{neg} is the precision of negatives.

	Nodes	% Id	Ac	Correct class.	Incorrect class.	κ	R_{TP}	R_{TN}	P_{pos}	P_{neg}
H+W	77	30	0.606	789	513	0.212	0.215	0.997	0.986	0.559
		40	0.605	788	514	0.21	0.214	0.997	0.986	0.559
		50	0.604	787	515	0.209	0.21	0.998	0.993	0.558
W+S	167	30	0.664	865	437	0.329	0.333	0.995	0.986	0.599
		40	0.664	864	438	0.327	0.332	0.995	0.986	0.598
		50	0.662	862	440	0.324	0.329	0.995	0.986	0.597
H+S	153	30	0.651	848	454	0.303	0.309	0.994	0.98	0.59
		40	0.651	847	455	0.301	0.307	0.994	0.98	0.589
		50	0.65	846	456	0.3	0.304	0.995	0.985	0.589
H+W+S	176	30	0.683	889	413	0.366	0.372	0.994	0.984	0.613
		40	0.682	888	414	0.364	0.37	0.994	0.984	0.612
		50	0.681	887	415	0.363	0.367	0.995	0.988	0.611

explained filters and performing the similarity searching, 43 lead hits were retrieved (Attachments **G**). Figure 4.5 shows the step-by-step hierarchical virtual screening. Until today, these repurposed sequences do not have reported tumor homing activity, demonstrating their high potential as tumor homing agents.

4.2.2 Tumor Homing Optimization

A library of 180 sequences (Attachments **H**) was obtained from optimization of 43 hits in TumorHPD with a higher TH score than the originals, non-toxicity, and less hemolytic activity. Mutations enriched the sequences in **W** and **C**, where mainly, **G** and **V** residues from originals were mutated to **W**, and **R**, **K**, and also some **W** to **C**. Studies report that the presence of **W** contributes positively to the intracellular translocation of peptides [179]. Moreover, it was reported that **W** enhances the stability of peptides in serum and salt [180].

41 peptides from the library were prioritized by studying their CSN where 50% scaffold extraction by local alignment was accomplished. To perform the scaffold extraction, the sequences were clustered and ranked according to the global harmonic centrality, and only the most central sequences with a similarity between them lower than 50% were kept. 41 sequences have higher predicted TH activity by TumorHPD than original peptides

Table 4.5: Results from statistical analysis of recovery performance of using models obtained from 30, 40, 50, and 60% of scaffold extraction in the CSN of giant components as queries. Ac is the accuracy, R_{TP} is the recall of true positives, R_{TN} is the recall of true negatives, P_{pos} is the precision of positives, and P_{neg} is the precision of negatives.

	% Sc	Nodes	% Id	Ac	Correct class.	Incorrect class.	κ	R_{TP}	R_{TN}	P_{pos}	P_{neg}
Harmonic	30	58	30	0.561	731	571	0.123	0.124	0.998	0.988	0.533
			40	0.561	730	572	0.121	0.123	0.998	0.988	0.532
			50	0.56	729	573	0.12	0.121	0.998	0.988	0.532
	40	140	30	0.658	857	445	0.316	0.323	0.994	0.981	0.595
			40	0.658	857	445	0.316	0.321	0.995	0.986	0.594
			50	0.659	858	444	0.318	0.32	0.998	0.995	0.595
	50	251	30	0.763	993	309	0.525	0.533	0.992	0.986	0.68
			40	0.763	994	308	0.527	0.533	0.994	0.989	0.68
			50	0.765	996	306	0.53	0.533	0.997	0.994	0.681
	60	368	30	0.859	1118	184	0.717	0.727	0.991	0.987	0.784
			40	0.859	1119	183	0.719	0.727	0.992	0.99	0.784
			50	0.862	1122	180	0.724	0.727	0.997	0.996	0.785
Weighted degree	30	60	30	0.589	767	535	0.178	0.18	0.998	0.992	0.549
			40	0.588	766	536	0.177	0.178	0.998	0.991	0.549
			50	0.588	765	537	0.175	0.177	0.998	0.991	0.548
	40	140	30	0.657	855	447	0.313	0.32	0.994	0.981	0.594
			40	0.657	855	447	0.313	0.318	0.995	0.986	0.593
			50	0.657	856	446	0.315	0.316	0.998	0.995	0.594
	50	255	30	0.761	991	311	0.522	0.525	0.997	0.994	0.677
			40	0.76	989	313	0.519	0.525	0.994	0.988	0.677
			50	0.5	651	651	0	0	1	0	0.5
	60	370	30	0.859	1119	183	0.719	0.728	0.991	0.988	0.785
			40	0.86	1120	182	0.72	0.728	0.992	0.99	0.785
			50	0.863	1123	179	0.725	0.728	0.997	0.996	0.786

with scores between 0.39 and 1.92. Moreover, they are anticancer and have less toxicity and hemolytic activity. 12 of 41 sequences come from fragments of original sequences of 5, 10, and 15 lengths; 15 obtained after 4 punctual mutations of originals; and 14 from fragments of mutated sequences of 5, 10, and 15 lengths. Two of 41 peptides, CNGRCGGKLA and WCAMS, are part of reported THPs, which validates the novel methodology to discover potential THPs described here. CNGRCGGKLA is the N-end of CNGRCGGKLAKLAKKLAKLAK peptide which contains NGR TH motif and a disulfide bridge that gives stability. CNGRCGGKLAKLAKKLAKLAK binds to CD13 of tumor cells acting as ACP and THP [181]. While WCAMS is the C-end of KLWCAMS peptide that homes mouse B16B15b melanoma [75].

From the combination of 43 lead and 41 optimized hits, 54 peptides (SET 1) were selected. Sequences from SET 1 present a diverse molecular structure, low toxicity, and

Table 4.6: Results from statistical analysis of recovery performance when a mixture of sets obtained from harmonic and weighted degree using 60% of scaffold extraction, and 99 outliers were used as queries. H is the set obtained when harmonic centrality was calculated, W is the set obtained when the weighted degree was calculated, Ac is the accuracy, R_{TP} is the recall of true positives, R_{TN} is the recall of true negatives, P_{pos} is the precision of positives, and P_{neg} is the precision of negatives.

	Nodes	% Id	Ac	Correct class.	Incorrect class.	κ	R_{TP}	R_{TN}	P_{pos}	P_{neg}
W+H	380	30	0.867	1129	173	0.734	0.743	0.991	0.988	0.794
		40	0.868	1130	172	0.736	0.743	0.992	0.99	0.795
		50	0.87	1133	169	0.74	0.743	0.997	0.996	0.795
		60	0.87	1133	169	0.74	0.74	1	1	0.794
		70	0.849	1106	196	0.699	0.699	1	1	0.769
H+S	467	30	0.932	1214	88	0.865	0.877	0.988	0.986	0.889
		40	0.933	1215	87	0.866	0.877	0.989	0.988	0.89
		50	0.935	1218	84	0.871	0.877	0.994	0.993	0.89
		60	0.935	1218	84	0.871	0.874	0.997	0.996	0.888
		70	0.913	1189	113	0.826	0.829	0.997	0.996	0.854
W+S	469	30	0.933	1215	87	0.866	0.879	0.988	0.986	0.891
		40	0.934	1216	86	0.868	0.879	0.989	0.988	0.891
		50	0.936	1219	83	0.873	0.879	0.994	0.993	0.891
		60	0.937	1220	82	0.874	0.877	0.997	0.997	0.89
		70	0.915	1191	111	0.829	0.833	0.997	0.996	0.856
H+W+S	479	30	0.941	1225	77	0.882	0.894	0.988	0.986	0.903
		40	0.942	1226	76	0.883	0.894	0.989	0.988	0.903
		50	0.944	1229	73	0.888	0.894	0.994	0.993	0.904
		60	0.945	1230	72	0.889	0.892	0.997	0.997	0.903
		70	0.923	1202	100	0.846	0.849	0.997	0.996	0.869

hemolytic activity, and most of them also show potential anticancer activity (Attachments I). The sequence diversity of the lead peptides was evaluated by using all vs. all global alignment where pairwise sequence identities were calculated. As Figures 4.6 and 4.7 show, they all mostly displayed sequence identities lower than 30% indicating structural singularity. Among the 54 lead hits, only one sequence has the well-known NGR motif. Therefore, SET 1 is composed of new structural entities within the known structural space of the THPs.

4.2.3 Discovery of THP motifs

As a consequence of the structural diversity of SET 1, the discovery of motifs accounting for the TH activity is not a straightforward task. In this sense, sensitive multiple sequence alignment (MSA) tools and alignment-free (AF) approaches (e.g., STREME) were applied to unravel new TH motifs.

Table 4.7: Statistic analysis of 9 best SSMs in Small Dataset as target.

	Nodes	% Id	Ac	Correct class.	Incorrect class.	κ	R_{TP}	R_{TN}	P_{pos}	P_{neg}
H+S	467	40	0.917	860	78	0.834	0.838	0.996	0.995	0.86
		50	0.916	859	79	0.832	0.836	0.996	0.995	0.858
		60	0.914	857	81	0.827	0.832	0.996	0.995	0.855
W+S	469	40	0.92	863	75	0.84	0.844	0.996	0.995	0.865
		50	0.92	863	75	0.84	0.844	0.996	0.995	0.865
		60	0.919	862	76	0.838	0.842	0.996	0.995	0.863
H+W+S	479	40	0.928	870	68	0.855	0.859	0.996	0.995	0.876
		50	0.928	870	68	0.855	0.859	0.996	0.995	0.876
		60	0.926	869	69	0.853	0.857	0.996	0.995	0.875

Table 4.8: Statistics analysis of 9 best SSMs in Main90 Dataset as the target.

	Nodes	% Id	Ac	Correct class.	Incorrect class.	κ	R_{TP}	R_{TN}	P_{pos}	P_{neg}
H+S	467	40	0.985	600	9	0.964	0.983	0.986	0.966	0.993
		50	0.99	603	6	0.976	0.983	0.993	0.983	0.993
		60	0.992	604	5	0.98	0.983	0.995	0.989	0.993
W+S	469	40	0.98	597	12	0.952	0.966	0.986	0.966	0.986
		50	0.984	599	10	0.96	0.966	0.991	0.977	0.986
		60	0.987	601	8	0.968	0.966	0.995	0.988	0.986
H+W+S	479	40	0.985	600	9	0.964	0.983	0.986	0.966	0.993
		50	0.989	602	7	0.972	0.983	0.991	0.977	0.993
		60	0.992	604	5	0.98	0.983	0.995	0.989	0.993

To make possible the application of MSA algorithms for motif identification, the resulting 54 lead THPs were mapped onto CSN space to identify putative communities. These networks communities were considered clusters containing related peptides. Finally, 6 clusters were conformed with 14, 10, 8, 4, 10, and 8 members, respectively. The last cluster grouped the singletons (peptides identified as atypical in the CSN).

Clustal-Omega [169], MAFFT [170], MUSCLE [171], and T-Coffee [172] which are MSA algorithms developed after the classical ClustalW were applied, so that they can deal with hard-to-align sequences shown in each cluster, and thus to detect any conserved signature or motif. Since each MSA has implemented a different algorithm to improve alignment quality, their altogether consideration for the estimation of consensus regions helped us to identify TH motifs by using the Jalview, EMBOSS Cons and Seq2Logo programs (Attachments J). As the EMBOSS Cons, gives a more legible output, only displaying high scored amino acids/positions (capital letters), less scored but positive residues (lower-case letters), and non-consensus positions (x), were selected as

Table 4.9: Comparison between the best SSM THP1 to predict THPs and the ML models reported in the literature as tumor homing benchmarking tests. P_{pos} corresponds to the sensibility, and $R_{TP}(\%)$ to specificity.

Dataset	Method	Ac(%)	$P_{pos}(\%)$	$R_{TP}(\%)$	MCC
Main	TumorHPD	86.56	80.63	89.71	0.7
	THPep	86.1	87.07	85.18	0.72
	THP1	94.47	99.66	89.25	0.894
Small	TumorHPD	81.88	73.13	90.92	0.65
	THPep	83.37	81.24	85.81	0.67
	THP1	92.64	99.5	85.71	0.861
Main90	TumorHPD	89.66	83.64	80.68	0.74
	THPep	90.8	91.8	87.97	0.77
	THP1	99.18	98.86	98.3	0.98

primary source to set consensus/conserved regions. The non-consensus positions were estimated using default parameters by visual inspection of the corresponding positions in the Jalview program [173] and in the Seq2Logo [174]. Table 4.10 depicts the consensus motifs, unraveled by each MSA algorithm.

Table 4.10: Discovered motifs by Multiple Sequence Alignment (MSA). **Taken from TumorHoPe (outside parenthesis), and starPepDB (inside parenthesis).

No.	Motif	EMBOSS consensus	Cluster	Cluster size	MSA Method	Frequency**
1	wwW	wwW	2	14	CLUSTALW-O	1/(1)
		xxW			MAFFT	0/(0)
2	C[fl][rg][vl]rW	CxxxrW	3	10	MAFFT	0/(0)
3	C[gpI][gs]cR	CxxxR			MUSCLE	0/(0)
4	[rkl]GLC	RGlc	4	8	CLUSTALW-O	0/(0)
		kGLC			MAFFT	0/(0)
		xGLc			MUSCLE	0/(0)
5	c[wp]kG	cwkG	1+5	4	CLUSTALW-O	0/(0)
		cxkG			MUSCLE	0/(0)
6	Not found	Non-consensus	6	10	T-Coffee	0/(1)
					CLUSTALW-O	0
					MUSCLE	
7	l[rp][cw]c	lxxc	Singletons	8	MUSCLE	0/(0)
					T-Coffee	

None of the motifs found by MSA have been reported as TH motifs. However, one of the motifs from No.3 “CxxxR”, “CGGCR” contains “CXXC” motif which is the active site of thioredoxin (Trx), a relevant protein in mammalian cells that act as an antioxidant and participates in programmed cell death inhibition and cell growth, commonly used as a target for cancer treatments [182, 183]. Moreover, “CIGCR” (from No.3 “CxxxR”) is

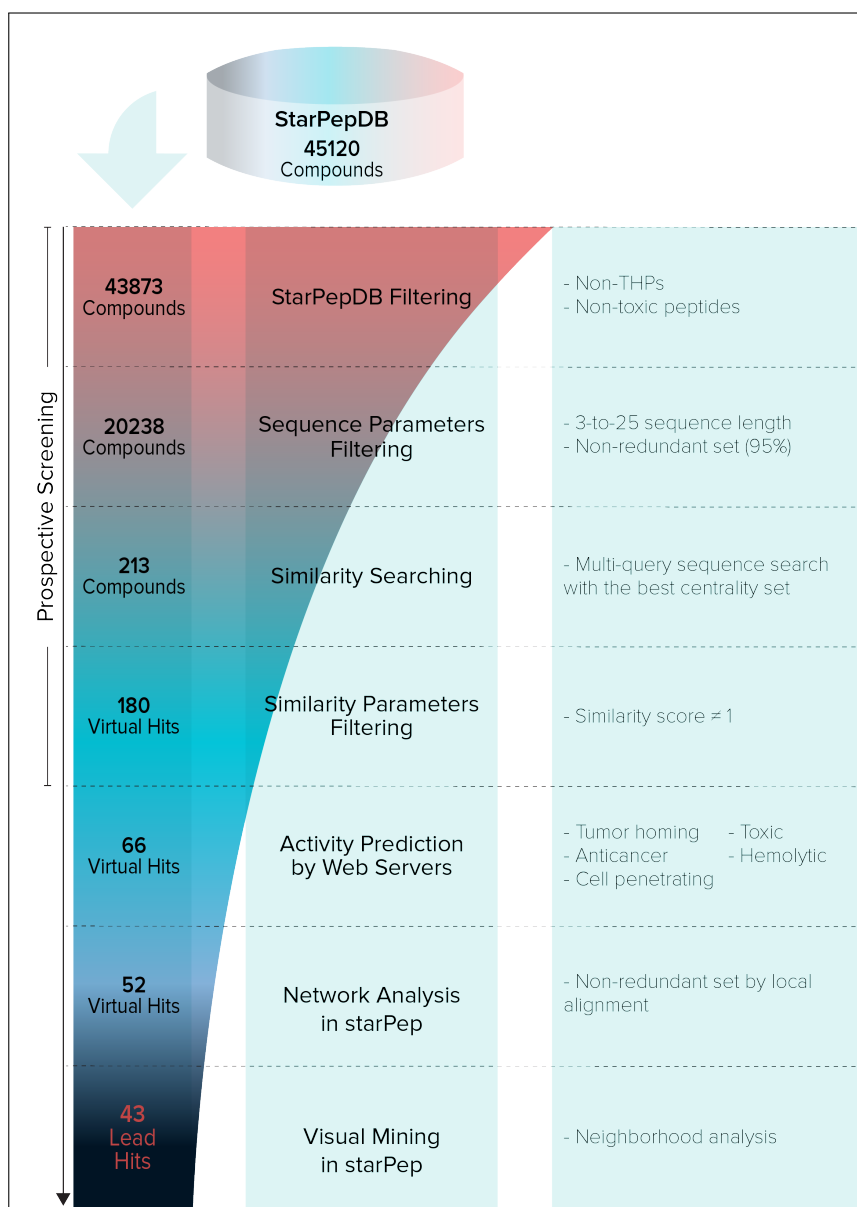


Figure 4.5: Hierarchical virtual screening for repurposing of peptides from starPepDB.

a motif from Epstein-Barr nuclear antigen 1 (EBNA1) epitope which binds to G protein-coupled receptor in pregnant women, related to pre-eclampsia [184], and “CWKG” (No.5) is contained in a nanoscale molecular platform used as drug delivery system in chemotherapy to enhance the conjugation of mitomycin C to the carrier [185].

On the other hand, the AF approach STREME was used to find unaligned patterns ranging 3-5 aa length within the overall 54 peptides and within each peptide cluster. STREME has been recently reported as the most accurate and sensitive algorithm among its competing state-of-art partners [175]. Unlike previous algorithms [186–188], STREME uses a position weight matrix (PWM) to count position matches efficiently for a motif

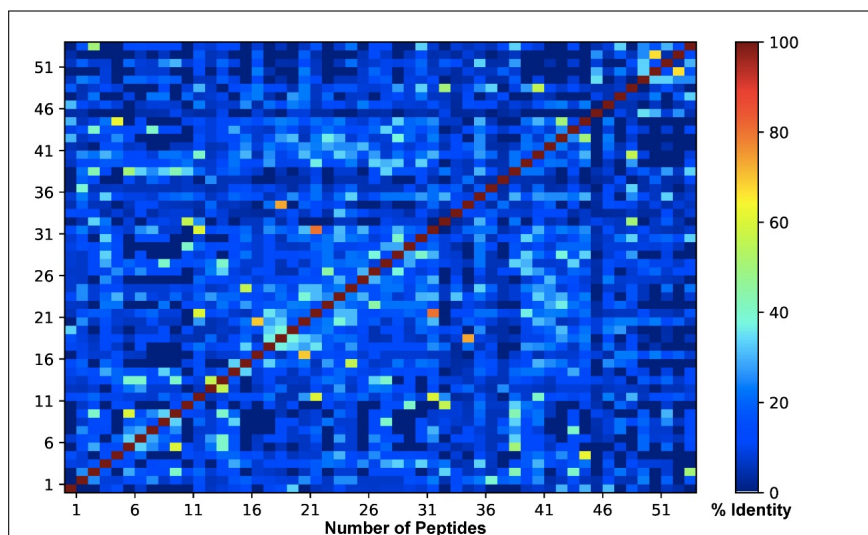


Figure 4.6: Heat map of SET 1 (54 lead compounds).

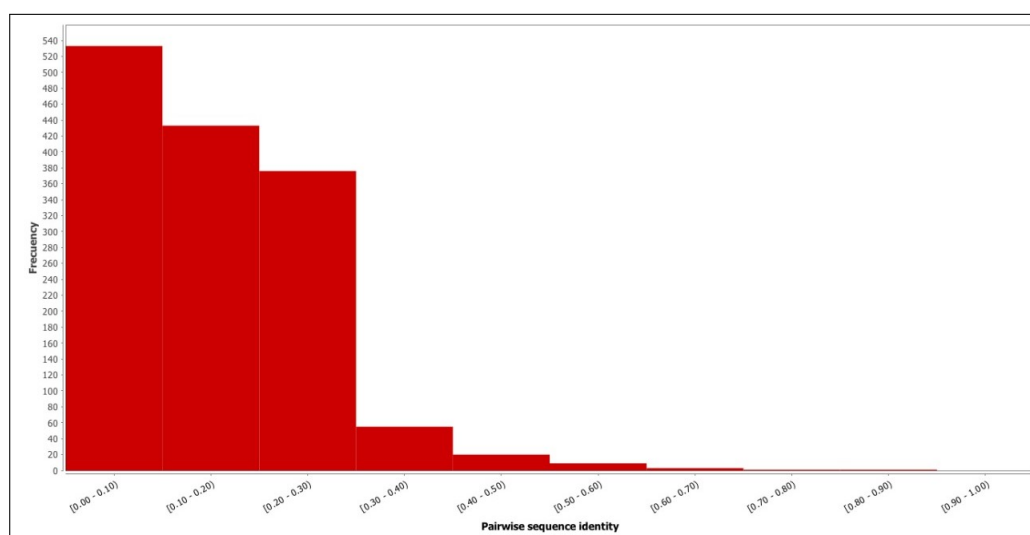


Figure 4.7: Histogram of pairwise sequence identity of SET 1 (54 lead compounds).

candidate against a Markov model built up from shuffling the same input set (control sequences). Table 4.11 displays the enriched motifs discriminating the 54 lead peptides against the control sequences. The same search was also performed by considering each cluster content. Motifs appearing in more than 20% of the query sequences are listed according to their statistical significance (score).

One of the motifs discovered by STREME had been reported as tumor homing, “WRP” which interacts with VEGF-C (Table 2.1) [82, 83]. Moreover, other found motifs were reported but not as TH, such as “WRPW”, “PRW”, “WKG”, and “PSHL”. “WRPW”, which contains “WRP” motif, is the binding site of 7 Enhancer of split E(spl) basic helix-loop-helix (bHLH) and Hairy proteins to the WD40 domain of corepressor

Table 4.11: Discovered Motifs by STREME. **Taken from TumorHoPe (outside parenthesis), and starPepDB (inside parenthesis).

No.	Motif	Cluster	Cluster size	Matches in positive seqs.	Matches in control seq.	Sites (%)	Score	Frequency**
1	WRP			7	1	50	1.6e-002	5/(5)
2	WVL	2	14	5	1	35.7	8.2e-002	0/(0)
3	WS[YR]			3	0	21.4	1.1e-001	1/(1)Y
4	WWWM			3	0	21.4	1.1e-001	0/(0)
5	CFRV			3	0	30	1.1e-001	1/(1)
6	HWK	3	10	2	0	20	2.4e-001	0/(0)
7	PRW			2	0	20	2.4e-001	3/(3)
8	CN[WG]			3	0	37.5	1.0e-001	34/(32)G
9	WARG	4	8	3	0	37.5	1.0e-001	0/(0)
10	GIG			2	0	25.0	2.3e-001	5/(4)
11	WKG	1-5	4	3	1	75.0	2.4e-001	0/(0)
12	KNKHK	6	10	3	0	30.0	1.1e-001	0/(0)
13	PSHL			3	0	30.0	1.1e-001	0/(0)
14	LRLRI	Singletons	8	2	0	25.0	2.3e-001	1/(1)
15	CC[CQ]			3	1	37.5	2.8e-001	0/(0)
16	LSP	All	54	11	1	20.4	3.4e-003	3/(3)
17	WSYG	sequences		7	0	13.0	8.2e-003	0/(0)
18	WRPW			5	0	9.3	3.2e-002	2/(2)

protein Groucho-TLE [189]. “PRW” is part of a biocatalyst, where it is conjugated to a lipid by an ester or amide bond [190]. “WKG” is a ribosomally synthesized and post-translationally modified peptide [191]. Moreover, “PSHL” is a tetrapeptide that affects the maturation and activation of HIV-1 protease (PR) [192].

Lastly, 54 lead THPs were queried against PROSITE’s pattern and profile databases by using the search engine Motif Search of the GenomeNet suite [193]. Only two query peptides had significant matches with motifs found in Gonadotropin-releasing hormones (GnRH) and Bombesin-like peptides (Table 4.12).

Table 4.12: Motifs found in PROSITE. **Taken from TumorHoPe (outside parenthesis), and starPepDB (inside parenthesis).

No.	Motif found	Hit Peptide	Accession	Match with	Signature	Related Seqs.	Frequency**
1	QHWSYGLRPG	starPep_07237	PS00473	Q[HY][FYW]Sx(4)PG	Gonadotropin-releasing hormones	67	1/(1)QHWSY
2	WARGHFM	starPep_10020	PS00257	WAxG[SH][LF]M	Bombesin-like peptides	36	0/(0)

These two peptide signatures and their receptors are involved in neuroendocrine signaling pathways associated with physiological states and tumors. GnRH is the hypothalamic decapeptide that plays a key role in the control of women's reproductive cycle. GnRH binds to specific receptors on the pituitary gonadotrophic cells, but it also is expressed in other reproductive organs, e.g. ovaries, and tumors derived from the ovaries. It has been shown GnRH is involved in the regulation of proliferation and metastasis of ovarian cancer either by indirect signaling pathway or direct interaction with the GnRH receptors placed at the surface of ovarian cancer cells [194].

Bombesin-like peptides were initially discovered from the frog skin, where they are secreted from cutaneous glands as a means of communication and defense. They were later found to be widely distributed in mammalian neural and endocrine cells represented by the neuromedin B (NMB) and the gastrin-releasing peptide (GRP), respectively. Bombesin-like peptide receptors are G protein-coupled and have seven membrane-spanning domains, so they are involved in signal transduction pathways [195]. Growing evidence shows bombesin-like peptides and their receptors play important roles in both physiological conditions and diseases. In fact, an abnormal expression of bombesin receptors has been observed in several types of tumors, which has motivated the development of more specific and safer bombesin-derivatives for tumors diagnosis and therapy [103].

The motif search by using different approaches may render a diversity of outcomes. However, some hits shared by different search approaches can support the reliability of the findings. For example, one motif "WSY" found by the PROSITE search was also encountered by STREME, an algorithm that works regardless of database and sequence similarity. Some of the motifs estimated by MSA algorithms were also identified by the AF approach STREME such as "WWW" and "WKG". All motifs were searched against TH databases, TumorHoPe, and starPepDB, in order to discriminate the possible new signatures from the existing ones (Table 2.1). New motifs appear at very low frequency contained within THPs (last column of Tables 4.10, 4.11 and 4.12), except "CNG" found by STREME, which appears 34 times in TumorHoPe and 32 in starPepDB. However, "CNG" has not been reported as TH motif.

4.3 Multi-Objective Optimization of THPs

4.3.1 Cell-Penetrating activity

Sequences from SET 1 show potent TH activity and singularity, but only 7 of them are permeable into cells. Improving their permeability was important to enhance their therapeutic activity due to it facilitates drug targeting. Thus, a library of 150 sequences (Attachments **K**) was obtained by punctual aa mutations, mainly to positively charged R or K residues, using CellPPD.

SET 1 and the library of 150 optimized sequences were combined and reduced by scaffold extractions and similarity searching. The stronghold is a SET 2 of 42 hits with TH scores between 0.19-3.61 according to TumorHPD, non-toxic by all models of ToxinPred, anticancer by at least one of the AntiCP models, and non-hemolytic by at least three models of HemoPI, where 34 hits are CPPs by at least one of CellPPD. Attachments **L** shows their predicted activities.

It was difficult to achieve cell-penetrating peptides according to both SVM models of CellPPD server while keeping the tumor homing activity, low toxicity and hemolysis due to mutations change considerably their structure affecting activity. Therefore, an alternative cell-penetrating optimization was performed based on the conjugation of the sequence with a well-known cell-penetrating sequence, such as TAT. The conjugation was applied as final step, after the selection of putative THPs, thus it is explained in detail later.

4.3.2 Half-Life Time

Half-life is a highly relevant parameter of therapeutic peptides since it governs the drug's pharmacokinetics, in consequence, the activity [196]. It directly influences the bioavailability, biodistribution, and necessary dosage of the drug. It is known that peptides show a short half-life in the gastrointestinal tract due to protease cleavage, which is the main reason why the preferred route of administration is parental [197]. Nevertheless, peptides present a short half-life in circulation as a result of enzymatic degradation but also renal clearance [33]. Therefore, it is essential to evaluate the theoretical half-life in the blood and digestive system of THPs and try to improve their stability, mainly by aa mutations.

Half-life in the blood of 42 sequences was improved by punctual mutations and shortening in fragments of 5 and 10 residues in PlifePred. A library of 206 sequences (Attachments **M**) with tumor homing activity, non-toxic, and non-hemolytic was obtained. After the application of scaffold extractions and similarity searching in the peptides from the library, 59 sequences (SET 3) with higher stability in blood than originals were retrieved. Sequences from SET 3 have predicted half-life time in blood between 13 to 33 minutes.

On the other hand, the stability of 59 sequences in the gastrointestinal tract was optimized using HLP by punctual mutations. The selection of sequences was based on those with predicted stability labeled as high (higher than 1 second) or normal (0.1-1 second) according to the server. Sequences with similar or higher TH scores than originals, anticancer activity, non-toxic and non-hemolytic were filtered, resulting in a library of 250 sequences (Attachments **N**). From the library, 78 sequences were retrieved (SET 4) after scaffold extractions and similarity searching. These sequences are TH active by the two servers (THP and TumorHPD) with a score between 0.46-3.30, non-toxic, the majority are anticancer by almost one SVM model of AntiCP, have a half-life in blood between 11-30 minutes with normal or high (0.799-6.599 seconds) gastrointestinal stability. Moreover, sequences from SET 4 were characterized by predicting other properties, including aggregation, and allergenic reaction (Attachments **O**).

To find a compromise between blood and gastrointestinal half-life time was not easy since neutral residues (E, S, T, and G) stabilized peptides in the gastrointestinal [141], while they decrease half-life in blood [28, 198]. Moreover, the proposed THPs was short sequences and small structural alterations modify their activity.

Although the stability of the sequences has been slightly increased, predicted half-life times are not adequate, considering that the chemotherapeutic drug needs a half-life of more than hours to days to decrease the number of doses administered to the patient [199]. Thus, parental administration is preferable.

On the other hand, the majority of sequences lost penetrability. The main reason was cell penetrability improves by increasing positively charged residues such as R and K, but with increasing charge, the sequence is more prone to degradation by the action of proteases [141].

Finally, the 78 sequences were reduced to 13 lead peptides (SET 5) (highlighted sequences in Attachments **O**) that accomplished SVM scores shown in Table 3.1. Compared

to the previously obtained SET 1, sequences from SET 5 have a higher potential to be THPs because scoring thresholds were more stringent. Nevertheless, the half-life is still low and a limiting factor in their pharmacokinetics. Thus, other optimization routes should be sought, such as increasing molecular weight or conjugation with stability enhancers [200].

4.3.3 Building Blocks

In a final attempt to enhance the tumor homing activity of SET 4, the reported motifs (Table 2.1), discovered in this study (Tables 4.10, 4.11 and 4.12), and the short sequences of SET 4 shown in Table 4.13 were attached and inserted to the 78 sequences from SET 4 and to motifs (Tables 2.1, 4.10, 4.11 and 4.12).

Table 4.13: Short optimized THPs with 5-8 aa length.

No	Sequence	SVM Score
1	WPGCHSWA	3.12
2	CSKGC	3.01
3	CRPGC	2.94
4	CRCGF	2.92
5	PYWLP	2.82
6	CPCKL	2.65
7	WRQLPWFG	2.52
8	PLSWA	2.5
9	AFPSWRM	2.36
10	AMDSRWM	2.22
11	YWRGF	2.07
12	ECGFW	2.03

In total, a library of 7923 sequences was built by the addition of motifs in the N-end, C-end, and 3 random positions. This large library was reduced to 62 lead peptides (SET 6), where all are TH active by the ML servers, and 13 are TH active by the THP1 model.

On the other hand, the library was filtered keeping only the non-redundant sequences with cell-penetrating activity, deriving 10 peptides (SET 7).

4.3.4 Final Selection of Potential THPs

At this point, there are 3 sets of lead sequences with optimized activities (SET 5, 6, and 7), totaling 85 sequences. As they were obtained in different steps, they presented redundancy

among the sequences. Therefore, redundant sequences were removed only from SET 5 and 6 by scaffold extraction, resulting in a set of 39 lead sequences. SET 7 was not reduced because they were sequences with high cell-penetrating potential according to both SVM models from CellPPD.

Finally, the 39 lead sequences were combined with SET 7, and peptides with the highest TH activity (highest ML scores) and the trade-off between all the different predicted activities were filtered. The result was a set of positively charged 27 potential THPs (SET: Putative THPs).

Predicted activities are shown in Attachments **P**. The range of size of 27 putative THPs is between 7-11 aa residues. In general, their physicochemical properties show a low score of hydrophobicity meaning that they were more hydrophilic, positively influencing solubility [201]. They are tumor homing according to the prediction of all SVM methods with a score between 1.16-3.34. Additionally, they are non-toxic, non-hemolytic, and they are anticancer according to all SVM models. According to the immunogenicity of the compounds, the scores that indicate how much allergic reaction they produce is less than 0.416, and they do not induce IL-10, an interleukin that has both tumor-inhibitory and tumor-promoting activity [202]. The scores that determine IL-4 induction are low, however, the production of this type of interleukin has an antiangiogenic effect and favors tumor cell growth inhibition [203]. On the contrary, predictions show that all the molecules are inducers of IL-12, an interleukin widely studied for its immunotherapy potential in cancer treatments because it mediates tumor regression [203]. In addition, they may exhibit other activities such as antimicrobial or antiviral agents.

Putative THPs showed simple structures and were classified as random coiled or single helix (Figure 4.8). The next step would be to test their tumor homing activity experimentally, to corroborate their potential.

***De novo* Design of ACPs**

From the 27 sequences, 14 sequences with higher anticancer scores, and commitment to TH scores, low toxicity, low hemolysis, and high solubility were prioritized. 8 of them were mutated by ROSE to build an extended peptide library as a source of finding new ACPs with enhanced cell permeability. Then, the resulting libraries were screened to identify putative ACPs by using the several ML-based programs trained to identify some

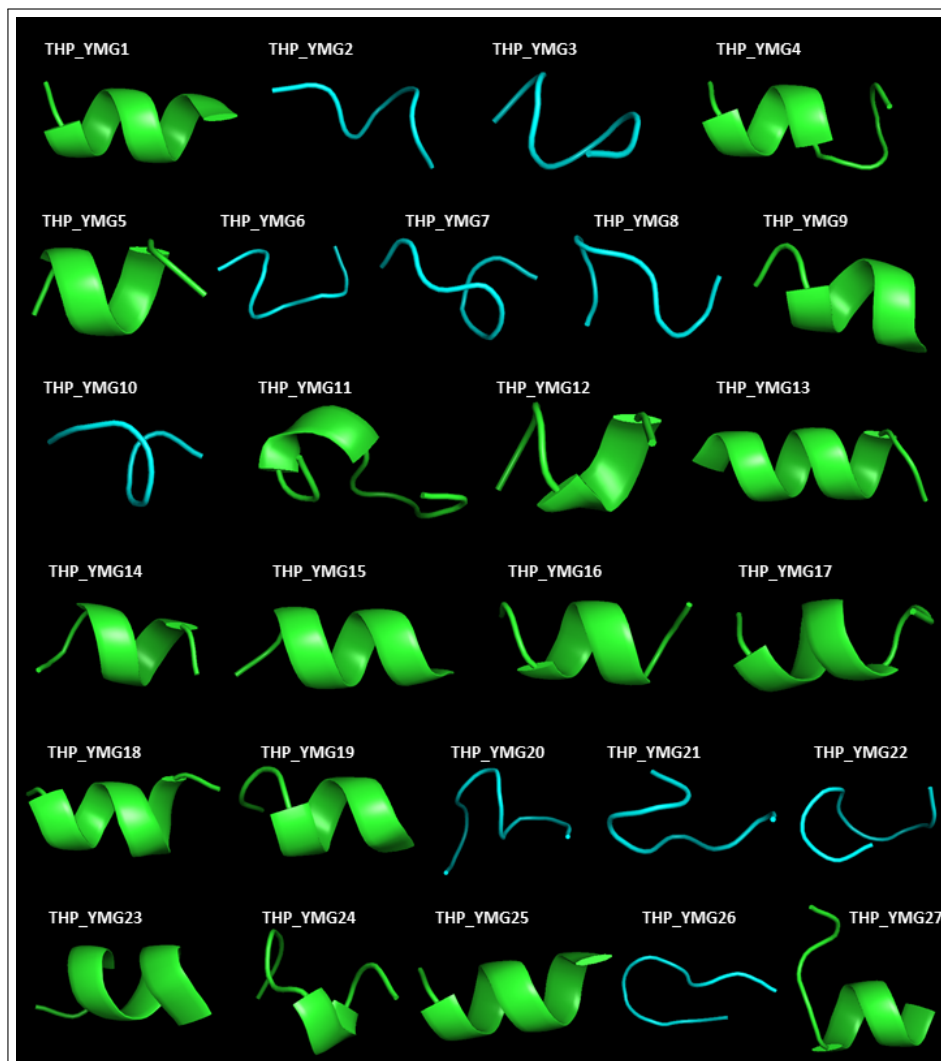


Figure 4.8: 3D structure of 27 putative THPs generated with PEP-FOLD 3 and visualized with PyMOL.

therapeutic peptides (Table 2.2). Finally, among the top-ranked candidates, i.e., those with higher anticancer scores according to AntiCP, lower human toxicity, and higher % of sequence similarity to ancestor peptides, the most potent and orthogonal 8 sequences were selected.

In general, the application of evolutionary approaches has been devoted to the optimization steps of peptide drugs. Here, a different approach for the design of bioactive peptides is proposed, which also leverages ML models and evolutionary algorithms but in a different mode. The strategy repurposes the simulation of sequences evolution to the rational generation of diversity-oriented peptide libraries that are subsequently explored with ML models of several pharmacological and ADME-TOX endpoints. This is achieved by applying a flexible evolutionary algorithm, as implemented in ROSE, that

comprises parameters such as average genetic distance, tree topology, and insertion and deletion events, among others. The advantage of using evolutionary algorithms to build libraries of candidates lies in the application of previous knowledge on the sites/residues that account for biological activity when mutations are performed. Thus, a consensus (root) peptide with its corresponding conservation scoring profile can be used to assign different mutation rates to each position in the sequence.

On the other hand, the sequence diversity of the peptides in the library can be controlled by evaluating the ROSE output with an all vs all global alignment [204]. Here, ROSE parameters were calibrated to produce peptide libraries with an overall 60% of identity by using the software starPep toolbox. All these evolutionary considerations provide rationality to the generation of peptide libraries. Thus, the probability to find new biologically relevant peptides is higher than approaches using stochastic mutations. The resulting peptide library was screened with several web servers to identify putative ACPs and, at the same time, to diminish the likelihood of action with the human counterpart as well ADME properties like cell permeability.

Table 4.14 shows the physicochemical properties of ACPs. Moreover, Table 4.15 summarizes the obtained scores for the 14 putative ACPs and Figure 4.9 shows their 3D structures generated with PEP-FOLD 3. Notably, ACP-YMG1 and ACP-YMG10, which is derived from one of the motif WRP shown in Table 2.1, is predicted to potentially bind VEGF-C. Similarly, ACP-YMG12 is predicted to bind to the tumor neovasculature, when precisely this peptide originates from the motif PSP. The other structures do not have known motifs, so *a priori* it is not possible to determine what they would be bound to and require experimental studies. Putative ACPs also showed a simple structure that can be classified as the random coiled or single helix.

Table 4.14: Physicochemical properties of 14 putative ACPs.

CodeID	Sequence	Length	Hydrophobicity	Steric hindrance	Sidebulk	Hydrophaticity	Amphipathicity	Hydrophilicity	Net Hydrogen	Charge	pI	Mol wt
ACP.YMG1	WRPWPSHL	8	-0.14	0.38	0.38	-1.08	0.43	-0.64	0.89	1.5	10.11	1191.53
ACP.YMG2	EKFWPRSG	8	-0.3	0.53	0.53	-1.42	0.82	0.38	1	1	9.1	1063.3
ACP.YMG3	PRWPLSWA	8	-0.07	0.44	0.44	-0.52	0.27	-0.64	0.78	1	10.11	1083.37
ACP.YMG4	HHGTPRWC	8	-0.25	0.37	0.37	-1.33	0.59	-0.31	0.89	2	8.61	1096.37
ACP.YMG5	PSPAFKWW	8	0.01	0.46	0.46	-0.57	0.41	-0.72	0.56	1	9.11	1204.51
ACP.YMG6	AMYWRGFWWP	10	0.05	0.54	0.54	-0.36	0.22	-1.25	0.73	1	9.1	1496.9
ACP.YMG7	CTGCQNWWM	9	-0.03	0.57	0.57	-0.3	0.12	-1.01	0.7	0	5.82	1259.64
ACP.YMG8	WWYWRGFWM	9	0.08	0.55	0.55	-0.51	0.25	-1.67	0.9	1	9.1	1548.99
ACP.YMG9	LPWCKRLRT	9	-0.34	0.51	0.51	-0.6	0.86	0.06	1.2	3	10.87	1273.7
ACP.YMG10	WRPGSWAKQALKSI	14	-0.17	0.54	0.54	-0.62	0.74	-0.11	0.93	3	11.17	1741.29
ACP.YMG11	CYLHSSSCGCHNCK	15	-0.17	0.5	0.5	-0.31	0.41	-0.29	0.69	2	8.17	1757.22
ACP.YMG12	SNWWRLKT	8	-0.3	0.52	0.52	-1.27	0.68	-0.28	1.33	2	11.01	1191.48
ACP.YMG13	GKWARGW	7	-0.19	0.53	0.53	-1.15	0.77	-0.16	1	2	11.01	1046.31
ACP.YMG14	RQRICVPRR	9	-0.65	0.58	0.58	-1.19	1.1	0.79	1.8	4	12.01	1339.76

On the other hand, when adding TAT and A-TAT, permeability was enhanced. How-

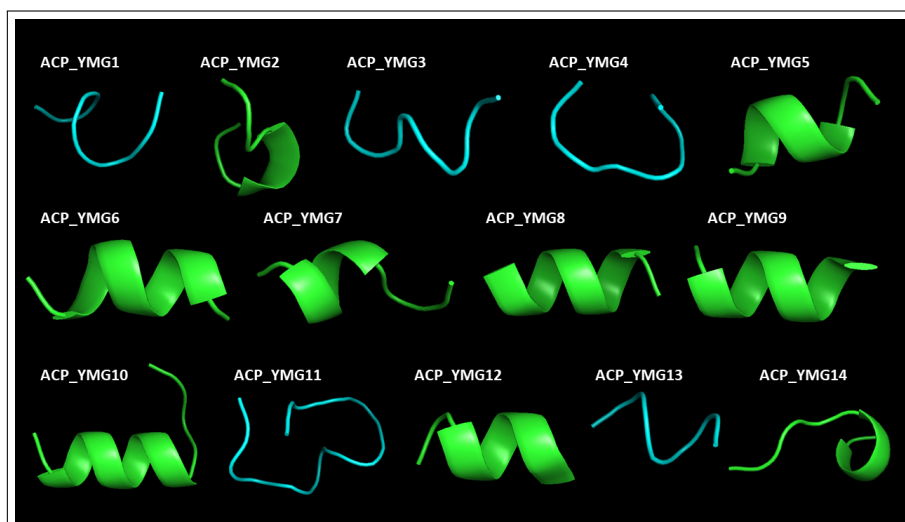


Figure 4.9: 3D structure of 15 putative ACPs generated with PEP-FOLD 3 and visualized with PyMOL.

ever, the predicted tumor homing action was considerably decreased. When comparing the 3D structures of sequences with and without TAT, it could be observed that in some sequences the peptide conformational structure changed subtly when A-TAT was added and not when only TAT was added, but different domains are not formed. Therefore, the peptides are expected to maintain the predicted activities when bound to TAT. Nevertheless, it requires further studies.

Table 4.15: Tumor homing and anticancer predictions of 14 putative ACPs.

CodeID	Sequence	TumorHPD SVM Score	THPep	AntiCP SVM Scores		iACP Prob	CpACpP Prediction					ACPred Prob	ENNAACT ACP Prob	AMPfun Score	
ACP.YMG1	WRPWPSHL	3.28	THP	1.11	0.78	0.147606	ACP	CpACP	ACP	CpACP	ACP	CpACP	0.977	0.776	0.4404
ACP.YMG2	EKFWPRSG	0.39	THP	1.06	0	0.965416	non-ACP	non-CpACP	ACP	CpACP	ACP	CpACP	0.988	0.095	0.4725
ACP.YMG3	PRWPLSWA	3.14	THP	1.02	0.87	0.959603	ACP	CpACP	ACP	CpACP	ACP	CpACP	0.819	0.095	0.385
ACP.YMG4	HHGTFRWC	2.07	THP	1.02	1.2	0.959603	ACP	CpACP	ACP	CpACP	ACP	CpACP	0.965	0.775	0.5314
ACP.YMG5	PSPAFKWW	2.07	THP	1.05	0.88	0.765597	ACP	CpACP	ACP	CpACP	ACP	CpACP	0.989	0.574	0.3503
ACP.YMG6	AMYWRGFWWP	3.02	THP	1.04	1.24	0.525148	ACP	CpACP	ACP	CpACP	ACP	CpACP	0.98	0.585	0.6715
ACP.YMG7	CTGCQNWWM	2.34	THP	1.16	0.91	0.701894	ACP	non-CpACP	ACP	non-CpACP	ACP	non-CpACP	0.954	0.653	0.4235
ACP.YMG8	WWYWRGFWM	2.78	THP	0.76	0.81	0.959603	ACP	CpACP	ACP	CpACP	ACP	CpACP	0.989	0.993	0.6427
ACP.YMG9	LPWCKRLRT	1.63	THP	1.09	1.41	0.482859	ACP	CpACP	ACP	CpACP	ACP	CpACP	0.084	0.998	0.3624
ACP.YMG10	WRPGSWAKQALKSI	0.19	THP	0.85	1.01	0.874125	ACP	CpACP	ACP	CpACP	ACP	CpACP	0.984	0.85	0.0375
ACP.YMG11	CYLHSSSCGCHNCK	2.51	THP	0.64	0.98	0.682829	ACP	non-CpACP	ACP	non-CpACP	ACP	non-CpACP	0.994	0.736	0.5205
ACP.YMG12	SNWWRLKT	1.49	THP	0.9	0.23	0.959603	ACP	CpACP	ACP	CpACP	ACP	CpACP	0.957	0.483	0.3282
ACP.YMG13	GKWARGW	2.06	THP	-0.04	1.41	0.985151	ACP	CpACP	ACP	CpACP	ACP	CpACP	0.987	0.997	0.2661
ACP.YMG14	RQRICVPRR	0.33	THP	0.81	0.44	0.835487	ACP	CpACP	ACP	CpACP	ACP	CpACP	0.908	0.453	0.4703

Chapter 5

Conclusions and Recommendations

5.1 Conclusions

In this study, a novel methodology based on network science and similarity searching was proposed and applied to explore the chemical space of THPs and discover potential THPs. Statistically, the performance of the strategy transcends current supervised ML approaches used in THPs predictions, demonstrating the potential of this alternative unsupervised approach. Hence, *in silico* predictions using the model based on representative THPs in conjunction with TumorHPD and THPep give high reliability to discover potential THPs. Herein, 54 lead compounds are repurposed as potential THPs that were obtained using the method in the starPep toolbox, followed by activity optimization using TumorHPD. In the set, novel motifs with tumor homing activity are proposed. Moreover, 54 lead molecules were subjected to punctual mutations and sequence shortening in order to find molecules with greater stability, and to enhance their tumor homing activity, identifying 27 putative THPs. In addition, a *de novo* design of ACPs was described, using evolutionary algorithms to find sequences that concentrate in tumor tissue, and have anticancer activity at the same time, where 14 ACPs were derived. The two sets of 27 THPs and 14 ACPs present a diversity structure, and would evolve the currently known chemical space of THPs.

5.2 Recommendations

This study is based on *in silico* approaches, consequently, biological assays are required to validate the tumor homing and anticancer activity. Once the activity of the peptides has been validated, it is recommended to optimize their pharmacokinetics, particularly their stability in blood, using other methodologies, such as PEGylation. On the other hand, the good performance of the methodology for predicting peptide activity based on similarity searching and network science suggests its application for the prediction of other endpoints in peptides, e.g. antimicrobial activity, toxicity, hemolytic, or anticancer.

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Attachments

A. FASTA of a set of representative 105 venom peptides obtained from starPepDB.

```
>starPep_42302
VRDAYIAKNYNCVYECFRDYSYCNDLCTKNGASSGYCQWAGKYGNACWCYALPDNVPIRVPGKCH
>starPep_26952
KKNGYAVDSSGKVAECLFNNYCNNECTKVYVYADKGYCCLLKCYCFLGADDPVLDIWDSTKNYCDVQIIDLS
>starPep_36890
RKCLIKYSQANESSKTCPSGQLLCLKKWEIGNPSGKEVKRGCVATCPKPWKNEIIQCCAADKDCNA
>starPep_35339
QAVGLPHGFCIQNRKTTWSNCSIGHRCLPYHMTCTYLYKPDENGENMKWAVKGCARMCPYAKSGERVKCTGASCNSD
>starPep_01487
KSCCPNTTGRNIYNTCRFAGGSRECAKLSGCKIISASTCPSDYPK
>starPep_11356
LVKCRGTSDCGRPCQQQTGCPNSKINRMCKCYGC
>starPep_08992
DCGHLHDPGPNDRPGHRTCCIGLQCRYGKCLVRV
>starPep_40522
SVNPCCDPVICKPRDGEHCISGPPCCNCKFLNSGTICQARARGDGNHDIYCTGITTDPCRNRYN
>starPep_17417
DCVRFWVGKCSQTSDDCPHLACKSKWPRNICVWDGVS
>starPep_10426
IPYCGQTGAECYSWCIKQDLSKDWCCDFVKDIRMNPADKCP
>starPep_24098
GTYCIELGERCPNPREGDWCCHKCVPEGKRFYCRDQ
>starPep_08211
ADDDCLPRGSKCLGENKQCKGTTMCFYANRCVGV
>starPep_20284
FRGLAKLLKIGLKSFAFVLLKVLKAAKAGKALAKSLADENAIRQQNQ
>starPep_14045
AACKCDDEGPDIRTAPLTGTVDLGSCNAGWEKASYYTHIADCCRKKK
>starPep_09101
DLWQFGKMLKVAGKLPFPYYGAYGCGYCGWGRGKPKDPTDRCCFVHDCC
>starPep_18579
EDPLYCQAIGCPTLYSEANLAVSKECRDQGKLGDDFHRCCCEEQCGSTTPASA
>starPep_09273
EPDEICRARMTHKEFNYSNVCNGCGDQVAACEAECFRNDVYTACHEAQK
>starPep_04906
ACVGDGQRCASWSGPYCCDGYCSCRSMPCRCRNNNS
>starPep_28739
LKCYPQHGKVVTCRDMKFCYHNTGMPFRNLKLLQGCSSSCSETENNKCCSTDRCNK
>starPep_32830
MNSSKLIRMLEEDGWRLVRVTGSHHHFKHPKPKGLVTVPHPKKDLPIGTVKSQKSAGL
>starPep_03482
MKLQNTLILIGCLFLMGAMIGDAYSRCQLQGFNCVVRSYGLPTIPCCRGLTCRSYFPGSTYGRQRY
>starPep_42730
VVIGQRCYRSPDCYSACKKLVGKATGKCTNGRDC
>starPep_09985
GPSFCKADEKPCYHADCCNCLSGICAPSTNWILPGCSTSSFFKI
>starPep_41131
TPFAIKCATDADCSRKCPGNPPCRNGFACT
>starPep_09189
ECLGFGKGCNPSNDQCKSSNLVCSRKHRWCKYEI
>starPep_09703
GCMKEYCAGQCRGKVSQDYCLKHKCKIPR
>starPep_27368
KNRPTFCNLLPETGRCNALIPAFYNSHLHKCQKFNYYGGCGGNANFKTIDECQRTCAAKYGRSS
>starPep_09902
GLIHKVTKVQLCAFNQDMAGWCEKSCQAAEGKNGYCHGTCKCKGKPLSYRRK
>starPep_01641
ADDKNPLEEFRETNYEVFLEIAKNGLKATSNPKRVVIVGAGMAGLSAAY
>starPep_24157
GVIPKKIWETVCPVPEWAKKCSGDIATYIKRECGKL
>starPep_36339
RDGYPLASNGCKFGCSGLGENNPTCNHVCEKAGSDYGYCYAWTCYCEHVAEGTVLWGDSTGPCRS
>starPep_01371
GKFSVFSKILRSIAKVFVGKVKRQFKTASDLKKNQ
>starPep_16015
CAKKRNWCGKNEDECCPMKCIYAWYNQCGSCQTITGLFKKC
>starPep_24272
GWCGDPGATCGKLRLYCCSGFDCYTKTCKDKSSA
>starPep_17657
DFPLSKEYESCVRPRKCKPPLKCNKAQICVDPNKGW
>starPep_15767
AVITGACERDLQCGKGTCCAVSLWIKSVRVCTPVGTSGEDCHPASHKIPFSGQRMHHTCPCAPNLACVQTSPPKFKLSKS
```

A. (Cont.) FASTA of a set of representative 105 venom peptides obtained from starPepDB.

```
>starPep_22698
GKRPRPVMCQCVDTTNGGVRLDAVTRAACSIDSFIDGYYTEKDGFCAKYSWDLFTSGQFYACLRYSHAGTNCQDPDQYE
>starPep_13668
WLGCAVKEACGPWEWPCCSGLKCDGSECHPQ
>starPep_20467
FVQHRPRDCESINGVCRHKDTVNCREIFLADCYNDEQKCCRK
>starPep_09596
FPRPRICNLACRAGIGHKYPFCHCR
>starPep_06936
MKTQFAIFLITLVLFQMFSDAIFKAIWSGIKSLFGKRGSLDLDLDESDFGEVVSQADIDFLKELMQ
>starPep_40558
SWDSIWKSANKMMDKIMRQKVAKWMAKKEGKSVEEVQAKVDAMSKKDIRMHVISHYGKKAFFEQLSKSLLE
>starPep_00486
GRGREFMSNLKEKLSGVKEKMKNS
>starPep_00538
RICRRRSAGFKGPCVSNKCAQVCMQEGWGGNCDGPLRRCKMRRR
>starPep_31970
MISMLRCTFFFVSVILITSYFVPTMSIKNRKRHVIKPHICRKCCKNG
>starPep_02059
ADPTFGFTPLGLSEKANLQIMKAYD
>starPep_24080
GTTTCYCGKTIGIYWFGTKTSPNRGYTGSCGYFLGICCPVD
>starPep_09161
DVTFSLLGANTKSYAAFITNFRKDVASEKK
>starPep_33838
NCVANILNINEAVIATGCVPAAGELRIFVGSSSHLYLIKATSSCGLSLTNQVFINGESVQSGGRC
>starPep_02272
IWLITALKFLGKNLKGHLAKQLAKL
>starPep_00607
FHPSLWVLPQYIQLIRKILKSG
>starPep_11966
MTKQSIIVLFAAIAMMAQLRVTAEPAPEPIAAPAEPYANPEAIASPEAKDLHTVVSAILQALGKK
>starPep_06767
MAQDIISTIGDLVKWIHTVKNKFTKK
>starPep_40811
TDDDESGNKCAKTKRRENVCRVCGNRSNDEYYECCESDYRYHRCLDLLRN
>starPep_13871
YCQKWMWTCDEERKCEGLVCRLWCKRIINM
>starPep_00286
GFFALIPKISSLPLFKTLLS AVGSALSSSGEQE
>starPep_41864
VIHYELNLQGTTKAQYSTILKQLRDDIKDPNLXYGXXDYS
>starPep_09560
FLPLLLGSLMTPPVIQAIHDAQR
>starPep_02364
PNPKVFFDMTIGGQSAGRIVMEEYA
>starPep_05690
GLKDWWNKHKDKIVEVVKDSGKAGLNAA
>starPep_05884
HGEGTFTSDLSKQMEEEAVRLFIEWLKNKGPPSSGAPPPS
>starPep_35905
QPQSHSIELDEVSKEAASTRAALTSNL
>starPep_13368
VAVKATTTTEETEIPAK
>starPep_32019
MKDLMSLVIAPIFVGLVLEMISRVLDEEDSRK
>starPep_09722
GEEELQENQELIREKSN
>starPep_10044
GSPRTEYEACRVRCQVAEHGVERQRRCCQVCEKRLREREGRE
>starPep_09011
DDRRSPLEECFQNDYEEFLEIARNSQLYQESLREDSSYHLSFIESLKSALFSYEKKFWEADGIHGGKVINDSLHDLPKREIQALCYPYSIKK
>starPep_01642
ADDRNPLEQCFRETDYEEFLEIARNNLKATSNPKHVIVGAGMAGLSAAYVLSGGGHQVTV
>starPep_01891
KVCRRRSAGFKGPCVSDKNCAQVCLQEGWGGNCDGPFRRCKCIRQC
>starPep_03728
AAPCFCSGKPGRGDLWILRGTCPPGGYGYTSNICYKWPNICYPH
>starPep_03969
FVQHRPRDCESINGVCRHKDTVNCREIFLADCYNDGQKCCRK
>starPep_08260
AEKDCIAPGAPCFGTDKPCCNPAWCCSSYANKCL
>starPep_09697
GCGGLMAGCDGKSTFCCSGYNCSPTWKVWCYARP
>starPep_09702
GCLGEGEKCADWSGPSCCDGFYCSRSMPYCRCRNNNS
>starPep_11811
MKTQFAILLVALVLFQMQFAQSDAILGKIWEGIKSLFGKRGSLDLDGLDELFDGEISKADRDFLRELMR
>starPep_14332
ADCNGACSPFVPPCRSRDRCVPIGLFVGFCHPTG
>starPep_16975
CSCNDINDKCEMYFCHQDVIWDEP
>starPep_18084
DRDSCVDKSRCAKYGYQECCQDCCKNAGHNGGTCMFFKCKCA
>starPep_18467
ECLEIFKACNPSNDQCCKSSKLVCSRKTRWCKYQI
```

A. (Cont.) FASTA of a set of representative 105 venom peptides obtained from starPepDB.

```
>starPep_11744
MKFLVNVVALVYFGRVHFLHLCVHFLHLWAPEPEPAPEAEAEADPEAGIGAVLKVLTTGLPALISWIKRKRQQG
>starPep_16979
CSCTDMSDLECMNFCHKDVIWNRN
>starPep_13928
YKQCHKKGGHCFPKKIKIPSSDLGKMDCRWVKWCKCKKGS
>starPep_39618
SGPADCCRMKECCTDRVNECLQRYSGREDKVFVFCYQEAATVTCGSFNEIVGCCYGYQCMIRVVKPNSLSGAHEACKTVSCGNPCA
>starPep_21163
GEATTIWGVGADEAIDKGTPSKNDLQNSADLAKNGFKGHQGVACSTVVKDGNKDVYMIKFSLAGGSNDPGGSPCSDD
>starPep_09979
GPMRIPEKHRIVREYIRKFLQLNEFVQETENAWYYIKNIRKVKHEVKKDPGLLKYPVKP
>starPep_32131
MKISQVFIFVFLLMISVAWANEAYEEESNYLSERFADVEEITPEFRGIRCPKSWKCKAFKQRLKRLLAMLRQHAF
>starPep_23891
GSCVPVDQPCLNTQPCCDDATCTQERNENGHTVYYCRA
>starPep_24192
GVPCRCSDGSPVHGNTLSGTVWVGSCASGWHKCNDEYNLAYECKE
>starPep_25916
ISIDPPCRFCYHRDGSNCVYDAYGCGAV
>starPep_29538
LTCVKNSNIWFPTSEDCPDGQNLCKFRWQYISPRMYDFTRGCAATCPKAEYRDVINCCGTDKCNK
>starPep_34640
NSVNPCCDPQTCKPIEGKHCISGPECENCYFLRSGTICQRARGDGNNDYCTGITPDCPRNRYN
>starPep_36661
RICYSHKASLPRATKTCVENTCYKMFIRTHRQYISERGGCPTAMWPYQTECKGDRCNK
>starPep_37544
RPTDIKCESYQCFPVCKSRFGKTNRCVNGFCDCF
>starPep_39452
SECVENGGFCDPEKMGDWCCGRICRNECRNG
>starPep_41191
TPYPVNCKTDRDCVMCGLGISCKNGYCGGCT
>starPep_44620
YKQCHKKGGHCFPKKIKIPSSDFGKMDCRWRWCKCKKRSKG
>starPep_32546
MKYFVIALALAVLVCAESTAYEVNEELENELDDLDAAWLAVAEELQGLEDFEESRGLFGKLIKFKGRKAISYAVKARGKN
>starPep_14712
AKACTPLLHDCSHDRHSCCRGDMFKYVCDCFYPEGEDKTEVCSCQQPKSHKIAEKIIDKAKTTL
>starPep_32810
MNSKIFAVLLLLAFLSCLSDQYCPKSSITACKKMIRNDCCCKDDDDCTGGSWCCATPCGNFCKYPTDRPGGKRAAGGKCKTGYVY
>starPep_28501
LFECFSCEIEKEGDKPKKKKCKGGWKCKFNMCVKV
>starPep_06838
MGAALKMTIFLLIVACAMIATTEAAVRIGPCDQVCPRIVPERHECCRAHGRSGYAYCSGGMYCN
>starPep_02749
MEKIANAVKSAIEAGQNQDWTKLGTSLDIVSNGVTELSKIFGF
>starPep_35451
QEDGEIVCGEDDPCGTQICECDKAAAICFRNSMDT
>starPep_00644
FSFKRLKGFAGKLNKSLARKIRTKGLKYVKNFAKDMLESEGEAPPAAEPPVEAPQ
>starPep_02421
VFHAYSARGVRNNYKSAVGPADWVISAIRGFIHG
```

B. FASTA of a set of representative 162 ACPs obtained from starPepDB.

```
>starPep_05497
GFKDLLKGAALKKTVLF
>starPep_05855
GWRKWIKKATHVKGKHIGKAALDAYI
>starPep_03042
FLGALFKVASKVLPSVKCAITKKC
>starPep_00126
GLFGKLIKFKGRKAISYAVKKARGKH
>starPep_06208
KILRGVSKKIMRTFLRRISKDILTGGK
>starPep_09845
GIPCGESCVPICLTSAIGCCKSKVCYRN
>starPep_11176
LKC�KLVPLFYKTCPAGKNL
>starPep_18164
DTAVTGLASPLSTGKILDQKAYSCANRLIVLCIENSFMTDARK
>starPep_24426
GYNYAKKLANLAKKFANALW
>starPep_09764
GFWSSVWDGAKNVGTAIKNAKVCVYAVCVSHK
>starPep_03287
IKIPSFERNILKKVKGKEAVSLIAGALKQS
>starPep_00640
FLSLIPHIVSGVASIAKHF
>starPep_00315
GLFDIVKKIAGHIASSI
>starPep_32958
MQFITDLIKKAVDVKGLFGNK
>starPep_00361
KWKVFKKIEKMRNIRNGIVKAGPAIAVLGEAKAL
>starPep_24256
GVWGIAGIAGKVLGNILPHVFSSNQS
>starPep_00657
GFLGILFHGVHHGRKKALHMNSERRS
>starPep_00807
KSSAYSLQMGATAIKQVKKLFKKWGW
>starPep_22576
GKEFKRIVWLSKTAKKL
>starPep_18824
EKSSRPEFYKVILGAHEEYIRG
>starPep_27320
KNECLWTDMLSNFGYPGYQSKHYACIRQKG
>starPep_14350
ADMDFTGIAESIIKKIKETNAKPPA
>starPep_12134
PAWRKAFRWAARMLKAA
>starPep_36015
QRTEIIHRALYDLIS
>starPep_13195
TFRAFLSSRLQDLYSIVRRADRAAV
>starPep_21198
GEILCNLCTGLINTLENLLTTKRKRQQ
>starPep_34474
NPEKALEKLIAIQKAIKMLNGWFTGVGFRRKR
>starPep_10217
HLRRINKLLTRIGLYRHAFG
>starPep_12142
PDEDAINNALNKVCSTGRRQRSICKQLLKK
>starPep_24842
HTHQDFQPVLHLVALNTPLSGGMGRGIR
>starPep_18008
DPFFKVPVNKLAAVSNFGYDLYRVRSSMSPPTN
>starPep_07864
VLLVTLTRLHQRGVYIRKWRHFSGRKYR
>starPep_41343
TTITGKKCQSWAAMPHRHST
>starPep_07104
MWKEFHNVLSGQLLADKRWARWYNRW
>starPep_07120
NLVSALIEGRKYLKNVLKKNLRLKEKNKAKNSKENN
>starPep_07882
VNWKKXLGKXIKXVK
>starPep_10591
KIAKVALAKLGIGAVLKVLTG
>starPep_11276
LPRNRWWSKIWKVTVFS
>starPep_12079
NRFTARFRTPWRLCLQFRQ
>starPep_13296
TRWLWLLRGGLKAAGWGIRAHLRNQ
```

B. (Cont.) FASTA of a set of representative 162 ACPs obtained from starPepDB.

```
>starPep_02535
FLHHIVGLIHHGLSLFGDRAD
>starPep_00758
GWKKWFTKGERLSQRHFA
>starPep_00419
FLGALIKGAIHGGRFIHGMIQNH
>starPep_09994
GQVWEATATVNAIRGSVTPAVSQFNARTAD
>starPep_35821
QMIVIELGTNPLKSSGIENGAFQGMK
>starPep_00249
ACGILHDNCVYVPAQNPCCRGLQCRYGKCLVQV
>starPep_26183
KAKAKAVSR SARAGLQFPVGRHRHLK
>starPep_36692
RIIDLWRVWRPQKPKFVTVVWR
>starPep_41266
TRSRWRRFIRGAGRFARRYGWRIA
>starPep_02289
KRKCPKTPFDNTPGAWFAHLILGC
>starPep_14847
ALARQPLTGSPPNERAFFCSSLRR
>starPep_00719
GLLSVLGSSVVKHVIPHVVPVIAEHL
>starPep_14190
AAPFLECQGRQGTCHFFAN
>starPep_03327
KKCKFFCKVKKKIKSIGFQIPIVSIPFK
>starPep_41021
TKWTPCSRTC GMGISNRV
>starPep_23911
GSGSGSGLKIKFKPMVIGVTIPF
>starPep_19944
FLKDHRISTFKNWPF
>starPep_02569
GFKRIVQRIKDFLRNLV
>starPep_01434
GTGLPMSERRKIMLMMR
>starPep_10463
ITCPQVTQSLAPCVPYLSG
>starPep_11901
MRGIRGADFQAFQARAVGLAGTFR
>starPep_04365
LALERRSGWLRFLGLKPRRKH
>starPep_13155
TAGIKLTVPIEKFPVTTQTFWG
>starPep_30757
MFSPILSLEHLALATLQSVFAQPVICTTVGSAAEGS
>starPep_00686
GKGRWLERIGKAGGIIIGALDHL
>starPep_15907
AWYRGAAPPKQEFLDIEDP
>starPep_12249
PRFWEYALRLME
>starPep_03733
ACVNQCPDAIDRFIVKDKGCHGVEKKYYKQVYVACMNGQHLYCRTEWGGPCQL
>starPep_35471
QEPHRHSIFTPQTNPRADLEKN
>starPep_36506
RGFTKMPHVQIHTEASESL
>starPep_04324
KRMGIFHLFWAGLRKLGNIKNKIQQGIENFLG
>starPep_01246
ATPATPTVAQFVIQGSTICLVC
>starPep_29515
LSSTCILVLVKDILVLVKEILVLVVKDKPI
>starPep_40880
TFKRKNGSRKNGHRPGGYSLIALGNKKVLKAPYMESI
>starPep_26006
ITMQGIQKQKIRMIMF
>starPep_25650
IMRIKQGQIGQMTI
>starPep_00911
ANDPQCLYGNVAAKF
>starPep_26645
KIKSCYYLPCFVTS
>starPep_18575
EDMNQKLFDLRGKFKRPLRRVRMSADAML
>starPep_02982
CVLIGQRCDNRGPRCCSGQGNVPLPFLGGVCAV
>starPep_02368
QICKAPSQTFFGLCFMDSSCRKYCIKEKFTGGHCSKLQRKCLCTKPC
>starPep_27510
KRFKQDGGWSHSPWSSC
```

B. (Cont.) FASTA of a set of representative 162 ACPs obtained from starPepDB.

```
>starPep_41295
TSLDASIIWAMMQN
>starPep_43002
WGRAFSAGVHRLANGGNG
>starPep_16327
CDSDSITWDQLWDLMK
>starPep_00564
YRGGYTGPPIRPPPIGRPPFRPVCNACYRLSVSDARNCCIKFGSCCHLVK
>starPep_32937
MPTWAWWFLVLLALWAPARG
>starPep_07884
VNWXXILGXIXVVX
>starPep_29189
LPGLTGSKGVRGISGLPGFSG
>starPep_24115
GVDITVIRPNH
>starPep_21830
GFHDHGPCDPPSHK
>starPep_05790
GRKKRRQRRRGGMWVVTNLRTD
>starPep_16457
CGGYSGWHRLRSTSYRCG
>starPep_13094
STRUCTUREGIVEN
>starPep_04732
RWFKIQMQRIRWKNKK
>starPep_08575
AXQNMEILEXTPLTXVX
>starPep_42725
VVGSPSAQDEASPL
>starPep_22164
GHRATSDLASTGEESQD
>starPep_16341
CELDENNTPMC
>starPep_40547
SVSRAGSPSGGFC
>starPep_41784
VGTDFSGNDDISDVQK
>starPep_37972
RRPKGRAMRREKQRPSDKPRR
>starPep_33129
MRLVLSSLLCILLCFISFSTEGKRRPAKAWSGRRTRLCCHRVSPNSTNLKGHHVRLC
KPKLEPEPRLWVVPALPQV
>starPep_44212
XXLIXVWAXGFXAXXLFXGIG
>starPep_43649
WYTXXTWXWXY
>starPep_43904
XIXIILPPLPII
>starPep_09828
GIIKKIHKIHKIHKIHKI
>starPep_28706
LIXFXPX
>starPep_09658
FXYWKXT
>starPep_00089
SWLSKTAKKLENSAKKRRISEGIAIAIQGGPR
>starPep_00224
RIIDLWRVRRPQKPKFVTWVVR
>starPep_00260
DTHFPICIFCCGCCHRKCGMCKT
>starPep_00775
ILGPVLGLVSDTLDDVLGIL
>starPep_03342
KLKNFAKVAQSLLNKASCKLSGQC
>starPep_03814
CETPSKHFNGLCIRSSNCASVCHGEHFTDGRQCQVRRRCMCLKPC
>starPep_03965
FVKKLILNIINSIFKK
>starPep_04262
KKALKHALAKWLPALKALAHKLAKK
>starPep_04575
NFAEIFAAVNKLIKQGVVKG
>starPep_04632
QRSVSNAATRVSRTRGRSRWRDVSRRNFMR
>starPep_05117
CHTNGGYCVRAICPPSARRPGSCFPEKNPCKYK
>starPep_06981
MPKEKVFLKIEKMGRNIRN
```


B. (Cont.) FASTA of a set of representative 162 ACPs obtained from starPepDB.

```
>starPep_16391
CGESCVFIPCISVIGCACKSKVCYKNGSIP
>starPep_18108
DRSTREPIYMSTI
>starPep_18157
DSSPVSTEQ LAPTA
>starPep_19548
FFSLIPKLVKGLISAFK
>starPep_20122
FLSLIPAAISAVSALANHF
>starPep_24834
HTASDA A A A A A A L T A A N A A A A A A A S M A
>starPep_39171
SAPFIECHGRGTCNYYANS
>starPep_41038
TLPFAYCNIHQVCHYAQRNDRSYWL
>starPep_24565
HGLGHCHEQQHGLGHGHKFKLDDDEHQGGHVLD
>starPep_30471
MDSNKDERAYAQWVHILHNVGSSPFKIANLGLSWGKLYADGNKDKEVYP
>starPep_21042
GCRRLCWKQRCVTYCRGR
>starPep_41838
VIFEWTL L Q V L S E S D Q D Q S L E V F L T
>starPep_09784
GGVCPKILKKRRSDCPGACICRGNGYCGSGSD
>starPep_04761
SKWQHQQDSCRKQLQGVNLTPEKHIMEKIQGRGDDDDDDDDDD
>starPep_23735
GRFKRFRK K L K R L W H K V G P F V G P I L H Y
>starPep_18139
DSEGWKVQP N I R D Q D G N T A G S V R V Q K Q L G N H E V H A G A S R V F S G P N R G G P S Y N V G A T F N V
>starPep_01120
ITSISLCTPGCKTGALMGCNMKTATCNC SIH VSK
>starPep_26494
KGIRGYKGGYCKGAFKQTCKCY
>starPep_10026
GSEGPLKPGARIFSDGKDVLRHPT
>starPep_39822
SKRKS R P V S V K T F E D I P L E E P
>starPep_33979
NGREACLDPEAPMVQKIVQKMLKG
>starPep_41197
TQQAFQKFLAAVTSALGKQYH
>starPep_05425
FSPQMLQDIHEKTKIL
>starPep_21918
GFRKRFNKLVKKVKHTIKETANVSKDVAIVAGSGVAVGAAM
>starPep_29425
LRSRGELVAKFLAGEQSPEDYVAE
>starPep_18829
EKYEGKISKTMGLDCQAWDS
>starPep_28608
LHCPALVTYNTDTFESMPNPEGRYTFGASCV
>starPep_00270
FLIGMTQGLICLITRKC
>starPep_17955
DLWIRETLTSPKSLTG
>starPep_00021
ACYCRIPACIAGERRYGT CIYQGR LWAFCC
>starPep_02292
KSCCPNTTGRNIYNTCRLTGSSRETCAKLSGCKIISASTCPSNYPK
>starPep_00334
GLMSSIGKALGLLVDVLKPKTPAS
>starPep_26158
IYFDGRDIMTDPSPWQKVIWHGSSPHGVRLVDNYCEAWRTA
>starPep_29236
LPRFSTMPFIYCNINEVCHY
>starPep_03897
FCTCNVKGFN AKNKRGIIP
>starPep_01521
MRKEFHNVLSGQLLADKRPARDYNRK
>starPep_08214
ADDKNPLEECFRET D Y E E F L E I A R N G L K A T S N P K R V V
>starPep_34689
NVLSPLSVATALSALSLGAEQRTES
>starPep_15086
ANIKLSVQMKLFRHLKWKIIVKLNDRGRELSDA
>starPep_40840
TEENRELVSELKRP
>starPep_00124
GKPRPYSPRPTSHPRPIRV
```

C. CSN parameters of similarity threshold analysis.

Similarity threshold	Density	Communities	Modularity	Singletons	ACC
0.1	0.999	3	0.03	0	0.999
0.15	0.996	3	0.03	0	0.996
0.2	0.985	3	0.03	0	0.988
0.25	0.956	3	0.04	0	0.968
0.3	0.891	3	0.05	0	0.93
0.35	0.772	3	0.07	0	0.87
0.4	0.593	3	0.11	0	0.791
0.45	0.383	4	0.16	2	0.703
0.5	0.199	4	0.23	3	0.612
0.55	0.079	6	0.34	20	0.508
0.6	0.023	10	0.47	99	0.428
0.65	0.005	34	0.68	238	0.419
0.7	0.001	38	0.81	449	0.544
0.75	0	21	0.85	548	0.535
0.8	0	13	0.84	587	0.456
0.85	0	9	0.87	606	0.456
0.9	0	2	0.5	623	-

D. Output from Friedman test where 9 best SSMs were compared.

Output tables for 1xN statistical comparisons.

November 20, 2021

1 Average rankings of Friedman test

Average ranks obtained by each method in the Friedman test.

Algorithm	Ranking
CSN-TH-0.60Sc-467-H+s-0.40-578	6.9583
CSN-TH-0.60Sc-467-H+s-0.50-575	5.5833
CSN-TH-0.60Sc-467-H+s-0.60-571	5.375
CSN-TH-0.60Sc-469-W+s-0.40-579	6.7708
CSN-TH-0.60Sc-469-W+s-0.50-576	5.5833
CSN-TH-0.60Sc-469-W+s-0.60-573	5.0625
CSN-TH-0.60Sc-479-H+W+s-0.4-589	4.1667
CSN-TH-0.60Sc-479-H+W+s-0.5-586	3.1042
CSN-TH-0.60Sc-479-H+W+s-0.6-583	2.3958

Table 1: Average Rankings of the algorithms (Friedman)

Friedman statistic (distributed according to chi-square with 8 degrees of freedom): 60.372222.

P-value computed by Friedman Test: 0.

Iman and Davenport statistic (distributed according to F-distribution with 8 and 184 degrees of freedom): 10.54915.
P-value computed by Iman and Davenport Test: 0.000000000004.

2 Post hoc comparison (Friedman)

P-values obtained in by applying post hoc methods over the results of Friedman procedure.

i	algorithm	$z = (R_0 - R_i)/SE$	p	Holm	Hochberg	Hommel	Holland
8	CSN-TH-0.60Sc-467-H+s-0.40-578	5.771157	0	0.00625	0.006391		
7	CSN-TH-0.60Sc-469-W+s-0.40-579	5.533986	0	0.007143	0.007301		
6	CSN-TH-0.60Sc-467-H+s-0.50-575	4.031904	0.000055	0.008333	0.008512		
5	CSN-TH-0.60Sc-469-W+s-0.50-576	4.031904	0.000055	0.01	0.010206		
4	CSN-TH-0.60Sc-467-H+s-0.60-571	3.768381	0.000164	0.0125	0.012741		
3	CSN-TH-0.60Sc-469-W+s-0.60-573	3.373096	0.000743	0.016667	0.016952		
2	CSN-TH-0.60Sc-479-H+W+s-0.4-589	2.239947	0.025094	0.025	0.025321		
1	CSN-TH-0.60Sc-479-H+W+s-0.5-586	0.895979	0.370264	0.05	0.05		

Table 2: Post Hoc comparison Table for $\alpha = 0.05$ (FRIEDMAN)

Bonferroni-Dunn's procedure rejects those hypotheses that have a p-value ≤ 0.00625 .

Holm's procedure rejects those hypotheses that have a p-value ≤ 0.025 .

Hochberg's procedure rejects those hypotheses that have a p-value ≤ 0.016667 .

Hommel's procedure rejects those hypotheses that have a p-value ≤ 0.025 .

Holland's procedure rejects those hypotheses that have a p-value ≤ 0.05 .

3 Adjusted P-Values (Friedman)

Adjusted P-values obtained through the application of the post hoc methods (Friedman).

i	algorithm	unadjusted p	p_{Bonf}	p_{Holm}	$p_{Hochberg}$	p_{Hommel}
1	CSN-TH-0.60Sc-467-H+s-0.40-578	0	0	0	0	0
2	CSN-TH-0.60Sc-469-W+s-0.40-579	0	0	0	0	0
3	CSN-TH-0.60Sc-467-H+s-0.50-575	0.000055	0.000443	0.000332	0.000277	0.000277
4	CSN-TH-0.60Sc-469-W+s-0.50-576	0.000055	0.000443	0.000332	0.000277	0.000277
5	CSN-TH-0.60Sc-467-H+s-0.60-571	0.000164	0.001314	0.000657	0.000657	0.000657
6	CSN-TH-0.60Sc-469-W+s-0.60-573	0.000743	0.005946	0.00223	0.00223	0.00223
7	CSN-TH-0.60Sc-479-H+W+s-0.4-589	0.025094	0.200755	0.050189	0.050189	0.050189
8	CSN-TH-0.60Sc-479-H+W+s-0.5-586	0.370264	2.962113	0.370264	0.370264	0.370264

Table 3: Adjusted p -values (FRIEDMAN) (I)

i	algorithm	unadjusted p	$p_{Holland}$
1	CSN-TH-0.60Sc-467-H+s-0.40-578	0	0
2	CSN-TH-0.60Sc-469-W+s-0.40-579	0	0
3	CSN-TH-0.60Sc-467-H+s-0.50-575	0.000055	0.000332
4	CSN-TH-0.60Sc-469-W+s-0.50-576	0.000055	0.000332
5	CSN-TH-0.60Sc-467-H+s-0.60-571	0.000164	0.000657
6	CSN-TH-0.60Sc-469-W+s-0.60-573	0.000743	0.002228
7	CSN-TH-0.60Sc-479-H+W+s-0.4-589	0.025094	0.049559
8	CSN-TH-0.60Sc-479-H+W+s-0.5-586	0.370264	0.370264

Table 4: Adjusted p -values (FRIEDMAN) (II)

E. Confusion matrices of the best SSM THP1 against Main, Small, and Main90 datasets.

Confusion Matrix		THP1 Main Dataset				
		Percent	Positive	Negative	Total	
Precision -> Active	Positive	99.66	581	2	583	
Precision -> Inactiv	Negative	90.26	70	649	719	
Ac%	Total	94.47	651	651	1302	
THP1						
		Recall -> Active		Precision -> Active		
		MCC	Ac (Accuracy)	Especificidad	Sensibilidad	FAR%
		0.894	94.47	89.25	99.66	9.74

Confusion Matrix		THP1 Small				
		Percent	Positive	Negative	Total	
Precision -> Active	Positive	99.50	402	2	404	
Precision -> Inactiv	Negative	87.45	67	467	534	
	Total	92.64	469	469	938	
THP1						
		Recall -> Active		Precision -> Active		
		MCC	Ac (Accuracy)	Especificidad	Sensibilidad	FAR%
		0.861	92.64	85.71	99.50	12.55

Confusion Matrix		THP1 Main90				
		Percent	Positive	Negative	Total	
Precision -> Active	Positive	98.86	173	2	175	
Precision -> Inactiv	Negative	99.31	3	431	434	
	Total	99.18	176	433	609	
THP1						
		Recall -> Active		Precision -> Active		
		MCC	Ac (Accuracy)	Especificidad	Sensibilidad	FAR%
		0.980	99.18	98.30	98.86	0.69

F. Output from Friedman test where THP1 was compared with literature models.

Output tables for 1xN statistical comparisons.

November 20, 2021

1 Average rankings of Friedman test

Average ranks obtained by each method in the Friedman test.

Algorithm	Ranking
TumorHPD	2.5833
THPep	2.1667
THP1	1.25

Table 1: Average Rankings of the algorithms (Friedman)

Friedman statistic (distributed according to chi-square with 2 degrees of freedom): 11.166667.
P-value computed by Friedman Test: 0.00376.

Iman and Davenport statistic (distributed according to F-distribution with 2 and 22 degrees of freedom): 9.571429.
P-value computed by Iman and Davenport Test: 0.001021905094.

2 Post hoc comparison (Friedman)

P-values obtained in by applying post hoc methods over the results of Friedman procedure.

i	algorithm	$z = (R_0 - R_i)/SE$	p	Holm	Hochberg	Hommel	Holland
2	TumorHPD	3.265986	0.001091	0.025	0.025321		
1	THPep	2.245366	0.024745	0.05	0.05		

Table 2: Post Hoc comparison Table for $\alpha = 0.05$ (FRIEDMAN)

Bonferroni-Dunn's procedure rejects those hypotheses that have a p-value ≤ 0.025 .
Hochberg's procedure rejects those hypotheses that have a p-value ≤ 0.05 .
Hommel's procedure rejects all hypotheses.

3 Adjusted P-Values (Friedman)

Adjusted P-values obtained through the application of the post hoc methods (Friedman).

i	algorithm	unadjusted <i>p</i>	<i>p</i> _{Bonf}	<i>p</i> _{Holm}	<i>p</i> _{Hochberg}	<i>p</i> _{Hommel}
1	TumorHPD	0.001091	0.002182	0.002182	0.002182	0.002182
2	THPep	0.024745	0.049489	0.024745	0.024745	0.024745

Table 3: Adjusted *p*-values (FRIEDMAN) (I)

i	algorithm	unadjusted <i>p</i>	<i>p</i> _{Holland}
1	TumorHPD	0.001091	0.00218
2	THPep	0.024745	0.024745

Table 4: Adjusted *p*-values (FRIEDMAN) (II)

G. Predicted activities of 43 repurposed peptides obtained from hierarchical virtual screening of peptides from starPepDB.*Originally, these peptides contained a X aa, which was changed by an aa that gave them greater tumor homing potential.

ID	Sequence	TumorHPD		THPep	AntiCP			ToxinPred							
		SVM Score			SVM Score		CellPPD	SVM Score	SVM Score	SVM Score	SVM Score				
starPep_27924	KWCFRVAIRGISYRRRCR	0.62	THP	THP	1	Anticip	CPP	-1.45	Non-Toxin	-1.45	Non-Toxin	-0.73	Non-Toxin	-0.73	Non-Toxin
starPep_43589	WWWKNGKKNKNGKH	0.98	THP	THP	0.56	Anticip	CPP	-0.32	Non-Toxin	-0.32	Non-Toxin	-0.04	Non-Toxin	-0.04	Non-Toxin
starPep_24644	HKHGHLKHKKLNKNGKH	0.37	THP	THP	0.38	Anticip	CPP	-0.97	Non-Toxin	-0.97	Non-Toxin	-0.98	Non-Toxin	-0.98	Non-Toxin
starPep_02029	TPFKLSLHL	0.33	THP	THP	0.17	Anticip	Non-CPP	-1.02	Non-Toxin	-1.02	Non-Toxin	-1.29	Non-Toxin	-1.29	Non-Toxin
starPep_07234	QGRLLGTQWAVGHLM	0	THP	THP	-0.29	Non-Anticip	Non-CPP	-1.48	Non-Toxin	-1.48	Non-Toxin	-1.21	Non-Toxin	-1.21	Non-Toxin
starPep_43502	WWAMKWRIV	1.58	THP	THP	1.15	Anticip	CPP	-0.49	Non-Toxin	-0.49	Non-Toxin	-0.81	Non-Toxin	-0.81	Non-Toxin
starPep_13108	SVSWGKPKSPRQ	0.6	THP	THP	-0.46	Non-Anticip	Non-CPP	-1.4	Non-Toxin	-1.4	Non-Toxin	-1.76	Non-Toxin	-1.76	Non-Toxin
starPep_27446	KQCISLKGICKDLACT	0.38	THP	THP	1.8	Anticip	Non-CPP	-0.55	Non-Toxin	-0.55	Non-Toxin	-0.02	Non-Toxin	-0.02	Non-Toxin
starPep_27346	KNKGKQWWW	1.04	THP	THP	0.69	Anticip	CPP	-0.6	Non-Toxin	-0.6	Non-Toxin	-0.44	Non-Toxin	-0.44	Non-Toxin
starPep_26052	IVLVRWPK	-0.33	Non-THP	THP	0.54	Anticip	CPP	-0.9	Non-Toxin	-0.9	Non-Toxin	-0.99	Non-Toxin	-0.99	Non-Toxin
starPep_14535	AGIRRPFGFSPRLIA	0.46	THP	THP	0.15	Anticip	Non-CPP	-0.81	Non-Toxin	-0.81	Non-Toxin	-0.36	Non-Toxin	-0.36	Non-Toxin
starPep_16575	CKGGAKAARSQK	1.07	THP	THP	1.81	Anticip	CPP	1	Toxin	1	Toxin	1	Toxin	1	Toxin
starPep_10105	GWAGWLLSPRGRPSWGP	2.2	THP	THP	0.86	Anticip	CPP	-1.03	Non-Toxin	-1.03	Non-Toxin	-1.05	Non-Toxin	-1.05	Non-Toxin
starPep_35988	QRNGLRHH	-0.37	Non-THP	THP	0.33	Anticip	CPP	-0.55	Non-Toxin	-0.55	Non-Toxin	-0.43	Non-Toxin	-0.43	Non-Toxin
starPep_10014	GRRSTHWRI	0.64	THP	THP	-0.03	Non-Anticip	CPP	-0.92	Non-Toxin	-0.92	Non-Toxin	-1.51	Non-Toxin	-1.51	Non-Toxin
starPep_07641	RSQMQDGLQSCCQELQNVEEQCQC	0.28	THP	THP	-1.03	Non-Anticip	Non-CPP	0.24	Toxin	0.24	Toxin	-0.37	Non-Toxin	-0.37	Non-Toxin
starPep_18023	DPFSNSWG	0.51	THP	THP	-0.47	Non-Anticip	Non-CPP	-1.09	Non-Toxin	-1.09	Non-Toxin	-0.99	Non-Toxin	-0.99	Non-Toxin
starPep_25472	ILPVKWWPWVWRR	2.12	THP	THP	1.16	Anticip	CPP	-0.56	Non-Toxin	-0.56	Non-Toxin	-0.57	Non-Toxin	-0.57	Non-Toxin
starPep_43120	WKGHWYKTT	0.71	THP	THP	0.5	Anticip	CPP	-0.05	Non-Toxin	-0.05	Non-Toxin	-0.39	Non-Toxin	-0.39	Non-Toxin
starPep_13030	SPRGRSPSWGTPDPRRS	1.04	THP	THP	-0.1	Non-Anticip	CPP	-1.29	Non-Toxin	-1.29	Non-Toxin	-1.72	Non-Toxin	-1.72	Non-Toxin
starPep_04689	RLRLRGR	1.14	THP	THP	-0.04	Non-Anticip	CPP	-1.22	Non-Toxin	-1.22	Non-Toxin	-0.86	Non-Toxin	-0.86	Non-Toxin
starPep_15346	AQPSFAF	0.67	THP	THP	-0.3	Non-Anticip	Non-CPP	-1.2	Non-Toxin	-1.2	Non-Toxin	-0.88	Non-Toxin	-0.88	Non-Toxin
starPep_05157	DGPKKKKKKSPSKSSG	-0.19	Non-THP	non-THP	-0.12	Non-Anticip	CPP	-0.97	Non-Toxin	-0.97	Non-Toxin	-0.92	Non-Toxin	-0.92	Non-Toxin
starPep_07335	RCICRLGIC	2.77	THP	THP	1.57	Anticip	CPP	-0.65	Non-Toxin	-0.65	Non-Toxin	-0.3	Non-Toxin	-0.3	Non-Toxin
starPep_08545	AVESTVATLEASPEVIESPPE	-1.71	Non-THP	non-THP	-1.27	Non-Anticip	Non-CPP	-0.96	Non-Toxin	-0.96	Non-Toxin	-1.07	Non-Toxin	-1.07	Non-Toxin
starPep_41900	VIVRWRFY	0.36	THP	THP	1.2	Anticip	CPP	-0.58	Non-Toxin	-0.58	Non-Toxin	-0.86	Non-Toxin	-0.86	Non-Toxin
starPep_05293	FFRNLRWKGAKAFAFRAGHAAWRA	0.07	THP	THP	0.35	Anticip	CPP	-0.74	Non-Toxin	-0.74	Non-Toxin	-1.28	Non-Toxin	-1.28	Non-Toxin
starPep_36476	RFWVRGRRS	0.66	THP	THP	0.11	Anticip	CPP	-0.98	Non-Toxin	-0.98	Non-Toxin	-1.43	Non-Toxin	-1.43	Non-Toxin
starPep_17042*	VLIWC	1.97	THP	THP	0.83	Anticip	Non-CPP	-0.41	Non-Toxin	-0.41	Non-Toxin	-0.32	Non-Toxin	-0.32	Non-Toxin
starPep_12276	PTSNHSPTSCPPTCPGYRWMCLRRF	1.71	THP	THP	0.94	Anticip	Non-CPP	-0.52	Non-Toxin	-0.52	Non-Toxin	-0.68	Non-Toxin	-0.68	Non-Toxin
starPep_10092	GVGSPVVSRLTGICL	0.18	THP	non-THP	0.63	Anticip	Non-CPP	-0.91	Non-Toxin	-0.91	Non-Toxin	-1.13	Non-Toxin	-1.13	Non-Toxin
starPep_07237	QHWVSYGLRPG	1.5	THP	THP	0.29	Anticip	Non-CPP	-0.9	Non-Toxin	-0.9	Non-Toxin	-0.74	Non-Toxin	-0.74	Non-Toxin
starPep_12415	QSFNGQWARGHFH	0.37	THP	THP	-0.53	Non-Anticip	Non-CPP	-1.17	Non-Toxin	-1.17	Non-Toxin	-0.83	Non-Toxin	-0.83	Non-Toxin
starPep_08820	CPSHLDLFC	1.97	THP	THP	0.37	Anticip	Non-CPP	-1.04	Non-Toxin	-1.04	Non-Toxin	-0.9	Non-Toxin	-0.9	Non-Toxin
starPep_01400	GLLSGVLGVGKVKVDCGLSLGLC	0.28	THP	non-THP	1.26	Anticip	Non-CPP	-1.09	Non-Toxin	-1.09	Non-Toxin	-0.66	Non-Toxin	-0.66	Non-Toxin
starPep_43956*	IWRFP	2.9	THP	THP	1.13	Anticip	Non-CPP	-0.77	Non-Toxin	-0.77	Non-Toxin	-0.29	Non-Toxin	-0.29	Non-Toxin
starPep_42404	WRLRIRSAVIRA	-0.21	Non-THP	non-THP	-0.47	Non-Anticip	Non-CPP	-0.96	Non-Toxin	-0.96	Non-Toxin	-1.45	Non-Toxin	-1.45	Non-Toxin
starPep_12257	WRLRIRSAVIRA	1.15	THP	THP	1.02	Anticip	Non-CPP	-1.1	Non-Toxin	-1.1	Non-Toxin	-0.49	Non-Toxin	-0.49	Non-Toxin
starPep_18019	PRPGPIY	2.05	THP	THP	-0.08	Non-Anticip	Non-CPP	-1	Non-Toxin	-1	Non-Toxin	-0.46	Non-Toxin	-0.46	Non-Toxin
starPep_01691	DDPESDL	-0.07	Non-THP	THP	0.16	Anticip	Non-CPP	-1.03	Non-Toxin	-1.03	Non-Toxin	-1.01	Non-Toxin	-1.01	Non-Toxin
starPep_13827*	EGGIPQWAVGHFM	2.34	THP	THP	1.19	Anticip	Non-CPP	-1.13	Non-Toxin	-1.13	Non-Toxin	-0.46	Non-Toxin	-0.46	Non-Toxin
starPep_16808	CPRVC	-0.03	Non-THP	non-THP	1.74	Anticip	CPP	-0.07	Non-Toxin	-0.07	Non-Toxin	0.34	Toxin	0.34	Toxin
starPep_29033	LLLNKKGKKNKHKHGHHGKH	-0.02	Non-THP	THP	0.58	Anticip	Non-CPP	-1.2	Non-Toxin	-1.2	Non-Toxin	-1.03	Non-Toxin	-1.03	Non-Toxin

G. (cont.) Predicted activities of 43 repurposed peptides obtained from hierarchical virtual screening of peptides from starPepDB.*Originally, these peptides contained a X aa, which was changed by an aa that gave them greater tumor homing potential.

ID	Sequence	HemoPI				
		SVM Score 1	SVM Score 2	SVM Score 3	SVM Score 4	
starPep_01400	GLLSGVLGVGKKVDCGLSGLC	0.84	0.72	0.58	0.58	0.67
starPep_01691	EGGGPQWAVGHFM	0	0.06	0.49	0.49	0.45
starPep_02029	TPFKLSLHL	0.6	0.59	0.5	0.5	0.53
starPep_04689	RLRLRIGRR	0.96	0.79	0.48	0.48	0.42
starPep_05157	DGPKKKKKKSPSKSSG	0.54	0.49	0.49	0.49	0.44
starPep_05293	FFRNLWKGAKAAFRAGHAAWRA	0.73	0.94	0.49	0.49	0.46
starPep_07234	QGRLTQWAVGHLM	0.24	0.25	0.49	0.49	0.49
starPep_07237	QHWSYGLRPG	0.16	0.21	0.48	0.48	0.41
starPep_07335	RCICRLGIC	1	0.79	0.49	0.49	0.44
starPep_07641	RSQMQDGLQSCCQELQNVEEQCQC	0	0.15	0.49	0.15	0.44
starPep_08545	AVESTVATLEASPEVIESPPE	0	0	0.48	0.48	0.4
starPep_08820	CPSHLDAFC	0.54	0.5	0.49	0.49	0.43
starPep_10014	GRRSTHWRI	0.85	0.75	0.49	0.49	0.44
starPep_10092	GVGSPYVSRLGICL	0.63	0.58	0.51	0.51	0.43
starPep_10105	GWAGWLLSPRGSRPSWGP	0.6	0.53	0.49	0.49	0.4
starPep_12257	PRPGPIYY	0.09	0.31	0.49	0.49	0.43
starPep_12276	PTSNHSP TSCPPTCPGYRWMCLRRF	0.43	0.49	0.49	0.49	0.39
starPep_12415	QSFNGQWARGHFM	0.11	0.16	0.49	0.49	0.49
starPep_13030	SPRGSRPSWGP TDP RRRS	0.29	0.72	0.49	0.49	0.43
starPep_13108	SVSWGMPSPRQ	0.14	0.22	0.48	0.48	0.38
starPep_13827	RWCFRVCYGCCR	1	0.99	0.61	0.61	0.58
starPep_14535	AGIRRPFGFSP LRIA	0.33	0.34	0.48	0.48	0.34
starPep_15346	AQPSFAF	0.02	0.24	0.49	0.49	0.44
starPep_16575	CKGKGAKAARSGKC	1	1	0.48	0.48	0.39
starPep_16808	CNGRCGGK LAKLAKLAKLAK	1	1	0.42	0.42	0.41
starPep_17042	CTDYVLIWC	0.92	0.78	0.49	0.49	0.46
starPep_18019	DPPFSPRL	0	0.24	0.49	0.49	0.42
starPep_18023	DPSFN SWG	0.13	0.22	0.49	0.49	0.4
starPep_24644	HKHGHGHLKHK NK LK KNGKH	0.77	0.62	0.49	0.49	0.44
starPep_25472	ILPVKWPWWP WRR	0.91	0.86	0.65	0.65	0.72
starPep_26052	IVLVRWPK	0.92	0.81	0.4	0.4	0.31
starPep_27346	KNKGGKWWWW	1	0.68	0.49	0.49	0.44
starPep_27446	KQCISLKGICKDLACT	1	0.92	0.49	0.49	0.55
starPep_27924	KWCFRVAYRGISYRRCR	1	1	0.53	0.53	0.44
starPep_29033	LLNKKGKNKHKGHGHGHKH	0.82	0.69	0.49	0.15	0.45
starPep_35988	QRNKGLRHH	0.32	0.36	0.49	0.49	0.4
starPep_36476	RFWVRGRRS	1	0.94	0.49	0.49	0.41
starPep_41900	VIWRWRKFY	1	1	0.49	0.49	0.5
starPep_42404	VRLRIRSAVIRA	0.8	0.71	0.48	0.48	0.43
starPep_43120	WKGRWYKTT	0.81	0.63	0.49	0.49	0.43
starPep_43502	WWAMKWIRV	1	0.78	0.49	0.49	0.46
starPep_43589	WWWKNKGK KNGKH	0.92	0.69	0.49	0.49	0.45
starPep_43956	KWDPPPPSP	0.28	0.47	0.49	0.49	0.44

H. FASTA of 180 THPs derived from 43 lead hits.

>starPep_43956.L5	>starPep_25472.It4.5L.4.1.L12
WDPPP	ALPYLWPWWPWSR
>starPep_43956.It4.3.4L.1.5	>starPep_25472.It4.5L.4.1.12.L5
WWLLRPPSPP	YLWPW
>starPep_43956.It4.3.4L.1.6	>starPep_25472.It4.5L.4.1.12.L10
WWLLRPPSPP	YLWPWWPWSR
>starPep_43956.It4.3.4W.1.5	>starPep_25472.It4.5Y.4.1.12.L10
LWLWRPPSPP	LYWPWWPWSR
>starPep_43956.It4.3.5.1.4	>starPep_26052.L5.2
WWLRLPPSPP	VRWWP
>starPep_13827.L5.1	>starPep_26052.It4.9W.1.5.4
XRWCF	WVLCSSRPW
>starPep_13827.L5.2	>starPep_26052.It4.9W.1.5.4.L5.1
CFRVC	LCSRW
>starPep_13827.L5.3	>starPep_26052.It4.9W.1.5.4.L5.2
WCFRV	WVLCSS
>starPep_13827.L10	>starPep_27346.L5.2
RWCFRVCYXG	KKWWW
>starPep_14535.L5.3	>starPep_27346.It4.1.3.7.6.L5
IRRPP	CNCGK
>starPep_14535.It4.3.14W.2.4	>starPep_27346.It4.1.5.7.6.L5.1
AWWWRPPGFSPLRWA	CNKGC
>starPep_14535.It4.3.14W.8.4.L10	>starPep_27346.It4.3.5.7.6.L5
WWRPPWFSPL	KNCGC
>starPep_14535.It4.3.14W.8.5.L10.1	>starPep_27446.L5.1
WRWPPWFSPL	KGICK
>starPep_14535.It4.3.14W.8.5.L10.2	>starPep_27446.L5.2
WPPWFSPLRW	QCISL
>starPep_15346.It4.1W.2.5.7	>starPep_27446.L10
WHPSWAM	LKGICKDLAC
>starPep_15346.It4.6.2.5.7	>starPep_27446.L15
AHPSWWM	KQCISLKGICKDLAC
>starPep_15346.It4.1W.2.5.7.L5.1	>starPep_27924.L5.1
WHPSW	KWCFR
>starPep_15346.It4.6.2.5.7.L5.1	>starPep_27924.L5.2
HPSWW	WCFRV
>starPep_15346.It4.6.2.5.7.L5.2	>starPep_27924.L15
PSWWM	WCFRVAYRGISYRRC
>starPep_16575.L5.1	>starPep_27924.It3.1C.6.9.14.L5.2
CKGKG	WCFRC
>starPep_16575.It4.6.2.8.4.L5	>starPep_29033.L5.3
CGCKC	HGHKH
>starPep_16575.It4.6.2.8.7.L5	>starPep_29033.L10
KGCCC	KGKKNKHKGHG
>starPep_16808.L5.1	>starPep_29033.L15
CNGRC	KKGKKNKHKGHGHG
>starPep_16808.L5.2	>starPep_29033.It4.1.2.3.11.L5.1
NGRCG	CCCNK
>starPep_16808.L5.3	>starPep_35988.L5
RCGGK	GLRHH
>starPep_16808.L10	>starPep_35988.It4.4C.8.9.2.L5.1
CNGRCGGKLA	CGLRC
>starPep_17042.L5.1	>starPep_35988.It4.4C.8.9.7.L5.2
LIWCX	GLCCC
>starPep_17042.L5.2	>starPep_35988.It4.4C.8.2.1C.L5
VLIWC	WCNCG
>starPep_17042.It4.2.5.7.4.L5.1	>starPep_36476.It4.1.5.7.2C
CCGVL	WCVWGLRS
>starPep_17042.It4.2.5.7.4.L5.2	>starPep_36476.It4.1.5.7.2C.L5.1
CCCGV	CWVWG
>starPep_18019.It4.1W.2.4.7	>starPep_36476.It4.1.5.7.2C.L5.2
WWPYSPHL	WVWGL
>starPep_18019.It4.1W.6.4.7	>starPep_36476.It4.1.5.7.2A.L5
WPPYSWHL	AWVWG
>starPep_18019.It4.1W.2.4.7.L5	>starPep_41900.L5.4
WWPYS	WRKFY

H. (cont.) FASTA of 180 THPs derived from 43 lead hits.

>starPep_01400.L5.2	>starPep_07335.It4.3W.1.8.L5
LSGLC	CWCRL
>starPep_01400.It4.14.2.3.4.L5.1	>starPep_07641.L5.1
CCGVL	CCQEL
>starPep_01400.It4.14.7.2.6.L5	>starPep_07641.It4.3.5.8.14.L5
CLCGV	CCCEL
>starPep_01691.It4.9.6.3.4	>starPep_08545.It4.3.10.14.17.L5.1
EGWWPWWAWGHFM	WASPW
>starPep_01691.It4.9.6.3.10	>starPep_08545.It4.3.10.14.17.L5.2
EGWGPWWAWWHFM	LWASP
>starPep_01691.It4.9.6.2.4.L10	>starPep_08545.It4.10.14.17.21.L15
WPWWAWGHFM	ATLWASPWWIWSPPW
>starPep_01691.It4.9.6.3.4.L5	>starPep_10014.L5.3
WAWGH	THWRI
>starPep_01691.It4.9.6.3.4.L10	>starPep_10014.It4.9C.3.8.5.L5.1
WWPWWAWGHF	CSLHW
>starPep_08820.L5	>starPep_10014.It4.9C.3.8.5.L5.2
CPSHL	SLHWC
>starPep_08820.It4.6W.8W.2.4.L5	>starPep_10092.It4.13W.2.1.3
CSRLW	WWWSPYVSRLLGWCL
>starPep_08820.It4.6W.8C.2.4.L5.2	>starPep_10092.It4.13W.2.1.3.L5
CWSRL	WWSPY
>starPep_08820.It4.6C.8.2.4.L5.1	>starPep_10092.It4.13W.7.1.3.L10
LCAWC	PYWSRLLGWC
>starPep_08820.It4.6C.8.2.4.L5.2	>starPep_10092.It4.13W.7.1.12.L10.1
WSRLC	PYWSRLLWWC
>starPep_07237.It4.1.6W.10.8C	>starPep_10092.It4.13W.7.1.12.L10.2
WHWSYWLCPC	SPYWSRLLWW
>starPep_07237.It4.1.6C.10.8C	>starPep_10105.It4.1.4.11.17
WHWSYCLCPW	WWAWWLLSPRHSRPSWYP
>starPep_07237.It4.1.6W.10.8C.L5.1	>starPep_10105.It4.1.4.11.17.L10
WHWSY	WWAWWLLSPR
>starPep_07237.It4.1.6W.10.8C.L5.3	>starPep_10105.It4.1.4.11.17.L15
WLCPC	WAWWLLSPRHSRPSW
>starPep_07237.It4.1.6C.10.8C.L5.1	>starPep_10105.It4.1.11.4.17.L15
HWSYC	WWAHWLLSPRWSRPS
>starPep_02029.It4.4W.5.3.7	>starPep_12257.It4.4W.6.1.8
TPWWSYHL	WRPWPLYF
>starPep_02029.It3.4W.7.3.9	>starPep_12257.It4.4L.6.1.7
TPWWLSWHY	WRPLPWFY
>starPep_02029.It4.4W.5.3.7.L5	>starPep_12257.It4.4W.6.1.7.L5.1
WWSYH	WPLFY
>starPep_02029.It4.4W.5.3.9.L5	>starPep_12257.It4.4W.6.1.8.L5
WWSLH	WPLYF
>starPep_02029.It3.4W.7.3.9.L5.1	>starPep_12257.It4.4L.6.1.7.L5
WLSWH	LPWFY
>starPep_04689.L5.1	>starPep_12276.It4.2.8.13.4.L5.1
LRLRI	PWSWH
>starPep_04689.It4.1.3.5.8.L5.1	>starPep_12276.It4.2.8.13.4.L5.2
CIGCR	WHSPW
>starPep_04689.It4.1.3.5.8.L5.2	>starPep_12276.It4.2.8.13.18.L10.1
CLCIG	CPGYWWMCLR
>starPep_04689.It4.1.5.8.9.L5.2	>starPep_12276.It4.2.8.13.18.L10.2
LCIGC	WCPGYWWMCL
>starPep_05157.It4.1C.10.12.4.L5.1	>starPep_12415.L5.1
CGPCK	WARGH
>starPep_05157.It4.1C.10.12.4.L5.2	>starPep_12415.It4.1.6.3.12C
CPCKS	WSWGNWWARGHCM
>starPep_05157.It4.1C.10.12.4.L10	>starPep_12415.It4.1.6.3.5.L5.2
CGPCKKKKKC	CWWAR
>starPep_05157.It4.1C.10.12.6.L10	>starPep_12415.It4.1.6.3.5.L10.1
CGPKKCKKKC	SWGCVWARGH
>starPep_05157.It4.1C.10.14.4.L5	>starPep_12415.It4.1.6.3.5.L10.2
CPSKC	WSWGCWWARG
>starPep_05293.It4.7.10.9.L5	>starPep_13030.L5.1
WFAFR	RPSWG
>starPep_05293.It4.7.10.9.L10	>starPep_13030.It4.13W.12.1.3.L5.1
LWWGWWFAFR	PLWPR
>starPep_05293.It4.7.10.11.12.L5	>starPep_13030.It4.13W.12.1.3.L5.2
LWWGA	WGPLW
>starPep_05293.It4.7.10.11.15.L10.1	>starPep_13030.It4.13W.12.1.6.L10
LWWGAWFAFR	WPSWGPLWPR
>starPep_05293.It4.7.10.11.15.L10.2	>starPep_13108.L5.1
RNLWWGAWWA	SVSWG
>starPep_07234.It4.10W.1.7.6.L5.1	>starPep_13108.It4.12.7W.5.2.L5
RLGCW	HMWPS
>starPep_07234.It4.10W.1.7.6.L5.2	>starPep_13108.It4.12.7W.5.11.L10
LGCWW	SWHMWPSPHW

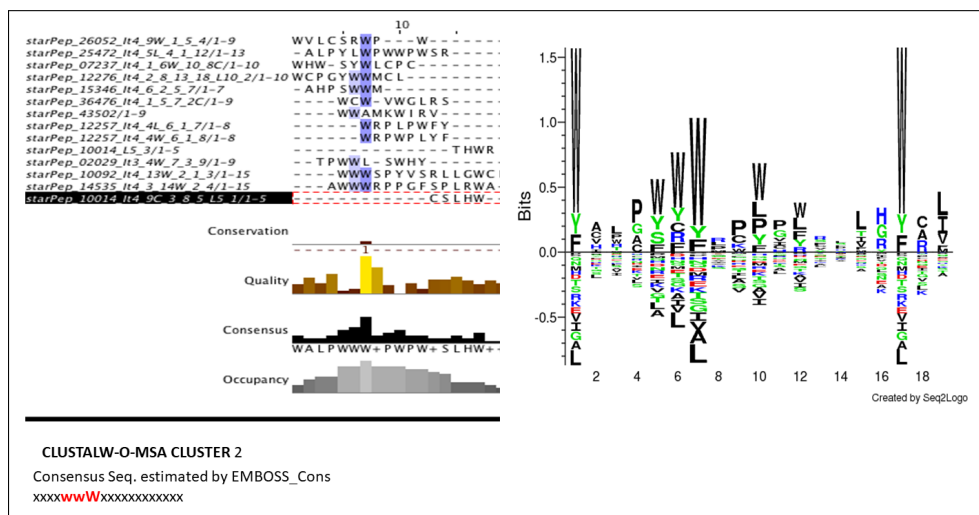
H. (cont.) FASTA of 180 THPs derived from 43 lead hits.

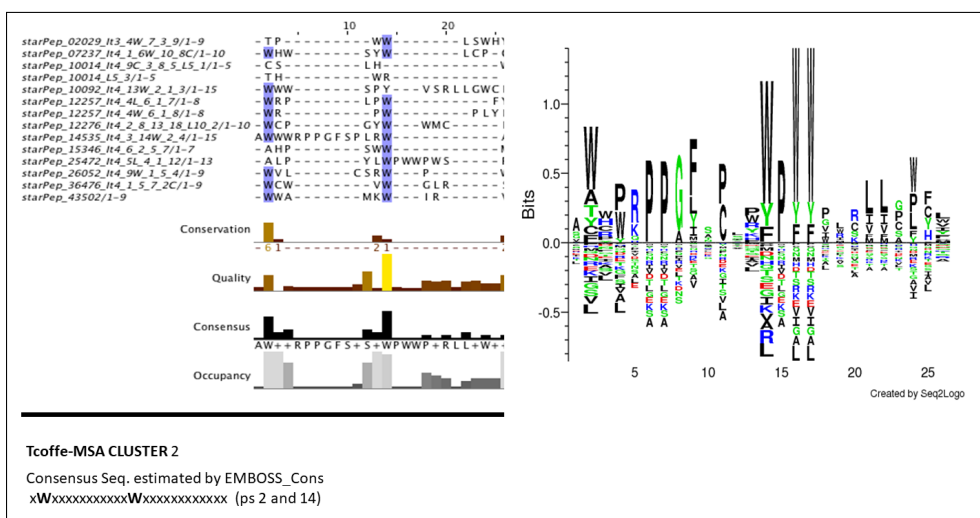
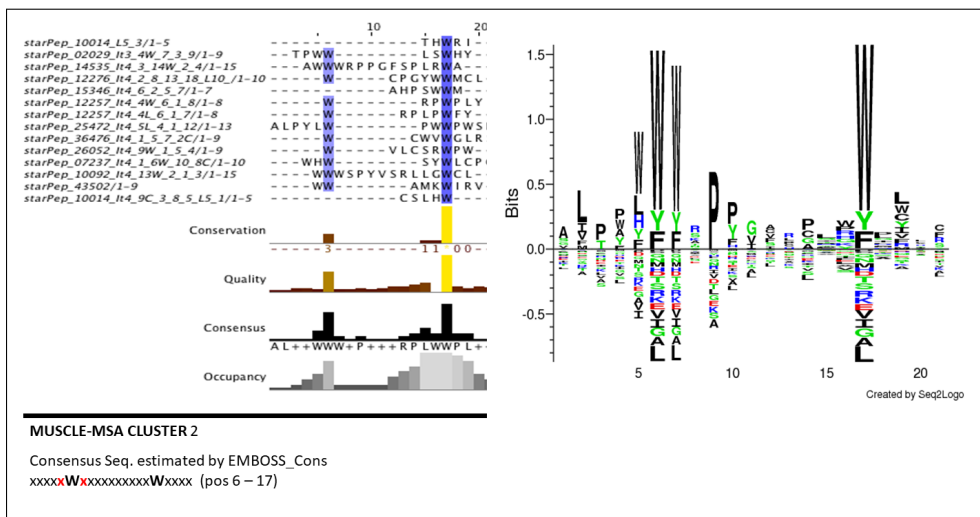
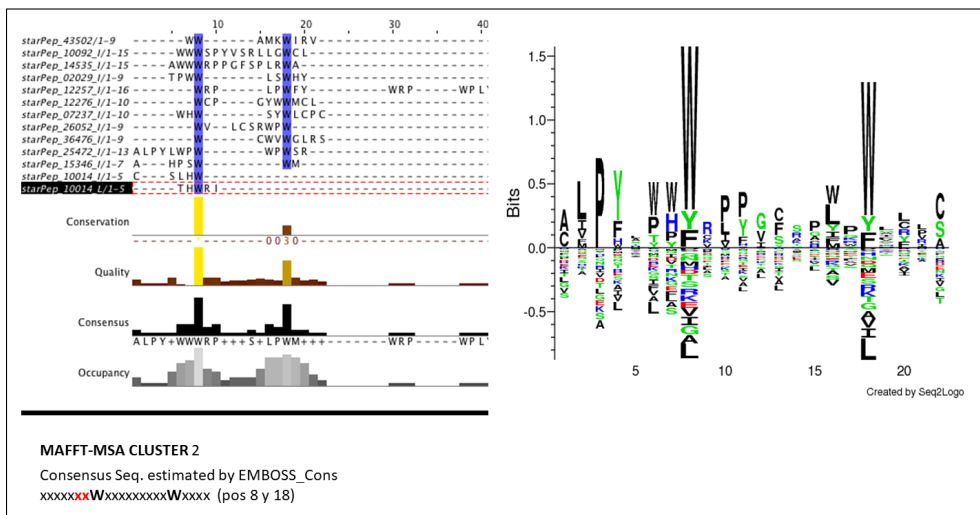
>starPep_18019.It4.1W_3.4.7.L5.2 WPWYS	>starPep_41900.It4.7C.1.2.8 CLWRWRCGY
>starPep_18019.It4.1W_6.4.7.L5 PYSWH	>starPep_41900.It4.7C.1.2.4.L5.1 CLWRW
>starPep_18023.It4.1H.8.5.4C HPSCWSWH	>starPep_41900.It4.7C.1.2.4.L5.2 LWRWC
>starPep_18023.It4.1W_5.8.4C WPSCHSWH	>starPep_42404.It4.1W_9.2.5.L5 WRSWA
>starPep_18023.It4.1H.8.5.4C.L5.1 HPSCW	>starPep_42404.It4.1W_9.2.10.L5.1 RSAWW
>starPep_18023.It4.1H.8.5.4C.L5.2 CWSWH	>starPep_42404.It4.1W_9.2.10.L5.2 SAWWR
>starPep_18023.It4.1W_5.8.4C.L5 WPSCH	>starPep_43120.L5.1 WKGRW
>starPep_24644.L10.1 HKHGHGHLKH	>starPep_43120.L5.2 KGRWY
>starPep_24644.L10.2 HGHGHLKHK	>starPep_43120.It4.2C_8.9.7L.L5.2 RWYLC
>starPep_24644.It4.8C_14.3.5.L5.1 HKCGC	>starPep_43502.It4.5S_7.9.1 CWAMSWCRC
>starPep_24644.It4.8C_14.3.5.L5.3 KCGCG	>starPep_43502.It4.5C_7.9.1 CWAMCWSRC
>starPep_24644.It4.8C_14.3.5.L10 CGCGHCKHKN	>starPep_43502.It4.5S_7.9.1.L5.1 CWAMS
>starPep_43502.It4.5S_7.9.1.L5.2 AMSWC	>starPep_07234.It4.1W_7.6.10.L10.1 WGRLGWWWAC
>starPep_43502.It4.5S_7.9.2.L5.1 WCAMS	>starPep_07234.It4.1W_7.6.10.L10.2 RLGWWWACGH
>starPep_43589.L5.1 WWWKN	>starPep_07335.L5 CICRL
>starPep_43589.L5.4 KGKKN	>starPep_07335.It4.3C.1.8.L5 CCRLG
>starPep_43589.It4.4.6.8.5.L5 CGCKN	>starPep_07335.It4.8.1.3.L5.1 CRLGC
>starPep_01400.L5.1 CGLSG	>starPep_07335.It4.8.1.3.L5.2 RLGCC
>starPep_13108.It4.12.7H.5.11.L10 SWWMHPSPHW	

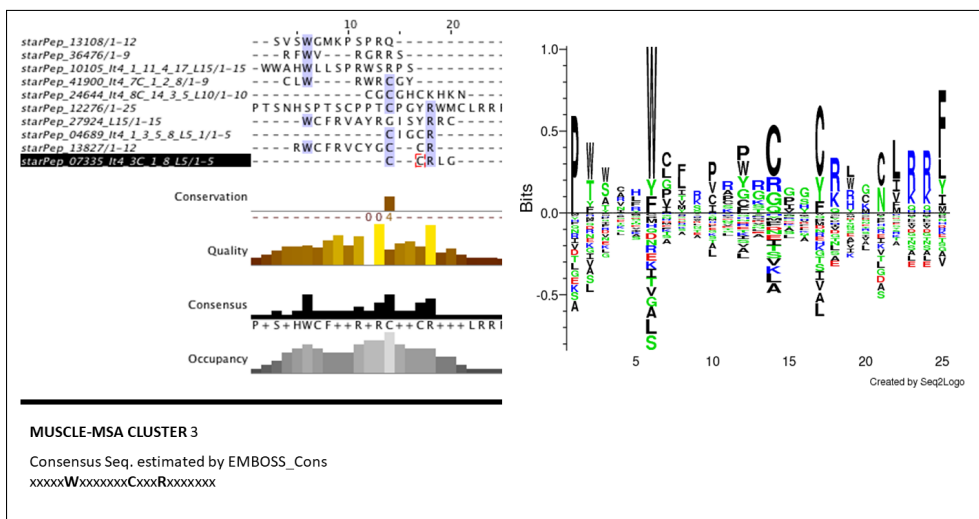
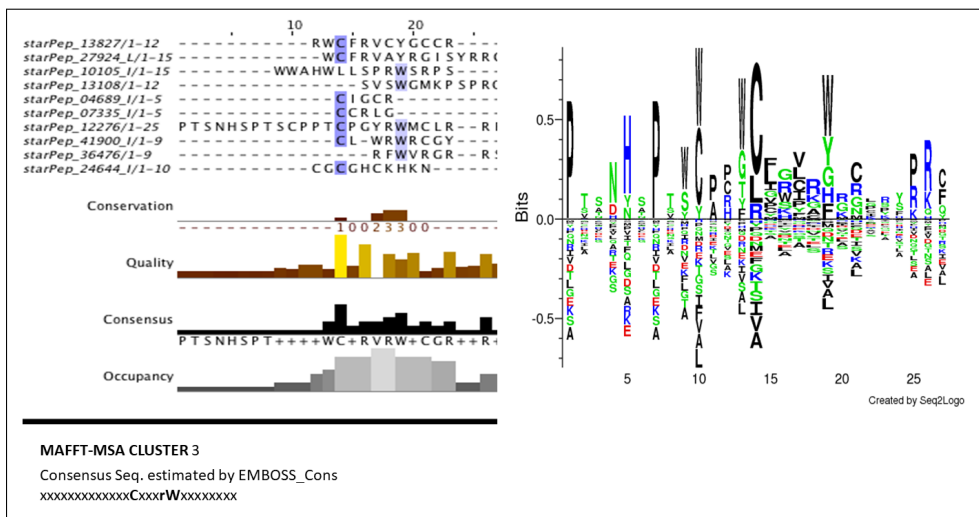
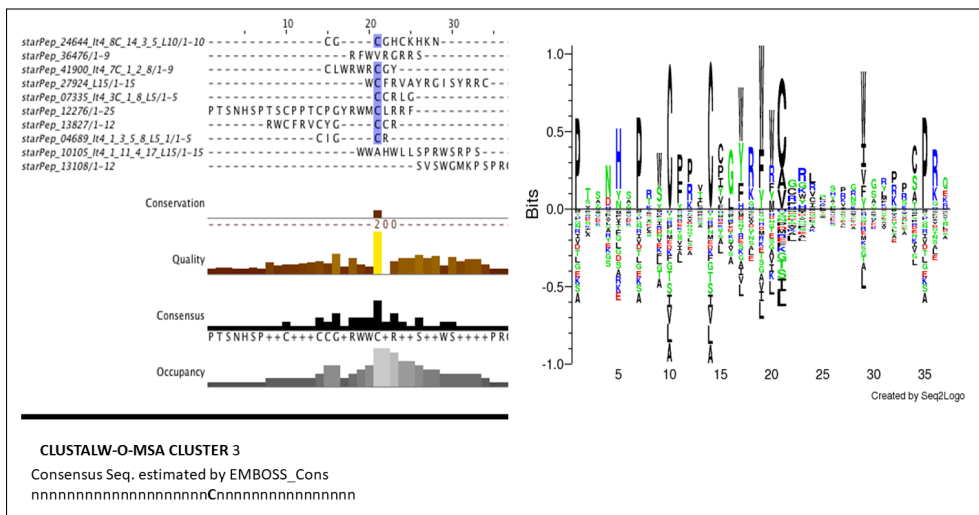
I. (cont.) Predicted activities of SET 1, conformed by 54 lead THPs.

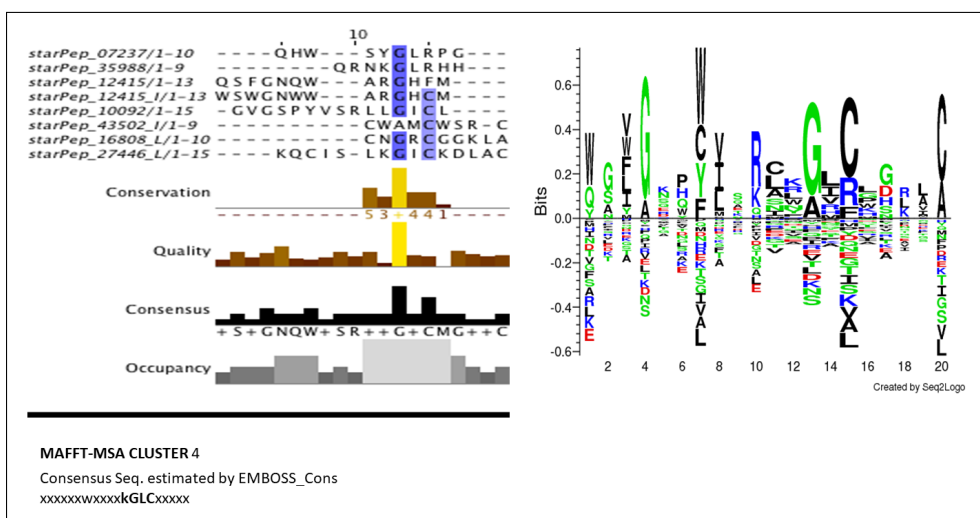
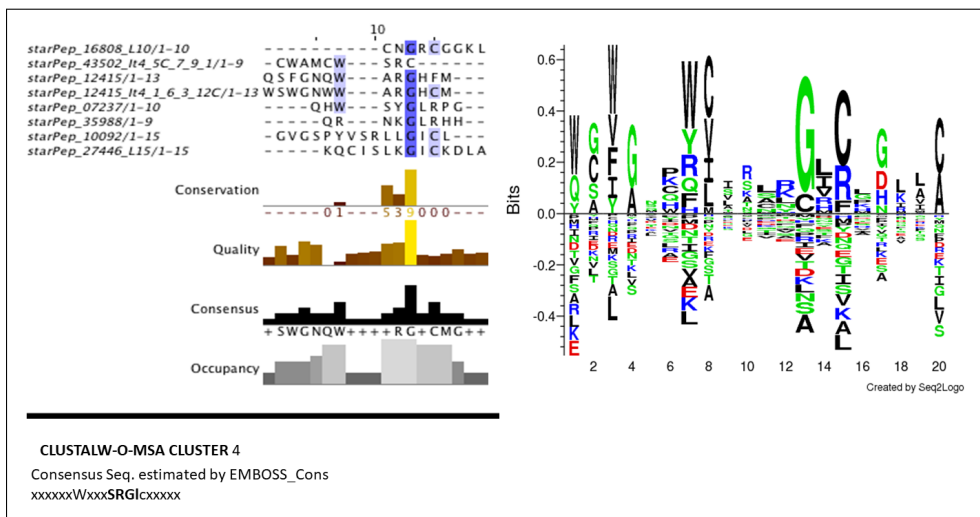
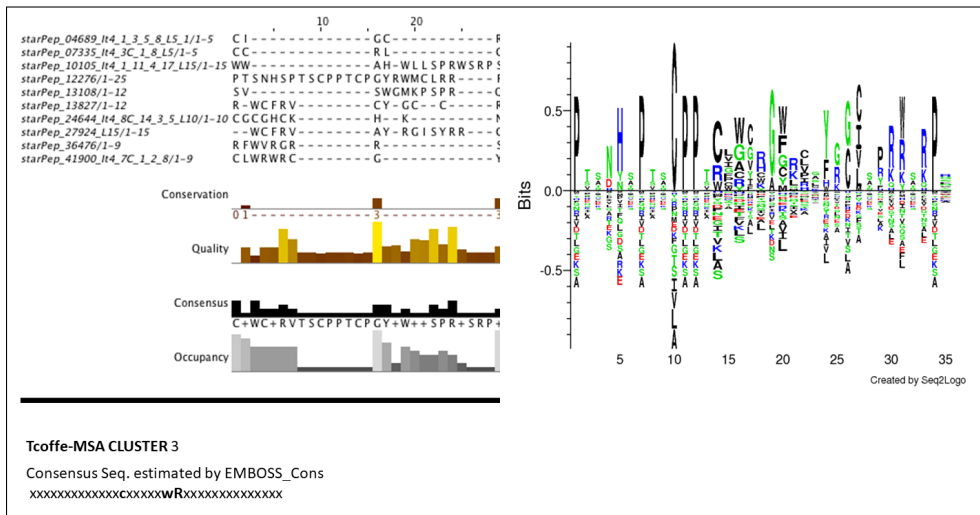
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		SVM Score 1	SVM Score 2	SVM Score 3	SVM Score 3	SVM Score 4
starPep_24644	HKHGHGHLKHKNKLKKNGKH	0.49	0.49	0.62	0.77	0.44
starPep_43502	WWAMKWIRV	0.49	0.49	0.78	1	0.46
starPep_13108	SVSWGMPKSPRQ	0.48	0.48	0.22	0.14	0.38
starPep_35988	QRNKGLRHH	0.49	0.49	0.36	0.32	0.4
starPep_04689	RLRLRIGRR	0.48	0.48	0.79	0.96	0.42
starPep_36476	RFWVRGRRS	0.49	0.49	0.94	1	0.41
starPep_12276	PTSNHSPTSCPPTCPGYRWMCLRRF	0.49	0.49	0.49	0.43	0.39
starPep_10092	GVGSPYVSRLLGICL	0.51	0.51	0.58	0.63	0.43
starPep_07237	QHWSYGLRPG	0.48	0.48	0.21	0.16	0.41
starPep_12415	QSFQNWARGHFM	0.49	0.49	0.16	0.11	0.49
starPep_08820	CPSHLDAFC	0.49	0.49	0.5	0.54	0.43
starPep_43956	KWDPPPPSPP	0.49	0.49	0.47	0.28	0.44
starPep_13827	RWCFRVCYGCCR	0.61	0.61	0.99	1	0.58
starPep_14535.It4.3.14W.2.4	AWWWRRPPGFSPLRWA	0.67	0.65	0.49	0.49	0.43
starPep_25472.It4.5L.4.1.12	ALPYLWVWPWSR	0.7	0.7	0.5	0.5	0.52
starPep_15346.It4.6.2.5.7	AHPSWWM	0.42	0.45	0.49	0.49	0.43
starPep_16808.L10	CNGRCGGKLA	0.91	0.71	0.48	0.48	0.38
starPep_17042.L5.1	LWVC	1	0.77	0.49	0.49	0.44
starPep_17042.It4.2.5.7.4.L5.1	CCGVL	1	0.73	0.49	0.49	0.44
starPep_18023.It4.1W.5.8.4C	WPSCHSWH	0.73	0.57	0.49	0.49	0.44
starPep_24644.It4.8C.14.3.5.L10	CGCGHCKHKN	0.93	0.67	0.49	0.49	0.41
starPep_26052.It4.9W.1.5.4	WVLC SRWP	1	1	0.49	0.49	0.47
starPep_27346.It4.1.5.7.6.L5.1	NKNGC	0.97	0.66	0.49	0.49	0.43
starPep_27446.L15	KQCISLKGICKDLAC	1	0.91	0.48	0.48	0.5
starPep_27924.L15	WCFRVAYRGISYRRC	1	1	0.51	0.51	0.44
starPep_29033.L10	KGKNKHKHGHG	0.67	0.57	0.49	0.49	0.44
starPep_29033.L15	KKGKNKHKHGHGHHK	0.71	0.58	0.49	0.15	0.44
starPep_35988.It4.4C.8.9.7.L5.2	GLCCC	0.95	0.58	0.49	0.49	0.44
starPep_36476.It4.1.5.7.2C	WCWVWGLRS	1	1	0.49	0.49	0.44
starPep_41900.It4.7C.1.2.8	CLWRWRCGY	1	1	0.49	0.49	0.48
starPep_42404.It4.1W.9.2.5.L5	WRS AW	0.88	0.65	0.49	0.49	0.44
starPep_43120.L5.1	WKGRW	0.97	0.67	0.49	0.49	0.46
starPep_43502.It4.5C.7.9.1	CWAMCWSRC	1	0.88	0.49	0.49	0.44
starPep_43502.It4.5S.7.9.2.L5.1	WCAMS	0.86	0.63	0.49	0.49	0.44
starPep_01400.L5.1	CGLSG	0.93	0.67	0.49	0.49	0.45
starPep_08820.L5	CPSHL	0.44	0.41	0.49	0.49	0.44
starPep_07237.It4.1.6W.10.8C	WHWSYWLCPW	1	0.83	0.49	0.49	0.44
starPep_07237.It4.1.6C.10.8C	WHWSYCLCPW	1	0.83	0.49	0.49	0.44
starPep_02029.It3.4W.7.3.9	TPWWLSWHY	0.68	0.61	0.49	0.49	0.43
starPep_04689.L5.1	LRRLRI	0.87	0.76	0.49	0.49	0.42
starPep_04689.It4.1.3.5.8.L5.1	CIGCR	1	0.75	0.49	0.49	0.44
starPep_05157.It4.1C.10.12.4.L5.2	CPCKS	1	0.73	0.49	0.49	0.44
starPep_05293.It4.7.10.11.12.L5	LWWGA	0.96	0.66	0.49	0.49	0.44
starPep_07335.It4.3C.1.8.L5	CCRLG	1	0.74	0.49	0.43	
starPep_07641.L5.1	CCQEL	0.44	0.46	0.49	0.49	0.44
starPep_10014.L5.3	THWRI	0.78	0.65	0.49	0.49	0.44
starPep_10014.It4.9C.3.8.5.L5.1	CSLHW	1	0.72	0.49	0.49	0.44
starPep_10092.It4.13W.2.1.3	WWWSPYVSRLLGWCL	1	0.91	0.49	0.49	0.44
starPep_10105.It4.1.11.4.17.L15	WWAHWLLSPRWSRPS	0.8	0.71	0.49	0.49	0.43
starPep_12257.It4.4W.6.1.8	WRPWPLYF	0.61	0.67	0.51	0.51	0.6
starPep_12257.It4.4L.6.1.7	WRPLPWFY	0.61	0.67	0.51	0.51	0.6
starPep_12257.It4.4W.6.1.8.L5	WPLYF	0.48	0.5	0.49	0.49	0.45
starPep_12276.It4.2.8.13.18.L10.2	WCPGYWWMCL	1	0.84	0.49	0.49	0.44
starPep_12415.It4.1.6.3.12C	WSWGNWWARGHCM	0.86	0.67	0.49	0.49	0.42

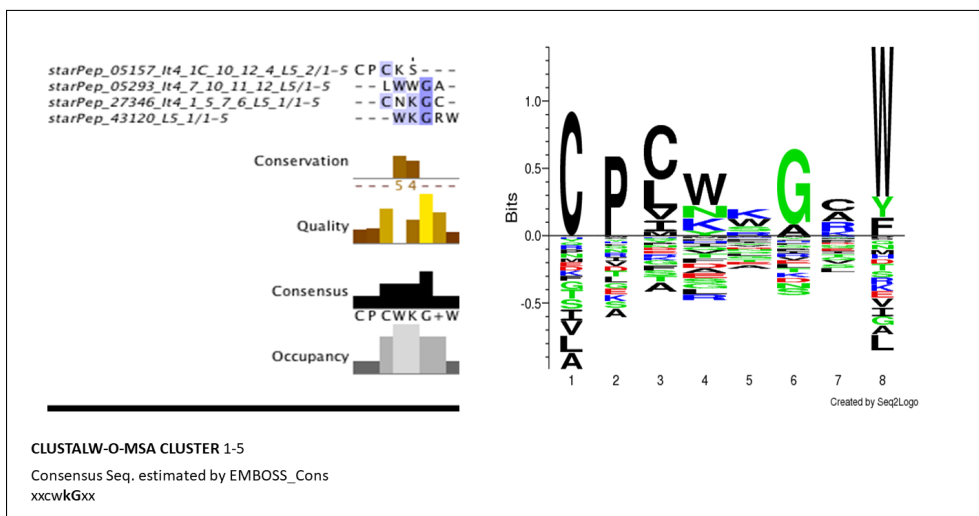
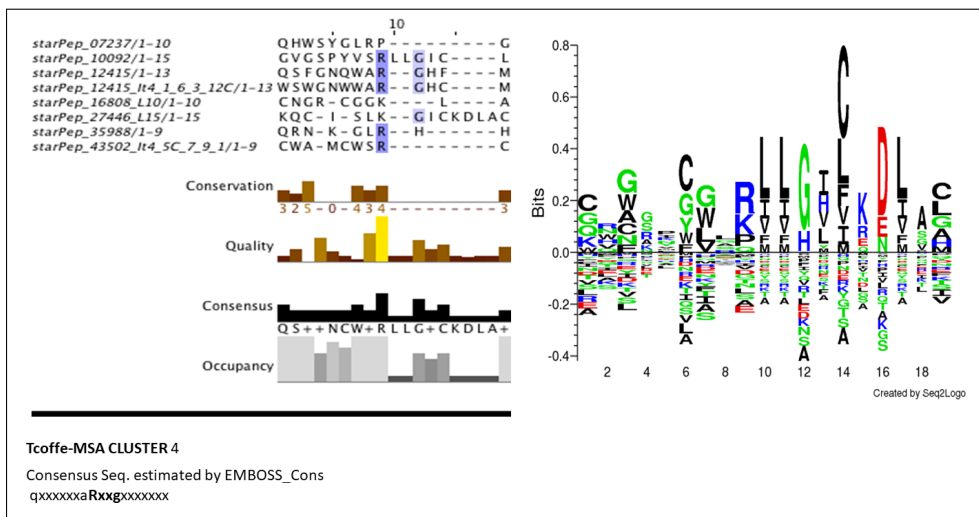
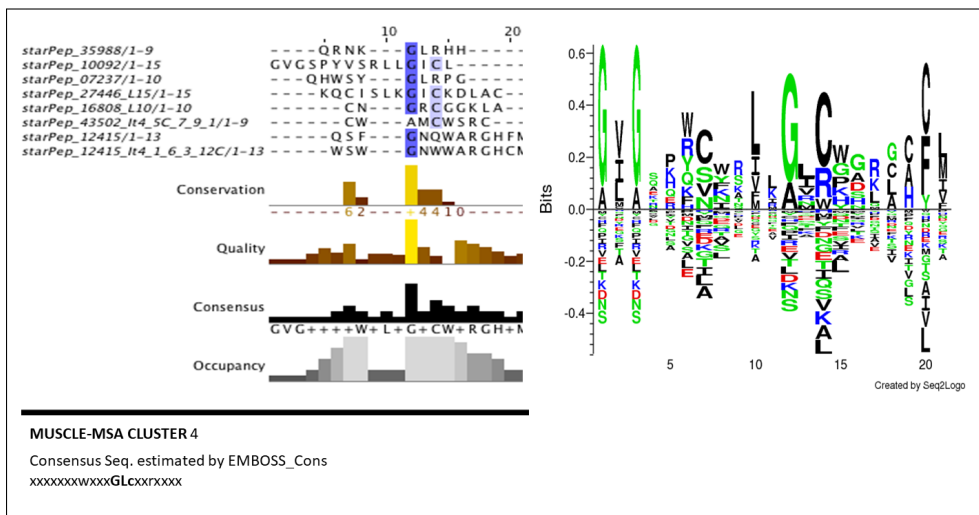
J. Jalview, EMBOSS Cons and Seq2Logo results of MSA.

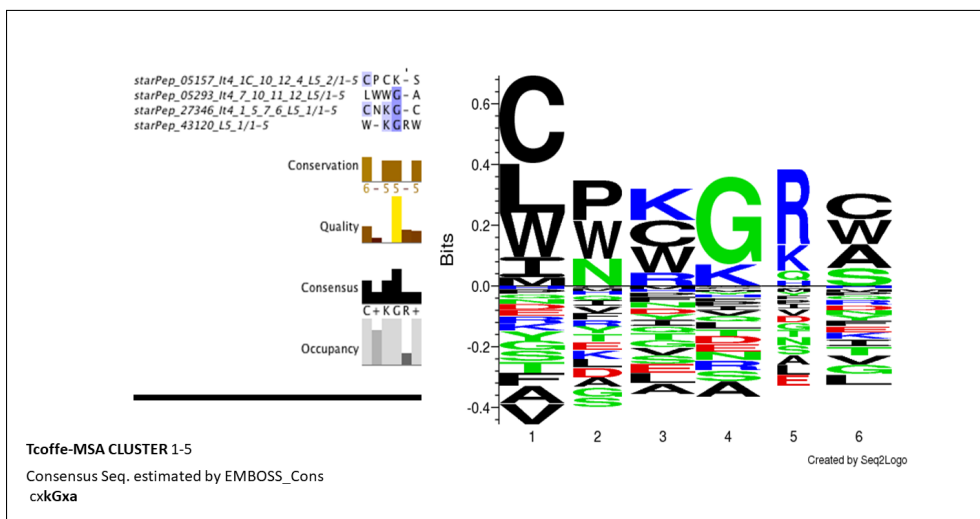
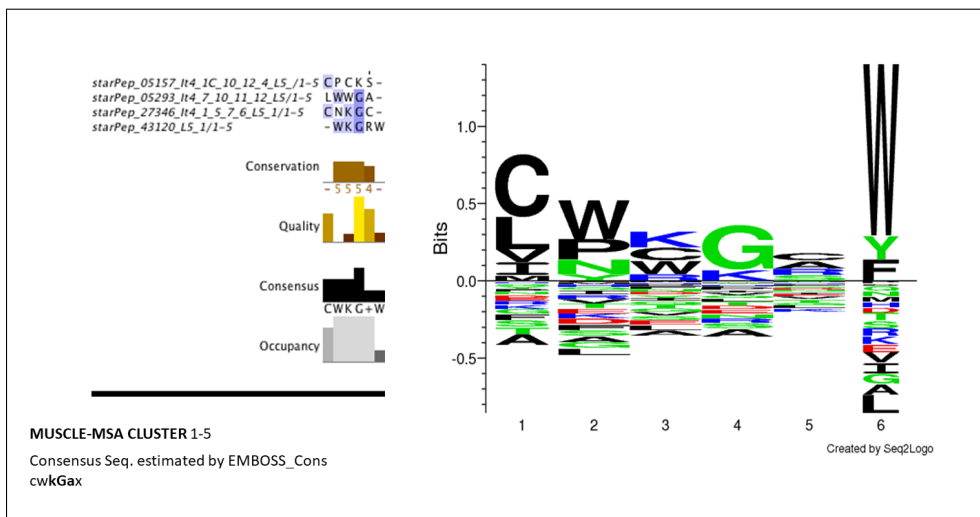
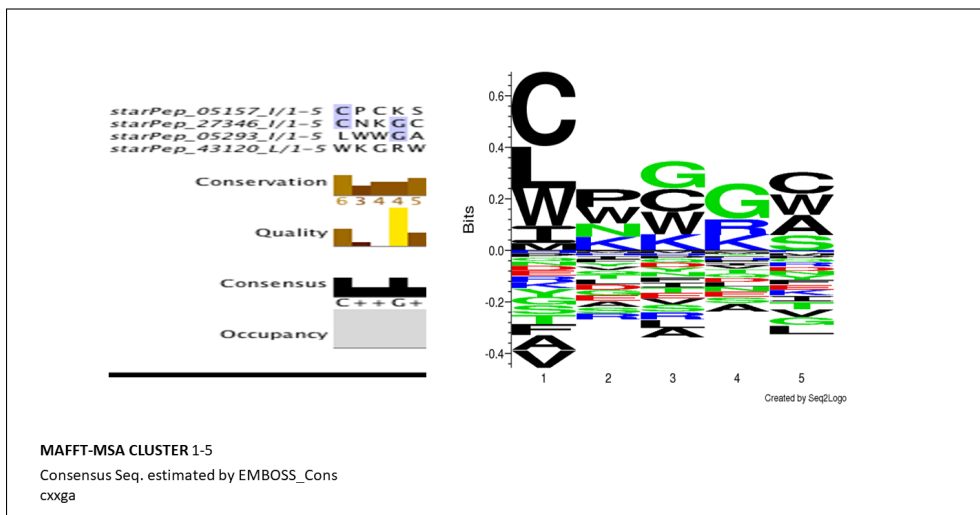


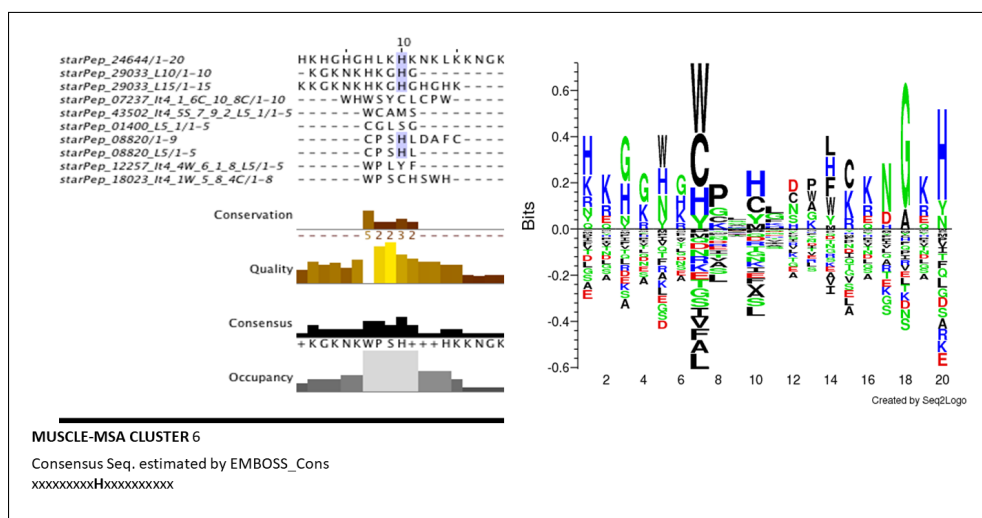
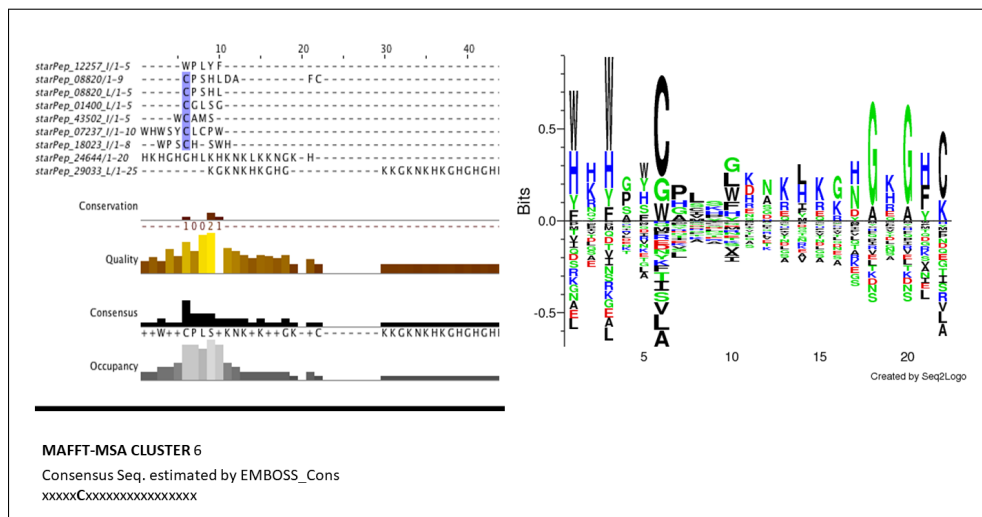
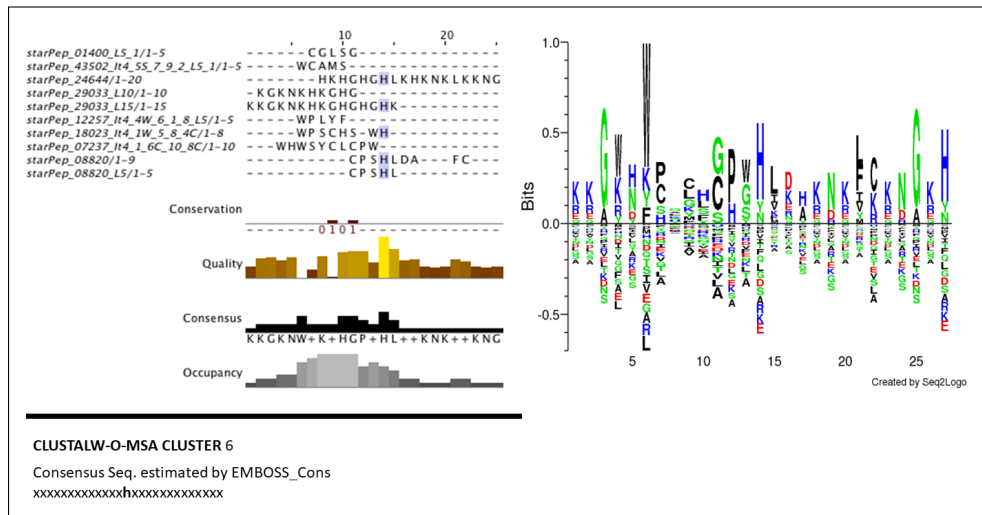


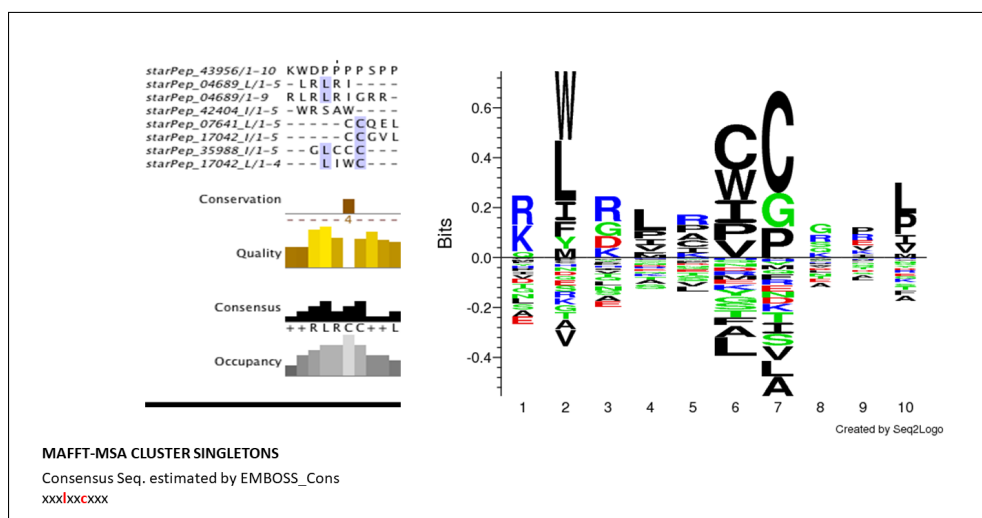
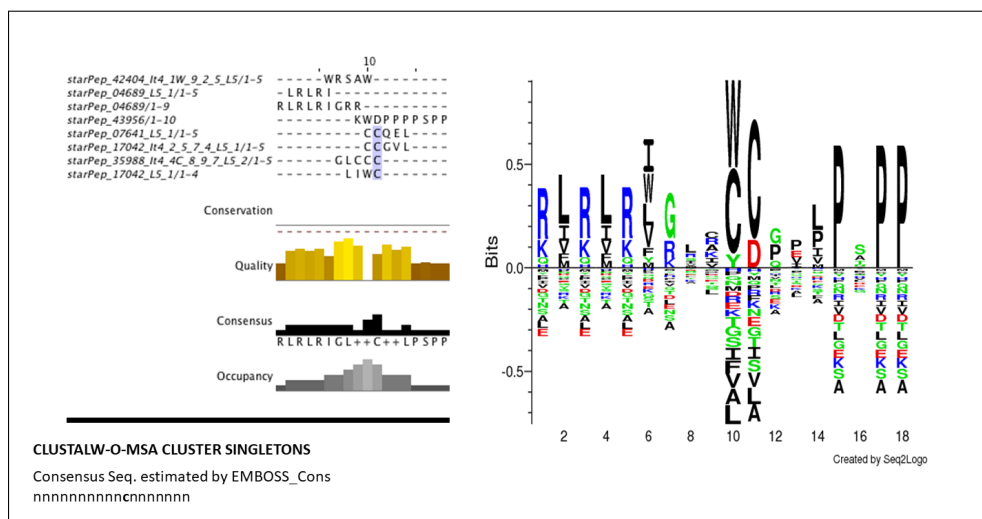
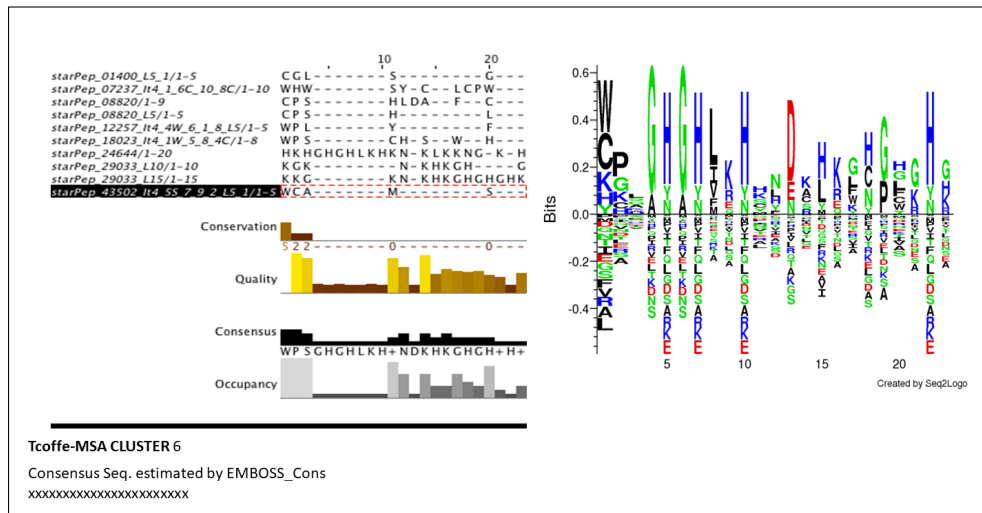


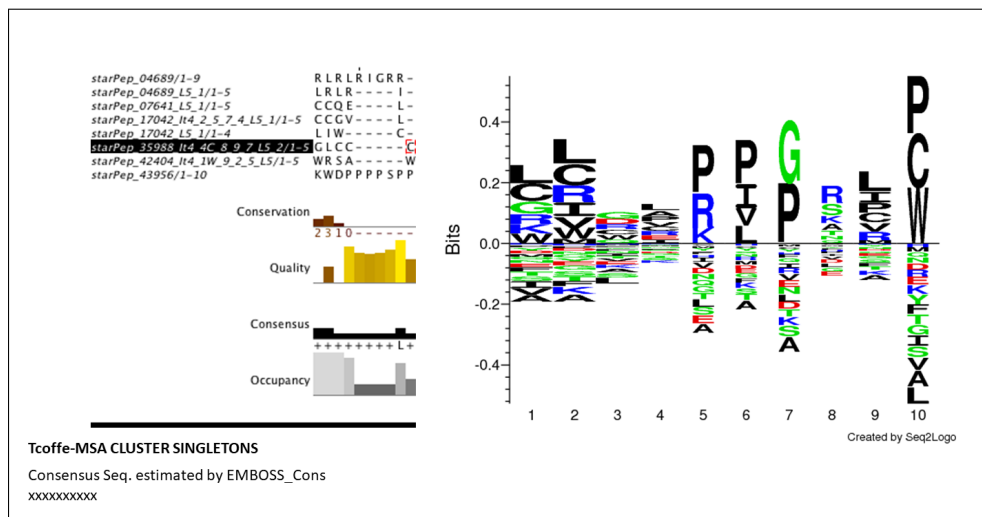
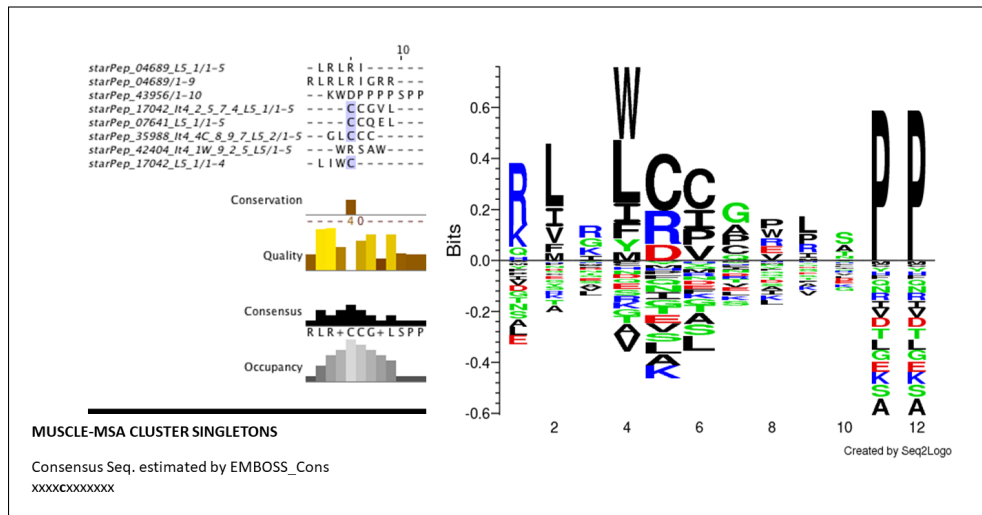












K. FASTA file of 150 cell-penetrating sequences derived from 54 lead hits.

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L. Predicted activities of SET 2, confirmed by 42 lead THPs with optimized cell-penetrating activity.

ID	Sequence	PflsePred		TumourHPD		THPep		AntiCp		ToxinPred		CellPPD	
		Half-time (seconds)	SVM Score	SVM Score	SVM Score	SVM Score	SVM Score 1	SVM Score 2	SVM Score 1	SVM Score 2	SVM Score 1	SVM Score 2	
starPop_4592	WVAMKRVIRV	849.01	1.58	THP	THP	0.5	Anticp	-0.49	Non-Toxin	-0.49	Non-Toxin	-0.81	Non-Toxin
starPop_13108.CPP1	SVFWRMKSPPRQ	868.51	0.86	THP	THP	0.8	Anticp	-1.24	Non-Toxin	-1.24	Non-Toxin	-1.46	Non-Toxin
starPop_13108.CPP4	SVFWRMKSPPRQ	990.11	1.07	THP	THP	0.56	Anticp	-1.47	Non-Toxin	-1.47	Non-Toxin	-1.77	Non-Toxin
starPop_13108.CPP5	SVFWRMKSPPRQ	1012.91	0.79	THP	THP	0.66	Anticp	-1	Non-Toxin	-1	Non-Toxin	-1.39	Non-Toxin
starPop_04689	RLRLRGR	845.01	1.14	THP	THP	0.94	Anticp	-0.24	Non-Toxin	-0.24	Non-Toxin	-0.86	Non-Toxin
starPop_12276	KTRNSHSPTCPGVRWMLRRF	841.71	1.71	THP	THP	0.8	Anticp	-0.22	Non-Toxin	-0.22	Non-Toxin	-0.68	Non-Toxin
starPop_42737.CPP2	KTNSHSPTCPGVRWMLRRR	924.81	1.37	THP	THP	0.8	Anticp	-0.52	Non-Toxin	-0.52	Non-Toxin	-0.41	Non-Toxin
starPop_08820	CHWSVLRER	955.31	1.58	THP	THP	0.9	Anticp	-1.4	Non-Toxin	-1.4	Non-Toxin	-1.1	Non-Toxin
starPop_6926	CPSHLDAR	689.51	1.97	THP	THP	0.99	Anticp	-1.04	Non-Toxin	-1.04	Non-Toxin	-0.9	Non-Toxin
starPop_6926.CPP3	KWVDPPEPSP	835.11	1.25	THP	THP	0.84	Anticp	-0.3	Non-Toxin	-0.3	Non-Toxin	-0.43	Non-Toxin
starPop_16898.L10	KWRPPEPSP	841.01	1.55	THP	THP	0.69	Anticp	-0.55	Non-Toxin	-0.55	Non-Toxin	-0.12	Non-Toxin
starPop_17042.L5.1	GNRCHCGKLA	874.81	2.3	THP	THP	0.74	Anticp	-0.33	Non-Toxin	-0.33	Non-Toxin	-0.06	Non-Toxin
starPop_17042.L5.1.CPP1	CGVIL	823.51	2.39	THP	THP	0.95	Anticp	-0.39	Non-Toxin	-0.39	Non-Toxin	-0.36	Non-Toxin
starPop_17042.L5.1.CPP2	WLPVM	834.11	3.22	THP	THP	1.21	Anticp	-0.04	Non-Toxin	-0.04	Non-Toxin	-0.29	Non-Toxin
starPop_17042.L5.1.CPP3	GLCC	823.51	1.49	THP	THP	1.09	Anticp	-0.73	Non-Toxin	-0.73	Non-Toxin	-0.93	Non-Toxin
starPop_45988.J14.JC.5.9.7.L5.1	KGNKNKHKHGHGHHK	834.81	3.18	THP	THP	0.93	Anticp	-1.16	Non-Toxin	-1.16	Non-Toxin	-0.24	Non-Toxin
starPop_45988.J14.JC.5.9.7.L5.2	KKCKK	834.71	1.07	THP	THP	-0.19	Anticp	-0.32	Non-Toxin	-0.32	Non-Toxin	-0.24	Non-Toxin
starPop_45992.J14.JC.5.9.7.L5.1	WCAMS	830.41	3.35	THP	THP	0.91	Anticp	-0.74	Non-Toxin	-0.74	Non-Toxin	-0.6	Non-Toxin
starPop_45992.J14.JC.5.9.7.L5.2	KKALK	823.31	0.19	THP	non-THP	0.85	Anticp	-0.46	Non-Toxin	-0.46	Non-Toxin	-0.71	Non-Toxin
starPop_01400.L5.1	CGLSG	829.91	1.42	THP	THP	0.7	Anticp	-1.13	Non-Toxin	-1.13	Non-Toxin	-0.45	Non-Toxin
starPop_04689.J14.L1.3.5.L5.1	CPKKK	833.31	1.07	THP	THP	0.69	Anticp	-0.89	Non-Toxin	-0.89	Non-Toxin	-0.71	Non-Toxin
starPop_05157.J14.L1.C.10.12.L1.5.2	CGCR	832.81	2.85	THP	THP	1.25	Anticp	-0.42	Non-Toxin	-0.42	Non-Toxin	-0.65	Non-Toxin
starPop_12415.CPP3	CPCKS	833.51	2.47	THP	THP	0.85	Anticp	-0.59	Non-Toxin	-0.59	Non-Toxin	0	Non-Toxin
starPop_04689.J14.L1.3.5.L5.1	QSFRRQWARRHFM	883.21	0.63	THP	THP	0.85	Anticp	-0.92	Non-Toxin	-0.92	Non-Toxin	-0.13	Non-Toxin
starPop_15346.J14.L6.2.5.7	AWWRRPPRESPLRWA	847.31	2.93	THP	THP	0.79	Anticp	-0.98	Non-Toxin	-0.98	Non-Toxin	-1.13	Non-Toxin
starPop_15346.J14.L6.2.5.7.CPP1	ALPYRVPWVPSR	841.41	3.34	THP	THP	0.92	Anticp	-0.56	Non-Toxin	-0.56	Non-Toxin	-0.45	Non-Toxin
starPop_15346.J14.L6.2.5.7.CPP2	AHPSWVM	837.81	3.5	THP	THP	0.58	Anticp	-0.77	Non-Toxin	-0.77	Non-Toxin	-0.16	Non-Toxin
starPop_18023.J14.L1.W.5.8.JC	CLPVM	832.71	0.71	THP	THP	1.2	Anticp	-0.53	Non-Toxin	-0.53	Non-Toxin	-0.5	Non-Toxin
starPop_18023.J14.L1.W.5.8.JC.CPP1	WPSCHRSWH	927.61	3.58	THP	THP	0.8	Anticp	-0.79	Non-Toxin	-0.79	Non-Toxin	-0.69	Non-Toxin
starPop_21644.J14.L1.C.3.5.L10	GRKKRHW	836.71	0.69	THP	THP	0.4	Anticp	-0.42	Non-Toxin	-0.42	Non-Toxin	-0.61	Non-Toxin
starPop_27346.J14.L5.7.6.L5.1	CGCGHCKHN	834.21	3.42	THP	THP	0.79	Anticp	-0.84	Non-Toxin	-0.84	Non-Toxin	-0.13	Non-Toxin
starPop_29033.L1.L5.CPP2	CNKGC	792.21	0.79	THP	THP	0.82	Anticp	-1.08	Non-Toxin	-1.08	Non-Toxin	-0.77	Non-Toxin
starPop_42929.J13.AW.7.3.9	KKGNKHKHKHKKK	844.71	3.56	THP	THP	0.73	Anticp	-1	Non-Toxin	-1	Non-Toxin	-0.51	Non-Toxin
starPop_42929.J13.AW.7.3.9.CPP1	TPWVLSVHY	835.11	0.61	THP	THP	1.02	Anticp	-0.8	Non-Toxin	-0.8	Non-Toxin	-1.1	Non-Toxin
starPop_42929.J14.L10.L1.L2.L5	IWAWGA	834.81	2.59	THP	THP	0.34	Anticp	-0.86	Non-Toxin	-0.86	Non-Toxin	-0.55	Non-Toxin
starPop_47385.J14.JC.L1.L5	CCRLG	833.71	3.61	THP	THP	0.99	Anticp	-0.4	Non-Toxin	-0.4	Non-Toxin	-0.33	Non-Toxin
starPop_47641.L5.1	CCOFL	834.41	3.04	THP	THP	0.87	Anticp	-0.07	Non-Toxin	-0.07	Non-Toxin	-0.14	Non-Toxin
starPop_10014.L5.3	THWRI	842.61	1.23	THP	THP	0.71	Anticp	-0.89	Non-Toxin	-0.89	Non-Toxin	-0.83	Non-Toxin
starPop_10014.J14.JC.3.8.5.L5.1	CSLHW	813.51	3.28	THP	THP	0.68	Anticp	-0.77	Non-Toxin	-0.77	Non-Toxin	-0.66	Non-Toxin
starPop_12257.J14.L6.L1.L7	WBPPLPWFY	837.51	3.43	THP	THP	1.17	Anticp	-0.87	Non-Toxin	-0.87	Non-Toxin	-0.66	Non-Toxin

M. FASTA of 206 sequences stable in blood.

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>starPep_12415_CPP3_H1      >starPep_27346.It4.1.5.7.6.L5.1.H4      >starPep_10014.It4.9C.3.8.5.L5.1.H5
QSFRLQWARRHFM             WNKGC                                     TSLHW
>starPep_12415_CPP3_H2      >starPep_27346.It4.1.5.7.6.L5.1.H5      >starPep_12257.It4.4L.6.1.7.HL5.1
QSFRLQWARRHFM             CNKGW                                     WRPLP
>starPep_12415_CPP3_H3      >starPep_29033.L15.CPP2.HL5.1          >starPep_12257.It4.4L.6.1.7.HL5.2
QSFRRQWALRHFM            KKGKN                                     RPLPW
>starPep_12415_CPP3_H4      >starPep_29033.L15.CPP2.HL5.2          >starPep_12257.It4.4L.6.1.7.H1
QSFRRQWARLHFM            KGKKN                                     PRPLPWFY
>starPep_12415_CPP3_H5      >starPep_29033.L15.CPP2.HL5.3          >starPep_12257.It4.4L.6.1.7.H2
QSFYRQWARRHFM            KNKHK                                     WRPLPPFY
>starPep_14535.It4.3.14W.2.4.CPP1.HL5.1 >starPep_29033.L15.CPP2.HL5.4          >starPep_12257.It4.4L.6.1.7.H3
SPLRW                     HKHKK                                     WRSLPWFY
>starPep_14535.It4.3.14W.2.4.CPP1.HL5.2 >starPep_29033.L15.CPP2.HL5.5          >starPep_12257.It4.4L.6.1.7.H4
PLRWA                     KHKKK                                     WRPLSWFY
>starPep_14535.It4.3.14W.2.4.CPP1.HL10.1 >starPep_29033.L15.CPP2.HL10.1         >starPep_12257.It4.4L.6.1.7.H5
WWRPPRFSP                GKKNKHKRHHK                             WRQLPWFY
>starPep_14535.It4.3.14W.2.4.CPP1.HL10.2 >starPep_29033.L15.CPP2.HL10.2         >starPep_43502.HL5.3
WRPPRFSP                KKNKHKRHHK                             AMKWI
>starPep_14535.It4.3.14W.2.4.CPP1.HL10.3 >starPep_29033.L15.CPP2.HL10.3         >starPep_43502.H1
RPPRFSP                NKHKRHHKHHK                             SWAMKWIRV
>starPep_14535.It4.3.14W.2.4.CPP1.H3      >starPep_29033.L15.CPP2.HL10.4         >starPep_43502.H2
AWWWRRQPRFSP            KHKRHHKHHK                             WSAMKWIRV
>starPep_14535.It4.3.14W.2.4.CPP1.H4      >starPep_29033.L15.CPP2.H1            >starPep_43502.H3
AWWWRRPFRFSQLR        KKHKNKHKRHHKHHK                         WWAMKSIRV
>starPep_14535.It4.3.14W.2.4.CPP1.H5      >starPep_29033.L15.CPP2.H2            >starPep_13108.CPP1.HL10
AQWWRRPFRFSP            KKGKNKHKHHKHHK                         VPWWMKPSPR
>starPep_25472.It4.5L.4.1.12.CPP2.HL5     >starPep_29033.L15.CPP2.H3            >starPep_13108.CPP1.H1
PYRWP                    KKKKNKHKRHHKHHK                         SVLWRMKPSPRQ
>starPep_25472.It4.5L.4.1.12.CPP2.H1      >starPep_29033.L15.CPP2.H4            >starPep_13108.CPP1.H2
ALPYRQPWWPWSR          KKGKKNKHKRHHKHHK                         SVPWRMKLSPRQ
>starPep_25472.It4.5L.4.1.12.CPP2.H2      >starPep_29033.L15.CPP2.H5            >starPep_13108.CPP1.H3
ALPYRWPQWWSR          KKGKNKHKHHKHHK                         SVPWRMKPSLRQ
>starPep_25472.It4.5L.4.1.12.CPP2.H3      >starPep_02029.It3.4W.7.3.9.H1        >starPep_13108.CPP1.H4
ALPYRWPQWWSR          TPRWLSWHY                               SVTWRMKPSPRQ
>starPep_25472.It4.5L.4.1.12.CPP2.H4      >starPep_02029.It3.4W.7.3.9.H2        >starPep_13108.CPP1.H5
ALPYRWPWWPQSR         TPWRLSWHY                               SVPWRMKTSPRQ
>starPep_25472.It4.5L.4.1.12.CPP2.H5      >starPep_02029.It3.4W.7.3.9.H3        >starPep_13108.CPP4.H1
ALPYRIPWWPWSR         TPWWLSRHY                               SVRWWMKLSPRQ
>starPep_15346.It4.6.2.5.7.H1             >starPep_02029.It3.4W.7.3.9.H4        >starPep_13108.CPP4.H2
AHPSWG                 TPQWLSWHY                               SVRWWMKPSLRQ
>starPep_15346.It4.6.2.5.7.H2             >starPep_02029.It3.4W.7.3.9.H5        >starPep_13108.CPP4.H3
AHPSLWM               TPWWLSQHY                               SVRWWMKTSPRQ
>starPep_15346.It4.6.2.5.7.H4             >starPep_02029.It3.4W.7.3.9.CPP1.HL5.1 >starPep_13108.CPP4.H4
AHPSRWM               SRRWR                                     SVRWWMKPSTRQ
>starPep_15346.It4.6.2.5.7.H5             >starPep_02029.It3.4W.7.3.9.CPP1.HL5.2 >starPep_13108.CPP4.H5
AHPSRWM               RRWRS                                     SVRWWMKISPRQ
>starPep_17042.It4.2.5.7.4.L5.1.CPP2.H1    >starPep_02029.It3.4W.7.3.9.CPP1.HL5.3 >starPep_13108.CPP5.H1
SLPVM                 RWRSR                                     SVRWGWKLSPRQ
>starPep_17042.It4.2.5.7.4.L5.1.CPP2.H2    >starPep_02029.It3.4W.7.3.9.CPP1.HL5.4 >starPep_13108.CPP5.H2
CNPVM                 WRSRH                                     SVRWGWKPSLRQ
>starPep_17042.It4.2.5.7.4.L5.1.CPP2.H3    >starPep_02029.It3.4W.7.3.9.CPP1.H1    >starPep_13108.CPP5.H3
CDPVM                 QRRWRSRHH                               SVRWGWKTSPRQ
>starPep_17042.It4.2.5.7.4.L5.1.CPP2.H4    >starPep_02029.It3.4W.7.3.9.CPP1.H2    >starPep_13108.CPP5.H4
CLPVR                 SRRWRQRHH                               SVRWGWKPSSTRQ
>starPep_17042.It4.2.5.7.4.L5.1.CPP2.H5    >starPep_02029.It3.4W.7.3.9.CPP1.H3    >starPep_13108.CPP5.H5
CLPRM                 SRRWRSRQH                               SVRWGWKASPRQ
>starPep_18023.It4.1W.5.8.4C.HL5.1         >starPep_02029.It3.4W.7.3.9.CPP1.H4    >starPep_04689.HL5
WPSCH                 SRRWRSRHQ                               RIGRR
>starPep_18023.It4.1W.5.8.4C.HL5.2         >starPep_05293.It4.7.10.11.12.L5.H2    >starPep_04689.H3
PSCHS                 LRWGA                                     RQRLRIGRR
>starPep_18023.It4.1W.5.8.4C.HL5.3         >starPep_05293.It4.7.10.11.12.L5.H3    >starPep_04689.H4
SCHSW                 LWRGA                                     RLRQRIGRR
>starPep_18023.It4.1W.5.8.4C.HL5.4         >starPep_05293.It4.7.10.11.12.L5.H4    >starPep_12276.H1
CHSWH                 LSWGA                                     PTSNHSPTSWPPTCPGYRWMCLRRF
>starPep_18023.It4.1W.5.8.4C.H1            >starPep_07641.L5.1.H1                 >starPep_12276.H2
WPSCGSWH              WCQEL                                     PTSNHSPTSCPPTWPGYRWMCLRRF
>starPep_18023.It4.1W.5.8.4C.H2            >starPep_07641.L5.1.H2                 >starPep_12276.H3
WPSCTSWH              CWQEL                                     PTSNHSPTSCPPTCPGYRWMWLLRRF
>starPep_18023.It4.1W.5.8.4C.H3            >starPep_07641.L5.1.H3                 >starPep_12276.H4
WPSCHSWT              NCQEL                                     WTSNHSPTSCPPTCPGYRWMCLRRF
>starPep_18023.It4.1W.5.8.4C.H4            >starPep_07641.L5.1.H4                 >starPep_12276.H5
WPSCASWH              CNQEL                                     PTSNHSWTS CPPTCPGYRWMCLRRF
>starPep_18023.It4.1W.5.8.4C.H5            >starPep_07641.L5.1.H5                 >starPep_12276.CPP1.H1
WPSCHSWA              ICQEL                                     KTRNHSQTS CPPTCPYRWMCLRRR
>starPep_18023.It4.1W.5.8.4C.CPP1.H1      >starPep_10014.L5.3.H1                 >starPep_12276.CPP1.H2
GTKKRRWC              TLWRI                                     KTRNHSPTSCQPTCPYRWMCLRRR
>starPep_18023.It4.1W.5.8.4C.CPP1.H2      >starPep_10014.L5.3.H2                 >starPep_12276.CPP1.H3
GRKKTRWC              TSWRI                                     KTRNHSPTSCQPTCPYRWMCLRRR
>starPep_18023.It4.1W.5.8.4C.CPP1.H3      >starPep_10014.L5.3.H3                 >starPep_12276.CPP1.H5
GRKKRTWC              TQWRI                                     KTRNHSPTSQPPTCPYRWMCLRRR
>starPep_24644.It4.8C.14.3.5.L10.H1        >starPep_10014.L5.3.H4                 >starPep_07237.CPP2.H1
CGCGRCKHKN            TAWRI                                     QHWSYRLRPK
>starPep_24644.It4.8C.14.3.5.L10.H2        >starPep_10014.It4.9C.3.8.5.L5.1.H1    >starPep_07237.CPP2.H2
CGCGHCKRKN            RSLHW                                     QHWSYRLKPR
>starPep_27346.It4.1.5.7.6.L5.1.H1        >starPep_10014.It4.9C.3.8.5.L5.1.H2    >starPep_07237.CPP2.H3
RNKGC                 SSLHW                                     QHWSYKLRPR
>starPep_27346.It4.1.5.7.6.L5.1.H2        >starPep_10014.It4.9C.3.8.5.L5.1.H3    >starPep_07237.CPP2.H4
CNKGR                 CSLSW                                     QHWSYRLRPT
>starPep_27346.It4.1.5.7.6.L5.1.H3        >starPep_10014.It4.9C.3.8.5.L5.1.H4    >starPep_07237.CPP2.H5
CRKGC                 PSLHW                                     QHWSYRLTPR

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M. (cont.) FASTA of 206 sequences stable in blood.

>starPep.08820_HL5.1	>starPep.29033_L15.CPP1_HL5.6
CPSHL	HGHGH
>starPep.08820_HL5.2	>starPep.29033_L15.CPP1_HL5.7
PSHL	GHHGH
>starPep.08820_HL5.5	>starPep.29033_L15.CPP1_HL10.1
LDAFC	GKNKHKHGH
>starPep.08820_H1	>starPep.29033_L15.CPP1_HL10.2
GPSHLDAFC	KNKHKHGH
>starPep.08820_H2	>starPep.29033_L15.CPP1_HL10.3
CPSHLDAFC	NKHKHGH
>starPep.08820_H3	>starPep.29033_L15.CPP1_HL10.4
CPSHLGAF	KHKHGH
>starPep.08820_H4	>starPep.29033_L15.CPP1_H1
CPSHGDAF	HKGKNKHKHGH
>starPep.08820_H5	>starPep.29033_L15.CPP1_H2
CPSHLDAGC	KHGKNKHKHGH
>starPep.43956_HL5	>starPep.29033_L15.CPP1_H3
KWDPP	KKGHNKHKHGH
>starPep.43956_H1	>starPep.29033_L15.CPP1_H4
KWDRPPSP	KKGKHKHGH
>starPep.43956_H2	>starPep.29033_L15.CPP1_H5
KWDPRPPSP	KKGKNHKKHGH
>starPep.43956_H3	>starPep.35988_It4.4C.8.9.7.L5.2.CPP2.H1
KWDPPRSP	CKCKK
>starPep.43956_H4	>starPep.35988_It4.4C.8.9.7.L5.2.CPP2.H5
KWDPPRSPP	KKCKK
>starPep.43956_H5	>starPep.43502_It4.5S.7.9.2.L5.1.H1
KWDPPSRP	WCAGS
>starPep.43956_CPP3_HL5	>starPep.43502_It4.5S.7.9.2.L5.1.H2
KWRPP	WGAMS
>starPep.43956_CPP3_H1	>starPep.43502_It4.5S.7.9.2.L5.1.H3
KWRYPPRPP	WLAMS
>starPep.43956_CPP3_H2	>starPep.43502_It4.5S.7.9.2.L5.1.H4
KWRPYPPRPP	GCAMS
>starPep.43956_CPP3_H3	>starPep.43502_It4.5S.7.9.2.L5.1.H5
KWRPPYRPP	WKAMS
>starPep.43956_CPP3_H4	>starPep.01400_L5.1.H1
KWRPPYRPP	CALSG
>starPep.43956_CPP3_H5	>starPep.01400_L5.1.H2
KWRPPRYP	CGLSA
>starPep.16808_L10_H5	>starPep.01400_L5.1.H3
FNGRCGGKLA	CPLSG
>starPep.17042.L5.1.H1	>starPep.01400_L5.1.H4
LIWT	CGLSP
>starPep.17042.L5.1.H2	>starPep.01400_L5.1.H5
LIWA	CGPSG
>starPep.17042.L5.1.H4	>starPep.08820_L5.CPP1_H1
LIWQ	CPGKK
>starPep.17042.It4.2.5.7.4.L5.1.H4	>starPep.08820_L5.CPP1_H2
ECGVL	CPKGG
>starPep.17042.It4.2.5.7.4.L5.1.H5	>starPep.08820_L5.CPP1_H3
CEGVL	CPKKG
>starPep.17042.It4.2.5.7.4.L5.1.CPP3_H1	>starPep.08820_L5.CPP1_H4
WLPVS	CPRKK
>starPep.17042.It4.2.5.7.4.L5.1.CPP3_H2	>starPep.08820_L5.CPP1_H5
WLSVM	CPKRK
>starPep.17042.It4.2.5.7.4.L5.1.CPP3_H3	>starPep.04689_It4.1.3.5.8.L5.1.H4
SLPVM	AIGCR
>starPep.17042.It4.2.5.7.4.L5.1.CPP3_H4	>starPep.04689_It4.1.3.5.8.L5.1.H5
WLPVR	CIGAR
>starPep.17042.It4.2.5.7.4.L5.1.CPP3_H5	>starPep.05157_It4.1C.10.12.4.L5.2.H1
WLPKM	GPCKS
>starPep.29033_L15.CPP1_HL5.1	>starPep.05157_It4.1C.10.12.4.L5.2.H2
GKNKH	CPGKS
>starPep.29033_L15.CPP1_HL5.2	>starPep.05157_It4.1C.10.12.4.L5.2.H3
KHKKH	SPCKS
>starPep.29033_L15.CPP1_HL5.3	>starPep.05157_It4.1C.10.12.4.L5.2.H4
HKKHG	CPSKS
>starPep.29033_L15.CPP1_HL5.4	>starPep.05157_It4.1C.10.12.4.L5.2.H5
KKHGH	CPCKT
>starPep.29033_L15.CPP1_HL5.5	
KHGHG	

N. FASTA of 250 sequences stable in gastrointestinal tract.

>starPep_43502.H2.G5.1 WSAMK	>starPep_17042.It4.2.5.7.4.L5.1.CPP3.H1.G.2 WLPNS	>starPep_15346.It4.6.2.5.7.H4.G1.2 AHPSWQM
>starPep_43502.H2.G5.2 SAMKW	>starPep_17042.It4.2.5.7.4.L5.1.CPP3.H1.G.3 WLPTS	>starPep_15346.It4.6.2.5.7.H4.G1.3 AHPSWAM
>starPep_43502.H2.G.1 WSAMKWIPV	>starPep_29033.L15.CPP1.H5.G5.1 GHGHK	>starPep_15346.It4.6.2.5.7.H4.G1.4 AFPSWRM
>starPep_43502.H2.G.2 WSAMKWITV	>starPep_29033.L15.CPP1.H5.G5.2 HGHHG	>starPep_15346.It4.6.2.5.7.H4.G1.5 AHTSWRM
>starPep_43502.H2.G.3 WSAMKWIRY	>starPep_29033.L15.CPP1.H5.G10.1 NHHKKHGHGH	>starPep_17042.It4.2.5.7.4.L5.1.CPP2.H3.G.1 CDPGM
>starPep_13108.CPP1.H3.G5 WRMKP	>starPep_29033.L15.CPP1.H5.G10.2 HHKKHGHGHK	>starPep_17042.It4.2.5.7.4.L5.1.CPP2.H3.G.2 CDPQM
>starPep_13108.CPP1.H3.G.1 SIPWRMKPSLRQ	>starPep_43502.It4.5S.7.9.2.L5.1.H4.G.1 GCQMS	>starPep_17042.It4.2.5.7.4.L5.1.CPP2.H3.G.3 CDPQM
>starPep_13108.CPP1.H3.G.2 SVPWRMKPSLPQ	>starPep_43502.It4.5S.7.9.2.L5.1.H4.G.2 GCAGS	>starPep_17042.It4.2.5.7.4.L5.1.CPP2.H3.G.4 CGPVM
>starPep_13108.CPP1.H3.G.3 SLPWRMKPSLRQ	>starPep_43502.It4.5S.7.9.2.L5.1.H4.G.3 GCAMW	>starPep_18023.It4.1W.5.8.4C.H5.G5.1 WPSCH
>starPep_13108.CPP1.H3.G.4 SLPWRMKPSLNQ	>starPep_43502.It4.5S.7.9.2.L5.1.H4.G.4 GFAMW	>starPep_18023.It4.1W.5.8.4C.H5.G5.2 SCHSW
>starPep_04689.H3.G5.1 RQRLR	>starPep_01400.L5.1.H4.G.1 CGLLP	>starPep_18023.It4.1W.5.8.4C.H5.G5.3 CHSWA
>starPep_04689.H3.G5.2 QRLRI	>starPep_01400.L5.1.H4.G.2 CGLPP	>starPep_18023.It4.1W.5.8.4C.H5.G WPGCHSWA
>starPep_04689.H3.G.1 RQRLPIGRR	>starPep_01400.L5.1.H4.G.3 CGFSP	>starPep_24644.It4.8C.14.3.5.L10.H2.G.4 CGCGDCKRKN
>starPep_04689.H3.G.2 RQRLRPGR	>starPep_01400.L5.1.H4.G.4 CGLNP	>starPep_27346.It4.1.5.7.6.L5.1.H3.G.1 CRPGC
>starPep_12276.H5.G5.1 PGYRW	>starPep_01400.L5.1.H4.G.5 CGDLP	>starPep_27346.It4.1.5.7.6.L5.1.H3.G.3 CRFGC
>starPep_12276.H5.G5.2 CPGYR	>starPep_05157.It4.1C.10.12.4.L5.2.H5.G.1 CPCKL	>starPep_27346.It4.1.5.7.6.L5.1.H3.G.4 CSKGC
>starPep_12276.H5.G10.4 PTSNHSWTSC	>starPep_12415.CPP3.H4.G5.1 WARLH	>starPep_27346.It4.1.5.7.6.L5.1.H3.G.5 CRCGF
>starPep_12276.H5.G15.2 PPTCPGYRWMCLRRF	>starPep_12415.CPP3.H4.G5.2 QWARL	>starPep_02029.It3.4W.7.3.9.H5.G5.1 TPWWL
>starPep_12276.H5.G.1 PTSNHSWTSCPPTCPGGRWMCLRRF	>starPep_12415.CPP3.H4.G5.3 SFRRQ	>starPep_02029.It3.4W.7.3.9.H5.G5.2 PWWLS
>starPep_12276.H5.G.2 PTSGHSWTSCPPTCPGYRWMCLRRF	>starPep_12415.CPP3.H4.G10.4 FRRQWARLHF	>starPep_02029.It3.4W.7.3.9.H5.G.1 TPWWLNQHY
>starPep_12276.H5.G.3 PTSNHSWTSCPPGCPGYRWMCLRGF	>starPep_12415.CPP3.H4.G.1 QSFRQWARLGF	>starPep_02029.It3.4W.7.3.9.H5.G.2 TPWWLPQHY
>starPep_07237.CPP2.H2.G.1 QHWSYRLLPR	>starPep_12415.CPP3.H4.G.2 QSFRQWARLPFM	>starPep_02029.It3.4W.7.3.9.H5.G.3 TPWWLSIHY
>starPep_07237.CPP2.H2.G.2 QHWSYRLLPR	>starPep_12415.CPP3.H4.G.3 QSIRQWARLPFM	>starPep_02029.It3.4W.7.3.9.H5.G.4 LPWWLSQHY
>starPep_07237.CPP2.H2.G.3 QHDSYRLLPR	>starPep_14535.It4.3.14W.2.4.CPP1.HL10.2.G5 FSPLR	>starPep_02029.It3.4W.7.3.9.H5.G.5 TPWWLSGHY
>starPep_07237.CPP2.H2.G.4 QHDSYRLLPR	>starPep_14535.It4.3.14W.2.4.CPP1.HL10.2.G.1 WRPPGFSPLR	>starPep_02029.It3.4W.7.3.9.CPP1.H1.G5.1 QRRWR
>starPep_08820.H3.G5 CPSHL	>starPep_14535.It4.3.14W.2.4.CPP1.HL10.2.G.2 WRPPRFPLPR	>starPep_02029.It3.4W.7.3.9.CPP1.H1.G5.2 RRWRS
>starPep_08820.H3.G.1 CPSHLPAFC	>starPep_14535.It4.3.14W.2.4.CPP1.HL10.2.G.3 WRPPRFSPLP	>starPep_02029.It3.4W.7.3.9.CPP1.H1.G.1 QMRWRSRHH
>starPep_08820.H3.G.3 CPSHLGFFC	>starPep_14535.It4.3.14W.2.4.CPP1.HL10.2.G.4 WLPPRFSPLR	>starPep_02029.It3.4W.7.3.9.CPP1.H1.G.2 QRRWTSRHH
>starPep_08820.H3.G.4 CPSHLPGFC	>starPep_14535.It4.3.14W.2.4.CPP1.HL10.2.G.5 WRPPRFGLPR	>starPep_02029.It3.4W.7.3.9.CPP1.H1.G.3 QCRWRSRHH
>starPep_43956.CPP3.H1.G5 WRYPP	>starPep_25472.It4.5L.4.1.12.CPP2.H2.G.1 ALPYRWPQWPWSI	>starPep_02029.It3.4W.7.3.9.CPP1.H1.G.4 QMRWDSRHH
>starPep_43956.CPP3.H1.G.1 KWRYPLRPP	>starPep_25472.It4.5L.4.1.12.CPP2.H2.G.2 ALPYRWPQLPWSR	>starPep_02029.It3.4W.7.3.9.CPP1.H1.G.5 QCRWDSRHH
>starPep_43956.CPP3.H1.G.2 KWRYPPRLP	>starPep_25472.It4.5L.4.1.12.CPP2.H2.G.3 ALPYRLPQWPWSR	>starPep_05293.It4.7.10.11.12.L5.H3.G.3 LWPGF
>starPep_43956.CPP3.H1.G.3 KWRYPPPLP	>starPep_25472.It4.5L.4.1.12.CPP2.H2.G.4 ALPYRWPQWPWSR	>starPep_05293.It4.7.10.11.12.L5.H3.G.5 YWRGF
>starPep_43956.CPP3.H1.G.4 KWRYPLRPL	>starPep_25472.It4.5L.4.1.12.CPP2.H2.G.5 ALPYRWQWPWSR	>starPep_07641.L5.1.H5.G.1 ICIEL
>starPep_43956.CPP3.H1.G.5 KWRYPLPLP	>starPep_15346.It4.6.2.5.7.H4.G5 PSRWM	>starPep_07641.L5.1.H5.G.2 ICQEE
>starPep_16808.L10.H5.G.1 FNGRCGKLP	>starPep_15346.It4.6.2.5.7.H4.G.1 ALPSRWM	>starPep_07641.L5.1.H5.G.3 ICQER
>starPep_16808.L10.H5.G.2 FPGRCGKLA	>starPep_15346.It4.6.2.5.7.H4.G.2 AFPSRWM	>starPep_07641.L5.1.H5.G.4 ECKL
>starPep_17042.It4.2.5.7.4.L5.1.H4.G.1 ECGVG	>starPep_15346.It4.6.2.5.7.H4.G.3 AMDSRWM	>starPep_10014.L5.3.H4.G.1 TAWSI
>starPep_17042.It4.2.5.7.4.L5.1.H4.G.2 ECGFG	>starPep_15346.It4.6.2.5.7.H4.G5.1 AHPSW	>starPep_10014.L5.3.H4.G.2 TSWRI
>starPep_17042.It4.2.5.7.4.L5.1.H4.G.3 QCGFL	>starPep_15346.It4.6.2.5.7.H4.G5.2 HPSWR	>starPep_10014.L5.3.H4.G.3 TAWRY
>starPep_17042.It4.2.5.7.4.L5.1.H4.G.4 ECGFW	>starPep_15346.It4.6.2.5.7.H4.G5.3 PSWRM	>starPep_10014.L5.3.H4.G.4 TSWSI
>starPep_17042.It4.2.5.7.4.L5.1.CPP3.H1.G.1 WLPGS	>starPep_15346.It4.6.2.5.7.H4.G1.1 ALPSWRM	>starPep_12257.It4.4L.6.1.7.H5.G.1 WRQLPWFG

N. (cont.) FASTA of 250 sequences stable in gastrointestinal tract.

>starPep.12257.It4.4L.6.1.7.H5.G.3 WRQLPAFY	>starPep.12276.H1.G15.1 TSWPTCPGYRWMCL	>starPep.12276_CPP1.H5.G5 CPRYR
>starPep.12257.It4.4L.6.1.7.H5.G.4 WRGLPWFY	>starPep.12276.H1.G15.2 HSPTSWPPTCPGYRW	>starPep.12276_CPP1.H5.G.1 KTRNHSPTSQPPTCPRYRWMCLRRRC
>starPep.12257.It4.4L.6.1.7.H5.G.5 WRLLPGFY	>starPep.12276.H1.G15.3 SWPPTCPGYRWMCLR	>starPep.12276_CPP1.H5.G.2 KTRNHSPTSQPPTCPRYRWMCLRCR
>starPep.43502.HL5.3.G.1 AMWWI	>starPep.12276.H1.G15.4 WPPTCPGYRWMCLRR	>starPep.12276_CPP1.H5.G.3 KTRNHSPTSQPPTCPRYRWMCLCRR
>starPep.43502.HL5.3.G.2 AMTWI	>starPep.12276.H1.G15.5 PPTCPGYRWMCLRRF	>starPep.12276_CPP1.H5.G.4 KTRNHSPTSQPPTCPRYCWMCLRRR
>starPep.43502.HL5.3.G.3 AMRWI	>starPep.12276.H2.G5 WPGYR	>starPep.07237_CPP2.H1.G5.1 QHWSY
>starPep.43502.HL5.3.G.5 AFKWW	>starPep.12276.H2.G10.1 TWPGYRWMCL	>starPep.07237_CPP2.H1.G5.2 HWSYR
>starPep.43502.HL5.3.G.7 AFGWY	>starPep.12276.H2.G10.2 PTWPGYRWM	>starPep.07237_CPP2.H1.G5.3 WSYRL
>starPep.43502.H3.G.3 WWAMKSIRY	>starPep.12276.H2.G10.3 PPTWPGYRWM	>starPep.07237_CPP2.H1.G5.4 YRLRP
>starPep.43502.H3.G.4 WWAMTSIRV	>starPep.12276.H2.G10.4 CPPTWPGYRW	>starPep.07237_CPP2.H1.G.2 QHWSYRLPPK
>starPep.43502.H3.G.5 WWAMKSIWV	>starPep.12276.H3.G15.3 CPPTCPGYRWMWLR	>starPep.07237_CPP2.H1.G.3 QHWSYRLRPG
>starPep.43502.H3.G.9 WWAMTSIES	>starPep.12276.H4.G.1 WTSNHSPTSCPPTCPGGRWMCLRRF	>starPep.07237_CPP2.H1.G.4 QHWSYRLRPE
>starPep.43502.H3.G.10 WWAMTSIEA	>starPep.12276.H4.G.2 WTSNHSPTSCPPGCPGYRWMCLRGF	>starPep.07237_CPP2.H1.G.5 QHWSYRLLPG
>starPep.13108_CPP1.HL10.G5 MKPSP	>starPep.12276.H4.G.3 WTSNHSPTSCPPGCPGGRWMCLRGF	>starPep.07237_CPP2.H3.G.1 QHWSYKLLPR
>starPep.13108_CPP1.HL10.G.1 LPWRMKPSPR	>starPep.12276_CPP1.H2.G5.1 RYRWM	>starPep.07237_CPP2.H3.G.2 QHWSYKLPFR
>starPep.13108_CPP1.HL10.G.2 VPWRMKPLPR	>starPep.12276_CPP1.H2.G5.3 YRWM	>starPep.07237_CPP2.H4.G.1 QHWSYRLLPT
>starPep.13108_CPP1.HL10.G.3 VPWRMKPSPN	>starPep.12276_CPP1.H2.G5.4 PRYRW	>starPep.07237_CPP2.H4.G.2 QHWSYRLPPT
>starPep.13108_CPP1.HL10.G.4 VPWRMLPSIR	>starPep.12276_CPP1.H2.G10.1 TRNHSPTSCQ	>starPep.08820.H5.G.1 CPSHLDSGC
>starPep.13108_CPP1.H1.G5.1 WRMKP	>starPep.12276_CPP1.H2.G10.2 RNHSPTSCQP	>starPep.08820.H5.G.2 CPSHLPAGC
>starPep.13108_CPP1.H1.G5.2 KPSPR	>starPep.12276_CPP1.H2.G10.3 NHSPTSCQPT	>starPep.08820.H5.G.3 CPSHLDPGC
>starPep.13108_CPP1.H1.G5.3 PSPRQ	>starPep.12276_CPP1.H2.G10.4 HSPTSCQPTC	>starPep.29033.L15_CPP1.HL5.5.G KCGHG
>starPep.13108_CPP1.H1.G10.1 SVLWRMKPSP	>starPep.12276_CPP1.H2.G15.1 TSCQPTCPRYRWMCL	>starPep.29033.L15_CPP1.HL5.5.G.2 KCGFG
>starPep.13108_CPP1.H1.G10.2 VLWRMKPSPR	>starPep.12276_CPP1.H2.G15.2 PTSCQPTCPRYRWM	>starPep.29033.L15_CPP1.HL5.5.G.3 KCGPG
>starPep.13108_CPP1.H1.G10.3 LWRMKPSPRQ	>starPep.12276_CPP1.H2.G15.3 NHSPTSCQPTCPRYR	>starPep.29033.L15_CPP1.HL5.5.G.4 KCGDG
>starPep.13108_CPP1.H1.G.1 SILWRMKPSPRQ	>starPep.12276_CPP1.H2.G15.4 HSPTSCQPTCPRYRW	>starPep.29033.L15_CPP1.HL5.5.G.5 KCGTG
>starPep.13108_CPP1.H1.G.2 SVLWRMKPLPRQ	>starPep.12276_CPP1.H2.G15.5 QPTCPRYRWMCLRRR	>starPep.29033.L15_CPP1.HL5.6.G HGSGH
>starPep.13108_CPP1.H1.G.3 SILWRMKPLPRQ	>starPep.12276_CPP1.H2.G15.6 SCQPTCPRYRWMCLR	>starPep.29033.L15_CPP1.HL10.2.G.1 KNKHKKPGHG
>starPep.13108_CPP1.H1.G.4 SILWRMKLLPRQ	>starPep.12276_CPP1.H2.G.1 KTRGHSPSTSCQPTCPRYRWMCLRRG	>starPep.29033.L15_CPP1.HL10.2.G.2 KNKHKKHGPG
>starPep.13108_CPP1.H4.G5 RMKPS	>starPep.12276_CPP1.H2.G.2 KTRNHSPTSCQPTCPRGRWMCLRRG	>starPep.29033.L15_CPP1.HL10.3.G.1 NKHKKPGHGH
>starPep.13108_CPP1.H4.G10.1 TWRMKPSPRQ	>starPep.12276_CPP1.H2.G.3 KTRNHSPTSCQPTCPRYRWMCLRCG	>starPep.29033.L15_CPP1.HL10.3.G.2 NKHKKHGPGH
>starPep.13108_CPP1.H4.G10.2 VTWRMKPSPR	>starPep.12276_CPP1.H2.G.4 KTRNHSPTSCQPTCPRYRWMCLCRG	>starPep.29033.L15_CPP1.HL10.4.G.1 KHKKPGHGK
>starPep.13108_CPP1.H4.G10.3 SVTWRMKPSP	>starPep.12276_CPP1.H3.G5 SPTSC	>starPep.29033.L15_CPP1.HL10.4.G.2 KHKKHGPGHK
>starPep.13108_CPP1.H4.G.1 SITWRMKPSPRQ	>starPep.12276_CPP1.H3.G10.1 RNHSPTSCPQ	>starPep.29033.L15_CPP1.H1.G.1 HPGKNKHKKHGHGK
>starPep.13108_CPP1.H4.G.2 SVTWRMKPSPNQ	>starPep.12276_CPP1.H3.G10.2 NHSPTSCPQT	>starPep.29033.L15_CPP1.H1.G5 GKNKHKKHGH
>starPep.13108_CPP1.H4.G.3 SGTWRMKPSPRQ	>starPep.12276_CPP1.H3.G10.3 HSPTSCPQTC	>starPep.29033.L15_CPP1.H2.G.1 KPGKNKHKKHGHGK
>starPep.13108_CPP1.H4.G.4 SITWRMKPLPRQ	>starPep.12276_CPP1.H3.G10.4 TRNHSPTSCP	>starPep.29033.L15_CPP1.H2.G.2 KHGKNKHKKHGHGK
>starPep.12276.H1.G5.1 PGYRW	>starPep.12276_CPP1.H3.G.2 KTRNHSPTSCPQTCPRYRWMCLRRG	>starPep.29033.L15_CPP1.H2.G.3 KHGKNKHKKHGHGK
>starPep.12276.H1.G5.2 CPGYR	>starPep.12276_CPP1.H3.G.4 KTGNHSPTSCPQTCPRYRWMCLRRR	>starPep.14535.It4.3.14W.2.4.CPP1.HL5.1.G.1 SPLPW
>starPep.12276.H1.G5.4 PTSNH	>starPep.12276_CPP1.H3.G.5 KTRNHSPTSCPQTCPRYRWMCLRRRC	>starPep.14535.It4.3.14W.2.4.CPP1.HL5.2.G PLSWA
>starPep.12276.H1.G5.5 HSPTS	>starPep.12276_CPP1.H3.G.6 KTRNHSPTSCPQTCPRYRWMCLCRR	>starPep.25472.It4.5L.4.1.12.CPP2.HL5.G.1 PYRLP
>starPep.12276.H1.G5.6 TSNHS	>starPep.12276_CPP1.H3.G.7 KTRNHSPTSCPQTCPRYRWMCLRRR	
>starPep.12276.H1.G10.4 TSNHSPTSWP	>starPep.25472.It4.5L.4.1.12.CPP2.HL5.G.2 PYWLP	

O. (cont.) Predicted activities of SET 4, conformed by 78 lead THPs with optimized gastrointestinal stability. 13 sequences from SET 4 are highlighted.

Sequence	PflifePred		TumorHPD		THP-pep		AntiGCP		ToxinPred				CoHPD		HemoPred		AMPInu		AigePred2		AmpPP
	Half-time (seconds)	Score	SVM Score	THP	SVM Score	THP	SVM Score 1	SVM Score 2	SVM Score 1	SVM Score 2	SVM Score 3	SVM Score 4	SVM Score 1	SVM Score 2	SVM Score 1	SVM Score 2	Anticancer	Score	Score	Allergen	
CSKWC	834.21	3.01	0.97	THP	0.97	THP	0.51	Anticp	-0.51	Non-Toxin	-0.2	Non-Toxin	-0.2	Non-Toxin	-0.26	Non-CPP	0.5103	0.411	0.411	Allergen	
AMWVC	834.81	2.79	1.12	THP	1.12	THP	-0.79	Anticp	-0.79	Non-Toxin	-0.59	Non-Toxin	-0.59	Non-Toxin	-0.26	Non-CPP	0.7749	0.412	0.412	Allergen	
CRGCF	818.81	2.92	0.88	THP	0.88	THP	-0.71	Anticp	-0.71	Non-Toxin	-0.26	Non-Toxin	-0.26	Non-Toxin	-0.26	Non-CPP	0.4826	0.405	0.405	Allergen	
CGFVA	826.51	1	1	THP	1	THP	-0.69	Anticp	-0.69	Non-Toxin	-0.42	Non-Toxin	-0.42	Non-Toxin	-0.21	Non-CPP	0.4002	0.304	0.304	Allergen	
CGOAS	810.21	1.18	0.82	THP	0.82	THP	-0.12	Non-Anticp	-0.12	Non-Toxin	-0.56	Non-Toxin	-0.56	Non-Toxin	-0.26	Non-CPP	0.4668	0.388	0.388	Allergen	
TAWSI	846.11	1.42	0.78	THP	0.78	THP	-0.91	Anticp	-0.91	Non-Toxin	-0.77	Non-Toxin	-0.77	Non-Toxin	-0.26	Non-CPP	0.5965	0.314	0.314	Allergen	
SPFSC	834.41	1.38	0.85	THP	0.85	THP	-0.88	Anticp	-0.88	Non-Toxin	-0.47	Non-Toxin	-0.47	Non-Toxin	-0.25	Non-CPP	0.3298	0.365	0.365	Allergen	
HSGGH	834.81	1.36	0.78	THP	0.78	THP	-0.78	Anticp	-0.78	Non-Toxin	-0.49	Non-Toxin	-0.49	Non-Toxin	-0.28	Non-CPP	0.4809	0.364	0.364	Allergen	
LCGVG	834.81	1.75	0.87	THP	0.87	THP	-0.89	Anticp	-0.89	Non-Toxin	-0.55	Non-Toxin	-0.55	Non-Toxin	-0.28	Non-CPP	0.7435	0.384	0.384	Allergen	
GCRGS	814.31	2.41	0.97	THP	0.97	THP	-0.66	Non-Anticp	-0.66	Non-Toxin	-0.28	Non-Toxin	-0.28	Non-Toxin	-0.25	Non-CPP	0.2971	0.372	0.372	Allergen	
GPCKL	834.41	2.65	0.95	THP	0.95	THP	-0.63	Anticp	-0.63	Non-Toxin	-0.13	Non-Toxin	-0.13	Non-Toxin	-0.24	Non-CPP	0.4697	0.434	0.434	Allergen	
ICIEL	834.71	1.3	0.89	THP	0.89	THP	-0.71	Anticp	-0.71	Non-Toxin	-0.37	Non-Toxin	-0.37	Non-Toxin	-0.27	Non-CPP	0.5471	0.326	0.326	Allergen	
KTRNSPTSCPMACPRHRWNCWHHC	830.01	2.42	0.8	THP	0.8	THP	-0.14	Anticp	-0.14	Non-Toxin	-0.61	Non-Toxin	-0.61	Non-Toxin	-0.12	Non-CPP	0.0881	0.254	0.254	Non-Allergen	
WWVCHSPTSQPPCPDRDNRNRCRR	933.11	2	0.73	THP	0.73	THP	-0.52	Anticp	-0.52	Non-Toxin	-0.78	Non-Toxin	-0.78	Non-Toxin	-0.23	Non-CPP	0.103	0.262	0.262	Non-Allergen	
WARNSHPTSCNTPQPRCWHSNRNG	849.91	1.89	0.76	THP	0.76	THP	-0.51	Anticp	-0.51	Non-Toxin	-0.66	Non-Toxin	-0.66	Non-Toxin	-0.04	Non-CPP	0.0681	0.304	0.304	Allergen	
LTRNSHSTSLPNTPRCSHWKQAWC	1439.91	1.91	0.75	THP	0.75	THP	-0.28	Anticp	-0.28	Non-Toxin	-0.66	Non-Toxin	-0.66	Non-Toxin	-0.03	Non-CPP	0.0367	0.265	0.265	Non-Allergen	
SPVWRKPSLRQ	1287.71	0.78	0.64	THP	0.64	THP	-1	Non-Anticp	-1	Non-Toxin	-1.48	Non-Toxin	-1.48	Non-Toxin	0.29	Non-CPP	0.0752	0.257	0.257	Non-Allergen	
SPVWRKPSRQ	1212.51	0.46	0.73	THP	0.73	THP	-1.55	Non-Anticp	-1.55	Non-Toxin	-2.05	Non-Toxin	-2.05	Non-Toxin	0.31	Non-CPP	0.1558	0.257	0.257	Non-Allergen	
SPVWRKPSRQ	1131.91	1.13	0.77	THP	0.77	THP	-1.18	Non-Anticp	-1.18	Non-Toxin	-1.48	Non-Toxin	-1.48	Non-Toxin	0.05	Non-CPP	0.1519	0.263	0.263	Non-Allergen	
SPVWRKPSRQ	1131.91	1.13	0.77	THP	0.77	THP	-1.18	Non-Anticp	-1.18	Non-Toxin	-1.48	Non-Toxin	-1.48	Non-Toxin	0.05	Non-CPP	0.1519	0.263	0.263	Non-Allergen	
SPVWRKPSRQ	1131.91	1.13	0.77	THP	0.77	THP	-1.18	Non-Anticp	-1.18	Non-Toxin	-1.48	Non-Toxin	-1.48	Non-Toxin	0.05	Non-CPP	0.1519	0.263	0.263	Non-Allergen	
LPVWRKPSRQ	905.91	1.92	0.84	THP	0.84	THP	-1.31	Anticp	-1.31	Non-Toxin	-0.67	Non-Toxin	-0.67	Non-Toxin	0.23	Non-CPP	0.3411	0.283	0.283	Non-Allergen	
QHWVYKLPDR	1154.31	1.92	0.68	THP	0.68	THP	-0.67	Anticp	-0.67	Non-Toxin	-0.96	Non-Toxin	-0.96	Non-Toxin	0.05	Non-CPP	0.1056	0.333	0.333	Allergen	
SLPVWRKPSLRQ	1174.41	0.96	0.67	THP	0.67	THP	-1.25	Non-Anticp	-1.25	Non-Toxin	-1.61	Non-Toxin	-1.61	Non-Toxin	0.16	Non-CPP	0.075	0.291	0.291	Non-Allergen	
QHWVYKLRPE	1230.41	1.43	0.79	THP	0.79	THP	-1.15	Anticp	-1.15	Non-Toxin	-0.85	Non-Toxin	-0.85	Non-Toxin	0	Non-CPP	0.2905	0.346	0.346	Allergen	
WTNSHPTSCNCPRYRWACRRNF	865.31	1.77	0.79	THP	0.79	THP	-0.58	Anticp	-0.58	Non-Toxin	-1.15	Non-Toxin	-1.15	Non-Toxin	-0.09	Non-CPP	0.0488	0.255	0.255	Non-Allergen	
SGTVWRKPSRQ	1152.81	0.62	0.73	THP	0.73	THP	-1.3	Non-Anticp	-1.3	Non-Toxin	-1.52	Non-Toxin	-1.52	Non-Toxin	0.24	Non-CPP	0.2584	0.25	0.25	Non-Allergen	
HSPTFSWPPFCGYRW	858.91	1.89	0.89	THP	0.89	THP	-0.51	Anticp	-0.51	Non-Toxin	-0.4	Non-Toxin	-0.4	Non-Toxin	-0.05	Non-CPP	0.2825	0.315	0.315	Allergen	
VIVWRKPSR	987.31	0.89	0.72	THP	0.72	THP	-1.35	Anticp	-1.35	Non-Toxin	-1.59	Non-Toxin	-1.59	Non-Toxin	0.25	Non-CPP	0.4058	0.269	0.269	Non-Allergen	
WASNHAMGSCACRAMPKGRKHMRECR	785.41	1.64	0.73	THP	0.73	THP	-0.12	Anticp	-0.12	Non-Toxin	-0.48	Non-Toxin	-0.48	Non-Toxin	-0.2	Non-CPP	0.05	0.264	0.264	Non-Allergen	
HHGTNRKHCNHSHC	834.61	1.9	0.77	THP	0.77	THP	-0.02	Anticp	-0.02	Non-Toxin	-0.22	Non-Toxin	-0.22	Non-Toxin	-0.06	Non-CPP	0.0498	0.409	0.409	Allergen	
WSAMKWHV	946.61	1.11	0.79	THP	0.79	THP	-0.69	Anticp	-0.69	Non-Toxin	-1.03	Non-Toxin	-1.03	Non-Toxin	-0.12	Non-CPP	0.3768	0.285	0.285	Allergen	
WSAMKWHV	915.51	1.19	0.79	THP	0.79	THP	-0.51	Anticp	-0.51	Non-Toxin	-0.73	Non-Toxin	-0.73	Non-Toxin	-0.06	Non-CPP	0.386	0.337	0.337	Allergen	
WWAMKSHV	844.41	1.95	0.67	THP	0.67	THP	-0.93	Anticp	-0.93	Non-Toxin	-1.14	Non-Toxin	-1.14	Non-Toxin	-0.12	Non-CPP	0.4153	0.282	0.282	Non-Allergen	
WSAMPVHV	960.11	2.53	0.83	THP	0.83	THP	-0.43	Anticp	-0.43	Non-Toxin	-0.63	Non-Toxin	-0.63	Non-Toxin	-0.15	Non-CPP	0.1963	0.258	0.258	Non-Allergen	
WVAMKSHV	1006.61	1.46	0.79	THP	0.79	THP	-0.77	Anticp	-0.77	Non-Toxin	-0.86	Non-Toxin	-0.86	Non-Toxin	-0.05	Non-CPP	0.4149	0.25	0.25	Non-Allergen	
AHRVY	844.81	1.74	0.94	THP	0.94	THP	-0.84	Anticp	-0.84	Non-Toxin	-0.8	Non-Toxin	-0.8	Non-Toxin	-0.25	Non-CPP	0.4291	0.233	0.233	Non-Allergen	
AFGVY	823.41	1.56	0.69	THP	0.69	THP	-0.82	Anticp	-0.82	Non-Toxin	-0.33	Non-Toxin	-0.33	Non-Toxin	-0.27	Non-CPP	0.3106	0.465	0.465	Allergen	
YGRCCGGLHP	828.81	2.09	0.98	THP	0.98	THP	-0.72	Anticp	-0.72	Non-Toxin	-0.55	Non-Toxin	-0.55	Non-Toxin	-0.19	Non-CPP	0.237	0.368	0.368	Allergen	
RYR	844.11	1.12	0.84	THP	0.84	THP	-1.01	Anticp	-1.01	Non-Toxin	-0.49	Non-Toxin	-0.49	Non-Toxin	-0.12	Non-CPP	0.174	0.352	0.352	Allergen	
QRLBI	846.91	1.49	0.6	THP	0.6	THP	-1.06	Anticp	-1.06	Non-Toxin	-0.71	Non-Toxin	-0.71	Non-Toxin	-0.22	Non-CPP	0.4062	0.346	0.346	Allergen	

P. Physicochemical properties of SET 8, conformed by 27 lead THPs.

ID	Sequence	Length	Hydrophobicity	Steric hindrance	Sidebulk	Hydrophobicity	Amphipathicity	Hydrophilicity	Net Hydrogen	Charge	pI	Mol wt
THP_YMG1	LPWCLRRLRI	9	-0.09	0.51	0.51	0.69	0.49	-0.56	0.9	2	10.38	1282.81
THP_YMG2	NGRCWKG	7	-0.35	0.58	0.58	-1.39	0.77	0.23	1.12	2	9.55	877.1
THP_YMG3	WRPWSHL	8	-0.14	0.38	0.38	-1.08	0.43	-0.64	0.89	1.5	10.11	1191.53
THP_YMG4	WSYWRQLPWFG	11	-0.03	0.53	0.53	-0.68	0.31	-1.11	0.92	1	9.1	1582.96
THP_YMG5	WRPLSWAP	8	-0.07	0.44	0.44	-0.52	0.27	-0.64	0.78	1	10.11	1109.41
THP_YMG6	PLSWPRWA	8	-0.07	0.44	0.44	-0.52	0.27	-0.64	0.78	1	10.11	1083.37
THP_YMG7	PRWPLSWA	8	-0.07	0.44	0.44	-0.52	0.27	-0.64	0.78	1	10.11	1083.37
THP_YMG8	HHGTTRWC	8	-0.25	0.37	0.37	-1.33	0.59	-0.31	0.89	2	8.61	1096.37
THP_YMG9	WSPYWLPR	8	-0.1	0.46	0.46	-0.87	0.27	-0.84	0.89	1	9.1	1260.58
THP_YMG10	RGDLRWC	7	-0.39	0.56	0.56	-0.94	0.61	0.35	1.25	1	8.6	1008.28
THP_YMG11	CGCGSRSR	10	-0.32	0.57	0.57	-0.13	0.45	0.24	0.91	2	8.67	1187.52
THP_YMG12	LRCWSRC	7	-0.35	0.52	0.52	-0.24	0.61	-0.11	1.25	2	9.1	1026.35
THP_YMG13	CNWWRLRAQFY	11	-0.22	0.57	0.57	-0.68	0.51	-0.71	1.25	2	9.55	1706.12
THP_YMG14	PSPAFKWW	8	0.01	0.46	0.46	-0.57	0.41	-0.72	0.56	1	9.11	1204.51
THP_YMG15	PYWARGWLP	9	-0.02	0.48	0.48	-0.56	0.25	-0.84	0.7	1	9.1	1242.58
THP_YMG16	TARGLCWRY	9	-0.23	0.54	0.54	-0.42	0.49	-0.34	1.1	2	9.55	1288.62
THP_YMG17	TAPYWLPRY	10	-0.05	0.49	0.49	-0.65	0.22	-1.01	0.82	1	8.93	1515.88
THP_YMG18	AMYWRGFWWP	10	0.05	0.54	0.54	-0.36	0.22	-1.25	0.73	1	9.1	1496.9
THP_YMG19	WWWMGCRGS	9	-0.03	0.55	0.55	-0.44	0.25	-0.92	0.8	1	8.6	1255.57
THP_YMG20	CPGCRHSGH	10	-0.21	0.44	0.44	-0.86	0.49	0.03	0.64	2	8.4	1147.41
THP_YMG21	HGSRPWVGH	9	-0.18	0.39	0.39	-1.59	0.54	-0.45	0.9	2	10.11	1256.5
THP_YMG22	WRPWSPTSC	9	-0.18	0.46	0.46	-0.93	0.25	-0.46	0.9	1	8.6	1222.52
THP_YMG23	HHWARGSHC	9	-0.24	0.35	0.35	-1.19	0.68	-0.31	0.9	2.5	8.61	1193.46
THP_YMG24	CPGCRWWWM	9	-0.02	0.52	0.52	-0.23	0.25	-1.05	0.7	1	8.39	1355.81
THP_YMG25	WYYWRGFWM	9	0.08	0.55	0.55	-0.51	0.25	-1.67	0.9	1	9.1	1548.99
THP_YMG26	RQRLPIGR	9	-0.64	0.57	0.57	-1.52	1.1	0.86	1.8	4	12.48	1307.71
THP_YMG27	QHWSYKLPPR	10	-0.34	0.5	0.5	-1.75	0.88	-0.15	1.2	2.5	10.01	1311.65

P. (cont.) Predicted activities of SET 8, conformed by 27 lead THPs.

ID	Sequence	ML Score		AllPred2		IL2Pred		IL-10Pred		Score		IL4Pred		AllPred		PRR		SVM Score	
		Score	Allergen	Score	Allergen	Score	Allergen	Score	Allergen	Score	Allergen	Score	Allergen	Score	Allergen	Score	Allergen	Score	Allergen
THP_YMG1	LPWCLRRLRI	0.381	Allergen	0.806	IL-2 inducer	-0.05212105	IL10 non-inducer	0.28	IL4 inducer	AIP	0.5296	0.44	Non-Pattern Recognition Receptor	Pattern Recognition Receptor	0.54	Pattern Recognition Receptor			
THP_YMG2	NGRCWKKG	0.393	Allergen	0.69	IL-2 inducer	-0.660145052	IL10 non-inducer	0.28	IL4 inducer	AIP	0.5209	0.58	Non-Pattern Recognition Receptor	Pattern Recognition Receptor	0.64	Pattern Recognition Receptor			
THP_YMG3	WRPWPSHL	0.312	Allergen	0.928	IL-2 inducer	-0.273554559	IL10 non-inducer	0.28	IL4 inducer	AIP	0.5163	0.47	Non-Pattern Recognition Receptor	Pattern Recognition Receptor	0.54	Pattern Recognition Receptor			
THP_YMG4	WSYWRQLPWFG	0.378	Allergen	0.716	IL-2 inducer	0.179412966	IL10 non-inducer	0.27	IL4 inducer	AIP	0.407	0.46	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.46	Non-Pattern Recognition Receptor			
THP_YMG5	WRPLSWAP	0.27	Non-Allergen	0.796	IL-2 inducer	-0.379288801	IL10 non-inducer	0.28	IL4 inducer	Non-AIP	0.3902	0.46	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.43	Non-Pattern Recognition Receptor			
THP_YMG6	PLSWPRVA	0.27	Non-Allergen	0.674	IL-2 inducer	-0.451490516	IL10 non-inducer	0.28	IL4 inducer	AIP	0.3651	0.46	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.43	Non-Pattern Recognition Receptor			
THP_YMG7	PRWP4SVA	0.27	Non-Allergen	0.674	IL-2 inducer	-0.451490516	IL10 non-inducer	0.28	IL4 inducer	AIP	0.3651	0.46	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.43	Non-Pattern Recognition Receptor			
THP_YMG8	HGTPRWK	0.283	Non-Allergen	0.714	IL-2 inducer	-0.676549063	IL10 non-inducer	0.28	IL4 inducer	AIP	0.4256	0.48	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.51	Non-Pattern Recognition Receptor			
THP_YMG9	WSPYWLPR	0.305	Allergen	0.858	IL-2 inducer	-0.1398894003	IL10 non-inducer	0.26	IL4 inducer	AIP	0.5186	0.42	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.41	Non-Pattern Recognition Receptor			
THP_YMG10	RGDLRWC	0.363	Allergen	0.838	IL-2 inducer	-0.764152544	IL10 non-inducer	0.28	IL4 inducer	Non-AIP	0.3722	0.52	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.42	Non-Pattern Recognition Receptor			
THP_YMG11	CGGCSRCR	0.373	Allergen	0.894	IL-2 inducer	-1.140550429	IL10 non-inducer	0.28	IL4 inducer	AIP	0.4628	0.48	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.49	Non-Pattern Recognition Receptor			
THP_YMG12	LRCSWRC	0.32	Allergen	0.984	IL-2 inducer	-0.506084432	IL10 non-inducer	0.28	IL4 inducer	AIP	0.5256	0.52	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.5	Non-Pattern Recognition Receptor			
THP_YMG13	CNWRRLAQFY	0.352	Allergen	0.868	IL-2 inducer	-0.050828942	IL10 non-inducer	0.27	IL4 inducer	AIP	0.5186	0.45	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.4	Non-Pattern Recognition Receptor			
THP_YMG14	PSPAFKWV	0.38	Allergen	0.86	IL-2 inducer	-0.294383533	IL10 non-inducer	0.28	IL4 inducer	AIP	0.5209	0.47	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.46	Non-Pattern Recognition Receptor			
THP_YMG15	PYWARGLP	0.285	Non-Allergen	0.75	IL-2 inducer	-0.101615187	IL10 non-inducer	0.28	IL4 inducer	AIP	0.4256	0.46	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.34	Non-Pattern Recognition Receptor			
THP_YMG16	TARGLCAVY	0.357	Allergen	0.402	IL-2 non-inducer	-0.174383382	IL10 non-inducer	0.24	IL4 inducer	AIP	0.5233	0.43	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.33	Non-Pattern Recognition Receptor			
THP_YMG17	TAPYWLPRY	0.29	Non-Allergen	0.724	IL-2 inducer	-0.063659525	IL10 non-inducer	0.27	IL4 inducer	AIP	0.4884	0.44	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.36	Non-Pattern Recognition Receptor			
THP_YMG18	AMVWRGFWWP	0.258	Non-Allergen	0.454	IL-2 non-inducer	0.013507739	IL10 non-inducer	0.27	IL4 inducer	AIP	0.5721	0.42	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.38	Non-Pattern Recognition Receptor			
THP_YMG19	WWWVGGRCG	0.317	Allergen	0.806	IL-2 inducer	-0.610054741	IL10 non-inducer	0.28	IL4 inducer	AIP	0.5628	0.5	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.54	Non-Pattern Recognition Receptor			
THP_YMG20	CF-CRRHSGH	0.349	Allergen	0.85	IL-2 inducer	-0.642309909	IL10 non-inducer	0.27	IL4 inducer	AIP	0.3884	0.47	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.52	Non-Pattern Recognition Receptor			
THP_YMG21	HGSWRPWGH	0.312	Allergen	0.852	IL-2 inducer	-0.629949166	IL10 non-inducer	0.26	IL4 inducer	AIP	0.4535	0.5	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.62	Non-Pattern Recognition Receptor			
THP_YMG22	WRPWSPTSC	0.298	Non-Allergen	0.898	IL-2 inducer	-0.526674545	IL10 non-inducer	0.19	Non IL4 inducer	AIP	0.4884	0.48	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.58	Non-Pattern Recognition Receptor			
THP_YMG23	HWARGSHC	0.277	Non-Allergen	0.832	IL-2 inducer	-0.593692415	IL10 non-inducer	0.28	IL4 inducer	AIP	0.4814	0.53	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.54	Non-Pattern Recognition Receptor			
THP_YMG24	CPGCRWVWM	0.323	Allergen	0.74	IL-2 inducer	-0.80685909	IL10 non-inducer	0.28	IL4 inducer	AIP	0.5419	0.44	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.48	Non-Pattern Recognition Receptor			
THP_YMG25	WVYWRGFWM	0.254	Non-Allergen	0.758	IL-2 inducer	-0.161308748	IL10 non-inducer	0.26	IL4 inducer	AIP	0.6512	0.38	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.42	Non-Pattern Recognition Receptor			
THP_YMG26	RQLPLGRR	0.416	Allergen	0.558	IL-2 inducer	0.429418195	IL10 non-inducer	0.25	IL4 inducer	AIP	0.5256	0.5	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.34	Non-Pattern Recognition Receptor			
THP_YMG27	QHWSYKLPFR	0.333	Allergen	0.862	IL-2 inducer	-0.353241021	IL10 non-inducer	0.1	Non IL4 inducer	AIP	0.4166	0.44	Non-Pattern Recognition Receptor	Non-Pattern Recognition Receptor	0.42	Non-Pattern Recognition Receptor			

P. (cont.) Predicted activities of SET 8, conformed by 27 lead THPs.

ID	Sequence	QSPred		KNN Physico		KNN Hybrid		RF Physico		RF Binary		RF Hybrid		SVM score		dPABBS	
		Comp	Physico	Comp	Physico	Comp	Physico	Comp	Physico	Comp	Physico	Comp	Physico	Comp	Physico	Prediction - dPABBS	WEKA Probability
THP_YMG1	LPWCLRRLRI	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	-0.47	Biofilm-inactive	Biofilm-inactive	0.74
THP_YMG2	NGRCWKKG	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	-0.33	Biofilm-inactive	Biofilm-inactive	0.71
THP_YMG3	WRPWPSHL	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	0.02	Biofilm-inactive	Biofilm-inactive	0.29
THP_YMG4	WSYWRQLPWFG	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	-0.21	Biofilm-inactive	Biofilm-inactive	0.46
THP_YMG5	WRPLSWAP	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	-0.01	Biofilm-inactive	Biofilm-inactive	0.33
THP_YMG6	PLSWPRVA	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	Non-QSP	QSP	QSP	-0.01	Biofilm-inactive	Biofilm-inactive	0.33
THP_YMG7	PRWP4SVA	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	Non-QSP	QSP	QSP	-0.01	Biofilm-inactive	Biofilm-inactive	0.33
THP_YMG8	HGTPRWK	Non-QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	0.17	Biofilm-inactive	Biofilm-inactive	0.55
THP_YMG9	WSPYWLPR	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	-0.26	Biofilm-inactive	Biofilm-inactive	0.23
THP_YMG10	RGDLRWC	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	-0.48	Biofilm-inactive	Biofilm-inactive	0.5
THP_YMG11	CGGCSRCR	Non-QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	Non-QSP	Non-QSP	Non-QSP	Non-QSP	Non-QSP	Non-QSP	-0.74	Biofilm-inactive	Biofilm-inactive	0.06
THP_YMG12	LRCSWRC	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	-0.27	Biofilm-inactive	Biofilm-inactive	0.49
THP_YMG13	CNWRRLAQFY	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	0.43	Biofilm-inactive	Biofilm-inactive	0.67
THP_YMG14	PSPAFKWV	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	Non-QSP	QSP	QSP	0	Biofilm-inactive	Biofilm-inactive	0.38
THP_YMG15	PYWARGLP	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	Non-QSP	QSP	QSP	-0.03	Biofilm-inactive	Biofilm-inactive	0.54
THP_YMG16	TARGLCAVY	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	Non-QSP	QSP	QSP	0.39	Biofilm-inactive	Biofilm-inactive	0.71
THP_YMG17	TAPYWLPRY	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	0.44	Biofilm-inactive	Biofilm-inactive	0.66
THP_YMG18	AMVWRGFWWP	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	0.47	Biofilm-inactive	Biofilm-inactive	0.52
THP_YMG19	WWWVGGRCG	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	-0.48	Biofilm-inactive	Biofilm-inactive	0.2
THP_YMG20	CF-CRRHSGH	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	-0.6	Biofilm-inactive	Biofilm-inactive	0.22
THP_YMG21	HGSWRPWGH	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	-0.13	Biofilm-inactive	Biofilm-inactive	0.31
THP_YMG22	WRPWSPTSC	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	Non-QSP	QSP	QSP	-0.71	Biofilm-inactive	Biofilm-inactive	0.19
THP_YMG23	HWARGSHC	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	0.43	Biofilm-inactive	Biofilm-inactive	0.49
THP_YMG24	CPGCRWVWM	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	-0.16	Biofilm-inactive	Biofilm-inactive	0.5
THP_YMG25	WVYWRGFWM	QSP	QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	0.16	Biofilm-inactive	Biofilm-inactive	0.54
THP_YMG26	RQLPLGRR	QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	0.67	Biofilm-inactive	Biofilm-inactive	0.26
THP_YMG27	QHWSYKLPFR	Non-QSP	Non-QSP	QSP	Non-QSP	QSP	Non-QSP	QSP	QSP	QSP	QSP	QSP	QSP	0.38	Biofilm-inactive	Biofilm-inactive	-0.2

P. (cont.) Predicted activities of SET 8, conformed by 27 lead THPs.

ID	Sequence	AMPDiscover				ClassAMP				CAMP+3				AMPClassifier			
		RF	ANN	AVP	APP	ABP	AFP	APP	AVP	Antibacterial	ANN	DA	RF	SVM	kNN	SVM	RF
THP_YMG1	LPWCLRRLRI	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	AMP	NAMP	AMP	AMP	AMP	AMP
THP_YMG2	NGRCWKG	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	AMP	AMP	AMP	AMP	AMP
THP_YMG3	WRPWPSHL	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG4	WSYWRQLPAWFG	AMP	ABP	AVP	nonAPP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	non-AMP	AMP
THP_YMG5	WRPLSWAP	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG6	PLSWPRWA	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG7	PRWPLSWA	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG8	HHGTPRWC	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG9	WSPYWLPR	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG10	RGDLRWC	AMP	nonABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG11	CGCGSCRSR	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG12	LRCWSRC	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG13	CNWWRLRAQFY	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG14	PSPAFKWW	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG15	PYWARGWLP	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG16	TARGLCWRY	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG17	TAPYWLPRY	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG18	AMYWRGFWWP	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG19	WWVWMCGRGS	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG20	CPGCRHGSCH	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG21	HGSRPWGSH	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG22	WRPWSPTSC	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG23	HHWARGSHC	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG24	CPGCRWWVM	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG25	WVYWRGFWM	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG26	RQRLPIGRR	AMP	ABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP
THP_YMG27	QHWYSYKLPFR	AMP	nonABP	AVP	APP	AMP	AFP	APP	AVP	AMP	AMP	NAMP	NAMP	non-AMP	AMP	AMP	AMP

P. (cont.) Predicted activities of SET 8, conformed by 27 lead THPs.

ID	Sequence	AMPfun				Meta-IAVP				iAMPred				AxPEP				AntiP				AntiThpPred			
		AMP	Antiparasitic	Antiviral	Anticancer	Targeting mammals	Anti-fungal	Targeting Gram (-)	Targeting Gram (+)	Score antibacterial	Score antiviral	Score antifungal	Score antimicrobial	Score	Prediction	Score	Score	Prediction	Score	Score	Prediction	Score	Score		
THP.YMG1	LPWCLRLRI	0.9844	0.5182	0.8667	0.4245	0.3333	0.5375	0.9167	0.8363	0.0555	0.628	0.071	0.71	-0.8118754	Non-Antifungal	0.93772771	0.7280815								
THP.YMG2	NGRCVKG	0.9862	0.2727	0.6	0.3636	0.0667	0.6723	0.775	0.6915	0.813	0.658	0.505	0.32	-0.23979716	Antifungal	-0.23979716	1.1932261								
THP.YMG3	WRPVPBHL	0.9763	0.1545	0.925	0.4404	0.0667	0.5453	0.6583	0.6642	0.783	0.406	0.527	0.74	-0.6262508	Non-Antifungal	-0.44607806	0.76633401								
THP.YMG4	WSYWRQLPWFG	0.996	0.1091	0.6333	0.6447	0.1	0.4749	0.6539	0.6539	0.828	0.388	0.394	0.63	-0.2356232	Non-Antifungal	0.78007734	2.10333341								
THP.YMG5	WRPLSWAP	0.9938	0.0545	0.7833	0.5128	0.0667	0.5525	0.6083	0.7273	0.783	0.229	0.361	0.85	-0.61259389	Non-Antifungal	0.22572689	0.79420516								
THP.YMG6	PLSWPRWA	0.99	0.0455	0.7167	0.4194	0.0667	0.3872	0.775	0.6996	0.777	0.224	0.354	0.51	-0.20982009	Non-Antifungal	0.22719556	0.80155775								
THP.YMG7	PRWPLSWA	0.995	0.1273	0.7167	0.385	0.0667	0.5236	0.6917	0.7847	0.777	0.223	0.354	0.63	-0.33683856	Non-Antifungal	0.21691015	0.79944312								
THP.YMG8	HHGTPRAW	0.9659	0.2273	0.6333	0.5314	0.1	0.7226	0.5583	0.5811	0.758	0.686	0.51	0.45	0.25321613	Non-Antifungal	-0.067360765	0.70995073								
THP.YMG9	WSPYWLPR	0.9825	0.1636	0.775	0.378	0.1	0.4922	0.6917	0.6286	0.78	0.33	0.578	0.76	-0.05488184	Non-Antifungal	-0.44186145	1.0573971								
THP.YMG10	RGDLRW	0.9638	0.3455	0.775	0.3859	0.1	0.5933	0.5833	0.5287	0.243	0.539	0.208	0.32	-0.32753846	Non-Antifungal	0.34355081	0.66435081								
THP.YMG11	CGCGSRSR	0.979	0.2	0.5833	0.6576	0.0333	0.6502	0.8	0.7469	0.874	0.898	0.799	0.66	1.1961358	Antifungal	-0.67749451	0.56310059								
THP.YMG12	LRCWSRC	0.99	0.2636	0.7667	0.4995	0.1	0.8337	0.7167	0.6923	0.481	0.772	0.399	0.28	0.24398566	Antifungal	-0.82271693	1.0600832								
THP.YMG13	CNWWLRAQFY	0.9867	0.3727	0.6167	0.4197	0.0667	0.4771	0.6	0.5417	0.647	0.507	0.167	0.5	-0.90660105	Non-Antifungal	1.3840615	2.074092								
THP.YMG14	PSPAPKWW	0.9867	0.2909	0.7583	0.3503	0.1333	0.5026	0.7167	0.5767	0.818	0.61	0.383	0.41	-0.41760293	Non-Antifungal	-0.062949073	0.65767449								
THP.YMG15	PYWARGWLP	0.9817	0.1273	0.7583	0.6639	0.2	0.4754	0.6917	0.5523	0.751	0.379	0.456	0.55	-0.86812146	Non-Antifungal	1.0873826	1.4211057								
THP.YMG16	TARGLCWRY	0.9863	0.1273	0.725	0.444	0.1333	0.676	0.7333	0.6556	0.482	0.566	0.569	0.4	-0.52801402	Non-Antifungal	2.6482984	1.8209758								
THP.YMG17	TAPYWLPAWRY	0.99	0.0091	0.8	0.644	0.2	0.3619	0.7	0.6267	0.727	0.461	0.518	0.48	-0.74428438	Non-Antifungal	0.58912696	1.1989485								
THP.YMG18	AMVVRGFWWP	0.9891	0.2545	0.7833	0.6715	0	0.4247	0.675	0.6045	0.776	0.672	0.389	0.57	-0.3361888	Non-Antifungal	1.0619798	1.5119185								
THP.YMG19	WWWVMGCRGS	0.9716	0.3636	0.575	0.4076	0	0.6874	0.7417	0.6765	0.711	0.638	0.44	0.54	0.33264286	Antifungal	-0.32469069	0.95468052								
THP.YMG20	CPGCRHSGH	0.9832	0.2545	0.6	0.3414	0	0.7226	0.775	0.6803	0.891	0.776	0.786	0.66	0.44008087	Antifungal	-0.26021683	0.33392327								
THP.YMG21	HGSWRPWGH	0.98	0.1182	0.8083	0.3211	0.1333	0.5176	0.7167	0.6482	0.72	0.507	0.488	0.69	0.024978864	Non-Antifungal	-0.1633616	0.82942079								
THP.YMG22	WRPWSPTSC	0.9863	0.1364	0.9	0.3045	0.0333	0.6333	0.5477	0.6333	0.767	0.317	0.457	0.73	0.091045668	Non-Antifungal	-0.67873198	0.60733054								
THP.YMG23	HHWARGSHC	0.97	0.2727	0.6583	0.4811	0.1	0.7814	0.75	0.7936	0.736	0.739	0.467	0.59	-0.14851988	Non-Antifungal	-0.0988906869	0.69554753								
THP.YMG24	CPGCRWWWM	0.9841	0.2182	0.725	0.3506	0	0.6013	0.7917	0.7895	0.705	0.717	0.477	0.47	0.070877086	Antifungal	-0.07097802	0.3764879								
THP.YMG25	WWWYWRGFWM	0.973	0.4	0.7917	0.6427	0.0333	0.6068	0.7667	0.677	0.666	0.609	0.433	0.53	-0.79464479	Non-Antifungal	-0.13798628	0.97381843								
THP.YMG26	RQLRPIGR	0.9907	0.5455	0.7417	0.4788	0.0667	0.7151	0.7417	0.6352	0.894	0.573	0.663	0.65	-0.30205662	Non-Antifungal	0.55902755	1.049842								
THP.YMG27	QHWYSKLPFR	0.9793	0.0909	0.5667	0.1056	0.1333	0.5046	0.65	0.5732	0.324	0.106	0.293	0.46	0.14825637	Antifungal	-1.0168668	1.0873239								

